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To cite this article: JJ Pearson, R Gerber, W Malherbe, NJ Smit & L de Necker (2024) A review of the reported and future potential ecological impacts of the invasive freshwater snail *Tarebia granifera* in South Africa., African Journal of Aquatic Science, 49:3, 179-195, DOI: [10.2989/16085914.2024.2357292](https://doi.org/10.2989/16085914.2024.2357292)

To link to this article: <https://doi.org/10.2989/16085914.2024.2357292>



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Published online: 27 Sep 2024.



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A review of the reported and future potential ecological impacts of the invasive freshwater snail *Tarebia granifera* in South Africa.

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The Southeast Asian gastropod *Tarebia granifera* is an invasive freshwater snail across several continents. Our review summarises the impacts of this invasive snail on invaded aquatic ecosystems. The most important impacts are those related to their ability to reproduce quickly and reach high densities within invaded ecosystems. The snail has reportedly caused declines and local extinctions in native snail populations globally. They can further significantly reduce algal standing stocks and may have severe impacts on benthic biomass, resulting in decreased benthic biodiversity and disturbances in aquatic ecosystem function. These invasive snails also cause bioturbation that can lead to changes in the nutrient cycles of invaded aquatic ecosystems and affect food web dynamics, with unknown implications for ecosystem function. Furthermore, *T. granifera* is well known to harbour a diverse range of parasitic species of medical and veterinary importance both within and outside their native ranges. The effective control of invasive *T. granifera* remains challenging as they are not easily preyed upon, and potential control methods may have unintentional side effects on native biota. This review provides evidence that there is a need to control this invader to avoid further degradation of freshwater habitats and aquatic biodiversity.

Keywords: disease transmission, ecological risk, exotic species, invasion biology, management, native species, quilted melania

Introduction

The loss of aquatic biodiversity attracts far less attention than that of terrestrial environments even though current trends indicate a more rapid loss of native biodiversity in aquatic environments (Cooke et al. 2016). The Living Planet Report of 2020 revealed that species in freshwater environments are at a higher risk of extinction than those in terrestrial environments, with almost a third of freshwater species at risk of extinction (Collen et al. 2014; WWF 2020). Despite the clear loss and great risk of extinction for freshwater biota, research and conservation efforts are still largely lacking (Thomaz et al. 2015; Di Marco et al. 2017).

Whether accidental or deliberate, the introduction and consequent establishment of non-native species remains a leading threat to, and cause of, global biodiversity loss (Meyerson et al. 2019; Reid et al. 2019; WWF 2020). This is particularly true for freshwater aquatic ecosystems, that have a far greater biodiversity per surface area than either marine or terrestrial environments (Dudgeon et al. 2006). Aquatic invasive species (AIS) are one of the most important threats to global freshwater biodiversity and it is thus critical to research and understand the factors leading to the introduction, establishment, spread and impact of AIS in these ecosystems (Havel et al. 2015; Reid et al. 2019; Jones et al. 2021). Despite their ubiquity and diversity, freshwater molluscs are one of the most

threatened groups of animals in the world (Bogan 2007; Johnson et al. 2013; Lopes-Lima et al. 2021) and one of the primary threats to these biota is the introduction of alien species (Almeida et al. 2018).

Alien or non-native species are those introduced into new environments, outside of their native range (Facon et al. 2006; Richardson and Pyšek 2006). Some alien species have the ability to adapt, survive and reproduce in new environments, becoming established and often rapidly increasing their population (Pimentel et al. 2005; Richardson and Pyšek 2006; Prentis et al. 2008). Not all alien species become invasive as some will die off naturally in their introduced environment or survive without causing extensive harm to the environment or native species (Lovell et al. 2005). Invasive species are alien species that have been introduced, become established, and spread while often causing harm to the environment, native species and the economy (Lovell et al. 2005; Pimentel et al. 2005; Richardson and Pyšek 2006).

The introduction of alien species is considered one of southern Africa's most important threats to freshwater biodiversity after habitat modification and pollution (Snoeks et al. 2011; Weyl et al. 2020). South African freshwater biota are particularly vulnerable to the introduction of alien species since many already-threatened species occur in the country. These habitats are further under pressure from the

considerable levels of development taking place (Snoeks et al. 2011) and high rates of international trade between South Africa and other countries, thus increasing the risk of invasive species introduction (Faulkner et al. 2020). Knowledge of the alien species that are present and their distribution in South Africa should be monitored since management and control of invasive species are of national priority (Weyl et al. 2020; Makherana et al. 2022a).

As with global trends, the rate of alien species introduction to South Africa has been directly linked to an increase in international trade (Faulkner et al. 2020). The number of introduced species in South Africa increased in the 1950s along with the improvement of technology that allowed for more rapid international trade and, consequently, more pathways for the introduction of alien species (Faulkner et al. 2020). Aside from international transport and import through cargo and ships (Lovell et al. 2005; Faulkner et al. 2020), the aquarium industry, particularly the aquarium hobby sales and the trade in aquatic plants, is one of the most important methods of freshwater molluscan introductions globally (Walker 1978; Madsen and Frandsen 1989; Cowie 1998; Pointier 1999; Letelier et al. 2007; Strayer 2010; Preston et al. 2022).

Thirteen freshwater molluscan species were introduced into South Africa between 1942 and 2006. Seven of these species have established, and up to 40% of introduced freshwater molluscan species have become invasive (Appleton 2003; Appleton and Miranda 2015a). Almost all freshwater molluscan species introduced into South Africa before 1990 originated from the Americas, apart from one Australian species (*Physastra gibbosa*; Gould 1846) and one East African species (*Biomphalaria angulosa*; Mandahl-Barth 1957) that were unable to establish (Appleton 2003; Appleton and Miranda 2015a). Imports of products from Asia to South Africa doubled from 1998 to 2011 and this increase most certainly led to increased rates of alien species introduction (Appleton and Miranda 2015a; Miranda et al. 2022). Indeed, four Asian molluscan species have been introduced into South Africa since 1999, namely *Tarebia granifera* (Lamarck 1822), *Radix rubiginosa* (Michelin 1831), *Gyraulus chinensis* (Dunker 1848) and *Sinotaia quadrata* (Benson 1842). With increased international trade, the risk of further alien species introductions will also increase, and better control is required to prevent this (Appleton and Miranda 2015a; Miranda et al. 2022).

Tarebia granifera (quilted melania) is a prosobranch gastropod native to Southeast Asia (Figure 1a) (Abbott 1952; Appleton et al. 2009; Appleton and Miranda 2015a). According to Sodeman (1991), *T. granifera* is also native to Madagascar, although Madhyastha and Dutta (2012) consider the presence of *T. granifera* in Madagascar uncertain. The native distribution for *T. granifera* was based on the IUCN Red-listing assessment completed by Madhyastha and Dutta (2012), while additional records of invasion were based on GBIF (2021) data. This mollusc has successfully invaded predominantly tropical and subtropical aquatic ecosystems across several continents (Figures 1b, c), including the Ukraine, Caribbean Islands, Texas (USA), Mexico, Hawaii, Israel, Egypt, Mozambique, Zimbabwe, Eswatini and South Africa (Chaniotis et al. 1980a;

Contreras-Arquieta and Contreras-Balderas 1999; Appleton 2003; Pointier et al. 2003; Appleton et al. 2009; Karatayev et al. 2009; López-López et al. 2009; Miranda et al. 2010; Appleton and Miranda 2015a; López-Altarrriba et al. 2019; Rico-Sánchez et al. 2020; Sullivan and Littrell 2020; Moustafa et al. 2021). This snail was first reported in Africa by Appleton and Nadasan (2002) from the northern parts of the South African province of KwaZulu-Natal in 1999, where it was discovered in a concrete reservoir at Mandeni. *Tarebia granifera* was most likely introduced into the country as a stowaway of the aquarium industry (Picker and Griffiths 2011). Since then, it has spread into numerous water bodies including several subtropical rivers and estuaries in southern Africa and throughout KwaZulu-Natal, and northwards into the Mpumalanga and Limpopo provinces of South Africa (Figure 1c; Appleton et al. 2009; Miranda et al. 2011b; Malherbe 2018). Appleton et al. (2009) collected *T. granifera* in 2006 from 13 rivers, five lakes, two impoundments and several estuaries in northern KwaZulu-Natal. Well-established populations have since been found in important conservation areas including the Kruger National Park, iSimangaliso Wetland Park and Ndumo Game Reserve (Appleton 2003; Wolmarans and de Kock 2006; Dube et al. 2017; Acosta et al. 2020; de Necker et al. 2021; Majdi et al. 2022).

Although reports have been published on the effects of *T. granifera* in invaded regions globally, there has been no comprehensive review or report of the current and potential future impacts of *T. granifera* in southern Africa. The aim of this review is thus to synthesise the reported effects of this invasive snail on invaded ecosystems globally and extrapolate these effects to South African environments and their associated aquatic biota. This is the first synthesis of the data on this ecologically important invader.

Biology and reproduction

Although *T. granifera* has been reported from rivers at an altitude of 983 m on Guam Island, Micronesia (Abbott 1952), it should be noted that temperatures at this altitude were consistently above 24 °C, suggesting that temperature, rather than altitude, plays the most important role in this mollusc's distribution patterns. Most localities invaded by *T. granifera* in South Africa lie below an altitude of 300 m above sea level (asl) (Appleton et al. 2009). Research has, however, found that *T. granifera* can survive extreme temperature ranges of 0–47.5 °C and may even be able to survive cold fronts if the temperature remains above –1 °C (Miranda et al. 2010). This is a much wider temperature range than previously proposed in the literature (7–40 °C) by Chaniotis et al. (1980c). This may make it possible for *T. granifera* to invade areas at elevated altitudes with colder climates, thus expanding their potential invasion range not only in southern Africa but other countries worldwide (Miranda et al. 2010). According to Albarrán-Mélzer et al. (2020), the harder and thicker shells of *T. granifera* not only offer better protection against harsh environmental conditions and predation, but also provide better thermoregulation performance in environments with high-temperature extremes (Figure 2). This may give them an advantage over native species and facilitate their

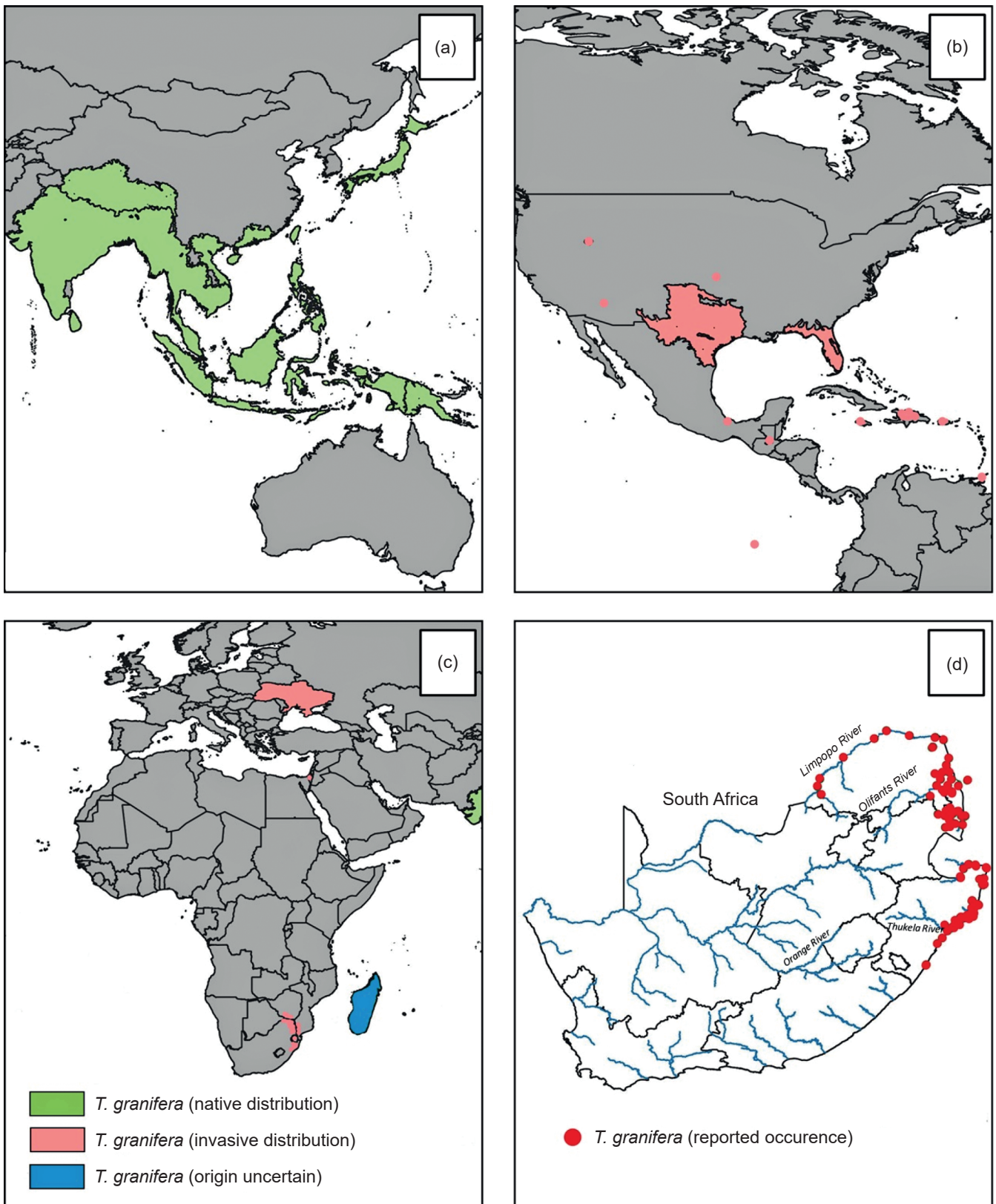


Figure 1: (a) – (c): Maps illustrating the global native and invasive distribution of *Tarebia granifera*. (d) Map of the reported occurrence of *T. granifera* in South Africa (Madhyastha and Dutta 2012; GBIF 2021)

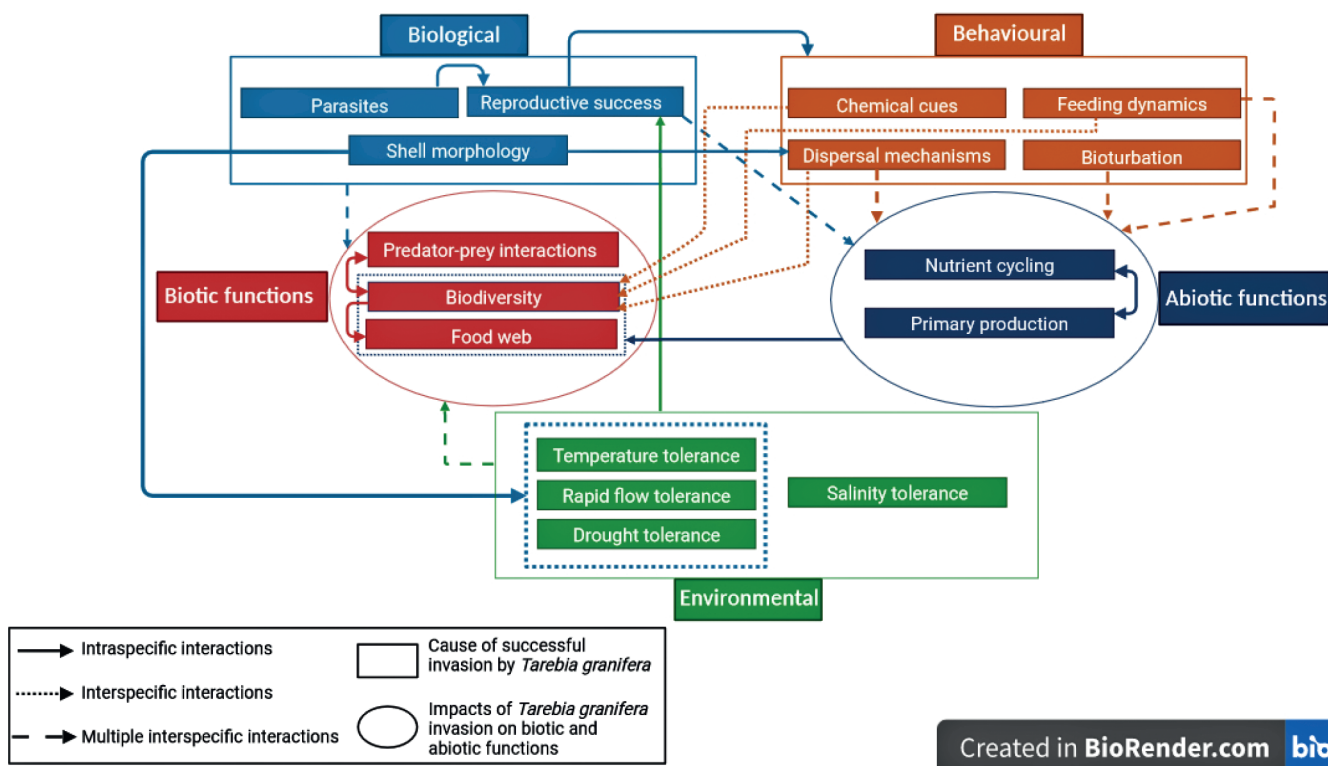


Figure 2: Summative infographic of reported causes of successful invasion by *Tarebia granifera* including biological, behavioural, and environmental causes and reported and proposed impacts of this species on both biotic and abiotic functions of invaded habitats

invasion success, especially since climate change will likely contribute to the spread of *T. granifera* in the future (Albarrán-Mélzer et al. 2020).

While *T. granifera* is considered a freshwater species, experiments have indicated that it can tolerate and survive at much higher levels of salinity than other freshwater species and for several weeks at a time (Figure 2) (Miranda et al. 2010). The snails achieve this by reducing their physical activity, closing their operculum, and burying into sediment until freshwater returns (Chaniotis et al. 1980a; Miranda et al. 2010). Miranda et al. (2010) found that lethal salinity for 50% of the population was reached at 30 PSU (Practical Salinity Unit) between 65 to 75 days and 40 PSU between 15 to 25 days. Indeed, the snail has been reported in highly saline and estuarine environments in South Africa, including the natural saline Lake Nyamithi (Acosta et al. 2020; de Necker et al. 2021) and the St Lucia estuary (Miranda et al. 2010). However, *T. granifera* are reportedly unable to survive acute increases in salinity as was evidenced by a salinity shock experiment by Miranda et al. (2010), as well as a study of the aquatic invertebrates of Lake Nyamithi. These studies determined that although *T. granifera* was able to live in the lake under natural saline conditions, they were unable to survive in the lake once salinity tripled for an extended period during a suprasedonal drought (de Necker et al. 2021).

Moreover, laboratory experiments have reported that this invasive snail can tolerate desiccation (air exposure) for two to three days at temperatures of 25 to 30 °C and 76 to 92%

relative humidity (Chaniotis et al. 1980c; Facon et al. 2004; Miranda et al. 2011b). Furthermore, *T. granifera* can tolerate fast-flowing currents due to their streamlined shape and ability to hold their position well during sudden flow changes (Sodeman 1991). They have been found migrating upstream in streams discharging up to 0.05 m³ s⁻¹ in the Caribbean islands (Prentice 1983) and up to 1.2 m³ s⁻¹ in KwaZulu-Natal (Appleton et al. 2009). Upstream migration was shown to be dependent on snail size, food distribution and habitat variety (Snider and Gilliam 2008).

The ability of *T. granifera* to tolerate wide temperature extremes, high salinity, desiccation and fast-flowing water facilitates its success as an invader and significantly increases the range of aquatic ecosystems it is capable of invading (Wolmarans and de Kock 2006; Appleton et al. 2009; Miranda et al. 2010; Jones et al. 2017). Invaders with high ecological flexibility are likely to reach much higher population densities and inhabit larger regions, which play an essential role in their impact on the environment because of their ability to survive more diverse environmental conditions and outcompete native biota (Nentwig et al. 2010; Kesner and Kumschick 2018; Sirza et al. 2020).

Tarebia granifera are parthenogenetic (capable of reproduction without fertilisation) as well as ovoviviparous (eggs hatch within the parent). These are traits that are also key to its success as an invader (Figure 2) (Appleton et al. 2009; Sirza et al. 2020). Shell length in adult snails ranges from 6 to 40 mm, but the most common size is ~25 mm (Abbott 1952; Chaniotis et al. 1980a). Sexual

maturity is reached at a size between 5.5–8 mm, typically within 122 days of age (Chanotis et al. 1980a; Prentice 1983), although observations made by Appleton et al. (2009) from snails collected in invaded regions in South Africa indicated that snails as small as 8 mm could reproduce, although most juveniles were born to snails >14 mm. New-born snails, 0.7–2.1 mm (Appleton et al. 2009) emerge through a birth opening on the righthand side of the head (Abbott 1952). These juveniles (0.5~1.25 mm) have a survival rate approaching 100% (Chen 2003) and can grow at a rate of about 0.16 mm per month (Chanotis et al. 1980a). *Tarebia granifera* can give birth to an average of one juvenile every 12 hours and would therefore be able to rapidly reach high densities in newly invaded areas (Abbott 1952; Chanotis et al. 1980a; Sodeman 1991; Appleton et al. 2009). Invasive snail species have been reported to have higher probabilities of causing economic and environmental harm due to their ability to reproduce rapidly (Keller et al. 2007; Rumi et al. 2010; Nash and Hoffmann 2012; Kesner and Kumschick 2018). Their ability to adapt to environmental changes, as well as their ability to reproduce quickly and rapidly reach high densities, enable *T. granifera* to often become the dominant invertebrate species in invaded aquatic habitats (Appleton 2003; de Kock and Wolmarans 2007; Appleton et al. 2009; Miranda et al. 2011a; Miranda and Perissinotto 2014; Makherana et al. 2022a; Sirza et al. 2020; Weyl et al. 2020; Purnama et al. 2021). Indeed, *T. granifera* has been reported to reduce native molluscan abundances due to their dominance in invaded aquatic habitats and this has made them uncontrollable in several ecosystems globally (Chuboon et al. 2013; Veeravechsukij et al. 2018a; Oliveira et al. 2020; Purnama et al. 2020, 2021; Sirza et al. 2020; Malatji et al. 2021; Nguyen et al. 2021; Makherana et al. 2022a; Yin et al. 2022). Similar trends have been reported in South Africa, where *T. granifera* may dominate molluscan and aquatic invertebrate abundances in invaded habitats (de Kock and Wolmarans 2007; Miranda and Perissinotto 2014; Jones et al. 2017; Majdi et al. 2022) and outcompete native molluscan species.

Disease transmission

Thiarid snails pose a severe threat to public health and are of veterinary importance as they transmit parasites of fishes, birds and mammals (Sodeman 1991; Mitchell et al. 2000; McKoy et al. 2011; Veeravechsukij et al. 2018a, b). These snails act as the first intermediate hosts for trematodes that have been reported to affect the respiratory, intestinal and hepatic systems of both wildlife and humans (Sodeman 1991; McKoy et al. 2011; Veeravechsukij et al. 2018a, b; Yin et al. 2022).

Tarebia granifera are well known to harbour diverse and prevalent species of trematodes both within and outside their native ranges (Bosma 1934; Abbott 1952; Grabda 1960; Giboda et al. 1991; Ditrich et al. 1992; Wang et al. 2002; Dechruksa et al. 2007; Ukong et al. 2007; Tolley-Jordan and Owen 2008; Chontanarath and Wongsawad 2010, 2013; McKoy et al. 2011; Heneberg et al. 2014; Le et al. 2017; Veeravechsukij et al. 2018a; Doanh et al. 2019; Tolley-Jordan and Chadwick 2019; Chalkowski et al. 2021;

Paller et al. 2021; Manh Hung 2022; Yin et al. 2022). These trematodes include: *Haplorchis pumilio* Looss 1896 (Veeravechsukij et al. 2018a), *Haplorchis taichui* Nishigori 1924 (Ditrich et al. 1992; Chontanarath and Wongsawad 2010, 2013; Le et al. 2017; Veeravechsukij et al. 2018a), *Loxogenoides bicolor* Kaw 1945 (Dechruksa et al. 2007; Veeravechsukij et al. 2018a), *Centrocestus formosanus* Nishigori 1924 (Dechruksa et al. 2007; Veeravechsukij et al. 2018a), *Acanthatrium histaense* Koga 1953 (Dechruksa et al. 2007; Veeravechsukij et al. 2018a), *Haematoloechus similis* Looss 1899 (Grabda 1960; Dechruksa et al. 2007), *Transversotrema laruei* Velasquez 1958 (Veeravechsukij et al. 2018a), *Cardicola aloseae* Meade and Pratt 1965 (Ukong et al. 2007; Veeravechsukij et al. 2018a), *Alaria mustelae* Bosma 1931 (Bosma 1934; Veeravechsukij et al. 2018a) and *Philophthalmus gralli* Mathis and Léger 1910 (Tolley-Jordan and Owen 2008; Heneberg et al. 2014; Veeravechsukij et al. 2018a).

Tarebia granifera has also been reported to serve as a host for invasive parasites in Jamaica (McKoy et al. 2011) and Texas (Tolley-Jordan and Owen 2008; Tolley-Jordan and Chadwick 2019). In Jamaica, *T. granifera* has been found to be infected with two species of invasive trematodes: a *Notocotylus* sp. and a *Philophthalmus* sp. (oriental avian eye fluke). This was also the first report of a *Notocotylus* sp. in *T. granifera* and a *Philophthalmus* sp. occurring in Jamaica (McKoy et al. 2011). In Texas, invasive oriental avian eye flukes (*Philophthalmus gralli* Mathis and Leger 1910) have also been found to infect *T. granifera* (Tolley-Jordan and Owen 2008; Tolley-Jordan and Chadwick 2019; Chalkowski et al. 2021).

Wading birds are definitive hosts for notocotylids (Skirnisson et al. 2004) and many species of birds, including birds of the orders Galliformes (ground-feeding birds) and Anseriformes (waterfowl), harbour philophthalmids (Murray 1964; Nollen and Murray 1978). Humans have been reported as incidental hosts for philophthalmids in Yugoslavia, Sri Lanka, Thailand, Mexico and the United States (Rajakpase et al. 2009), where these infections cause swelling of the semilunar fold in the eyes (Mimori et al. 1982).

The trematodes *H. pumilio* and *H. taichui* are widely distributed, occurring in at least 14 countries worldwide (Ditrich et al. 1992; Huston et al. 2014; Veeravechsukij et al. 2018a), and are known to infect *T. granifera* (Chontanarath and Wongsawad 2010, 2013; Le et al. 2017; Veeravechsukij et al. 2018a) and *Melanoides tuberculata* (O.F.Müller, 1774) (Tolley-Jordan and Owen 2008; Krailas et al. 2011, 2014; Huston et al. 2014; Lopes et al. 2020; Pulido-Murillo et al. 2018). Invasion by these parasites can have severe pathological consequences for a broad range of intermediate fish hosts (Huston et al. 2014) since the penetration and migration of large numbers of cercaria can be lethal to fry and adult fish species (Sommerville 1982; Umadevi and Madhavi 2006). Natural definitive hosts are fish-eating animals, including humans (Chai et al. 2009; Giboda et al. 1991).

In some cases, biological invaders have immunity against predators, pathogens or parasites within invaded habitats; or these threats are missing from invaded habitats, thus giving the invader advantage over native species (Genner et al. 2008). Field evidence from Lake Malawi indicated

that trematodes were absent from a highly invasive morph of *M. tuberculata* and that the invader was resistant to trematodes found in native *M. tuberculata* (Genner et al. 2008). It is suspected that the parasites naturally infecting the invasive morph may not have migrated with the host or were unable to survive in the invaded habitat due to the absence of suitable intermediate or final hosts (Keane and Crawley 2002; Torchin et al. 2003; Genner et al. 2008). Trematodes are known to play a vital role in host snail populations as they can affect changes in survivorship, behaviour and morphology, and cause reduced fecundity and growth (Sorensen and Minchella 2001; Lafferty and Kuris 2009; McKoy et al. 2011). Genner et al. (2008) report that certain native trematodes act as parasitic castrators of snails and cause reduced population growth within the snail populations.

To date, no trematodes have been reported in *T. granifera* from South Africa, likely giving it an advantage over native molluscan species (Figure 2), including *M. tuberculata* (Miranda et al. 2011b; Weyl et al. 2020) and contributing to their rapid population expansion. It may be that *T. granifera* has immunity to parasites in invaded areas of South Africa and the native parasites may be unable to infect this species due to their evolutionary adaptations to native host species (Ebert 1994, 1998; Prenter et al. 2004; Fromme and Dybdahl 2006).

Classical hypotheses such as the "spillover" and "spillback" hypotheses are important to consider with regard to invasive host species (Chalkowski et al. 2018). Spillover of parasites occurs when parasites from invasive host species are introduced into a new environment and subsequently infect native species, therefore posing a health risk to native species (Daszak et al. 2000; Lymbery et al. 2014; Iglesias et al. 2015; Chalkowski et al. 2018). Parasite spillback occurs when native parasites from a native host infect an invasive host, while also increasing the risk to infect other native species (Hoberg et al. 2002; Kelly et al. 2009; Chalkowski et al. 2018). Appleton et al. (2009) suggests that the rapid spread of *T. granifera* and the resultant replacement of native *M. tuberculata* in South Africa, may worsen this problem. Based on the many possible ways invasive and native host-parasite species can interact, it might be possible that *T. granifera* can (a) dilute native parasite populations via host displacement, (b) serve as a host to invasive parasites introduced via migratory birds, or (c) eventually serve as a host for native parasite taxa that spillover from native hosts (Chalkowski et al. 2018). Even though no parasites have been reported from *T. granifera* in South Africa, the native thiarid *M. tuberculata* was previously reported to be infected with at least nine unidentified cercariae (Faust 1921; Porter 1938), which have been described under the generic collective group *Cercaria* (Müller 1773). The last reports describing parasites from *M. tuberculata* in South Africa were, however, in 1921 by Faust and in 1938 by Porter. It is, therefore, quite likely that the estimated trematode diversity in this snail might be higher than expected considering *M. tuberculata* is known to be a host for diverse groups of trematodes worldwide (Pinto and de Melo 2011). Since *T. granifera* can reach high densities within invaded habitats, and may replace *M. tuberculata* and several other native snail species,

studies on the parasitic diversity of both these thiarid species in South Africa will be of great significance (Malatji et al. 2021). Experimental studies may further our understanding of the suitability of *T. granifera* and *M. tuberculata* as hosts for native and/or invasive trematode species in South Africa (Malatji et al. 2021). This will contribute not only to a better understanding of the biodiversity and biology of the snail hosts, but also to their evolutionary relationships with parasitic trematodes.

Dispersal mechanisms

Alien species with the ability to spread through a variety of mechanisms are able to become highly invasive over a wide distribution area and have a higher probability of causing environmental changes in newly invaded areas (Moodley et al. 2013; Novoa et al. 2016; Kesner and Kumschick 2018). *Tarebia granifera* may be able to spread successfully from one waterbody to another through a variety of mechanisms (Figure 2). One of these includes the digestive system of birds where aquatic birds eat snails whole in one habitat, and then may pass them out unharmed in another habitat (Appleton et al. 2009). Indeed, small individuals of *T. granifera* (5–7 mm) have been reported in the droppings of aquatic birds in the Mhlali River in KwaZulu-Natal (Appleton et al. 2009). As some of the shells were still intact, the snails may have been alive when they passed through the bird's digestive system, and it is thought that they would perhaps have survived if they had been dropped in water (Appleton et al. 2009). Thiarid snails, including *M. tuberculata*, are known to form part of the diet of birds (Appleton 2002), including the White-faced Duck, *Dendrocygna viduata* (Linnaeus 1766; Hockey et al. 2005). According to Halse (1984), food is retained in the gut of aquatic birds for between two and six hours, and thus dispersal of snails (that survive in the gut) is likely to occur as these birds move over short distances from one aquatic ecosystem to the next (Appleton et al. 2009).

Tarebia granifera have also been found attached to floating clumps of macrophytes in rivers, thus making dispersal downstream rapid and easy (Appleton et al. 2009). Another mode of dispersal may be through drying mud on large mammals, such as elephant (*Loxodonta africana* Blumenback 1797) and buffalo (*Syncerus caffer* Sparrman 1779), particularly in conservation areas such as the Kruger National Park (Vanschoenwinkel et al. 2008, 2011; van Leeuwen et al. 2013). It is likely that *T. granifera* is also dispersed between habitats by anthropogenic means, including on boats or mud attached to footwear or vehicle wheels. Although this has not specifically been determined for *T. granifera*, many aquatic species are reportedly transported this way including mussels, bivalves, invasive aquatic plants, such as *Salvinia molesta*, and large branchiopods (Ricciardi and MacIsaac 2000; Leuven et al. 2009; Waterkeyn et al. 2010).

Mechanisms underlying impacts of *Tarebia granifera*

The introduction of invasive species to aquatic ecosystems has become a key ecological factor, as invading organisms increasingly alter aquatic communities (Gurevitch and

Padilla 2004; Almeida et al. 2018). Species that establish themselves outside of their native ranges usually achieve densities higher than they would in their native areas and often have negative impacts on native species as a result (Carlton et al. 1990; Gurevitch and Padilla 2004; Gallardo et al. 2016; Majdi et al. 2022). *Tarebia granifera* populations can reach incredibly high densities, up to 10 000 m⁻², and are often the dominant invertebrate species in invaded aquatic communities (Pillay and Perissinotto 2008; Appleton et al. 2009; Jones et al. 2020; Scharler et al. 2020; Mujiono and Isnainingsih 2021; Purnama et al. 2021; Majdi et al. 2022; Nwoko et al. 2022). Appleton et al. (2009) reported an average density of 20 764 m⁻² from the Nseleni River in KwaZulu-Natal, while Weyl et al. (2020) reported on a snail community from the Phongolo River, indicating that 93% of the biomass consisted of *T. granifera*. It is, therefore, likely that this snail negatively impacts the indigenous benthos and community structures in invaded water bodies, resulting in a decline in the abundance of native species (Hillebrand et al. 2002; Appleton et al. 2009; Jones et al. 2017; Miranda et al. 2011a; Miranda and Perissinotto 2014; Sirza et al. 2020; Purnama et al. 2021; Majdi et al. 2022; Nwoko et al. 2022; Oetama and Purnama 2022). Native aquatic species may be particularly vulnerable to interference from dense populations of *T. granifera*. For example, high densities of *T. granifera* may cause decreased growth rates in native snail species, leading to delays in sexual maturity (reduced reproduction) and causing an increase in mortality and displacement or total eradication (Hillebrand et al. 2002; de Kock and Wolmarans 2008; Appleton et al. 2009; Jones et al. 2017; Miranda et al. 2011a, b; Miranda and Perissinotto 2012, 2014; Rico-Sánchez et al. 2020). Numerous studies, both globally and in South Africa, suggest that *T. granifera* may have severe negative impacts on native aquatic snail communities (Hillebrand et al. 2002; Appleton et al. 2009; López-López et al. 2009; Jones et al. 2017; Pointier et al. 2010; Miranda et al. 2011a; Miranda and Perissinotto 2014; Purnama et al. 2021; Nwoko et al. 2022; Oetama and Purnama 2022) although few studies have assessed the specific impacts. Further research on the ecological impact of this invader on aquatic community structures and biodiversity is urgently needed (Appleton et al. 2009; Weyl et al. 2020; Purnama et al. 2022; Majdi et al. 2022; Raphahlelo et al. 2022).

The invasion success of *T. granifera* may be directly related to the release of chemical cues that repel native gastropods (Figure 2) (Raw et al. 2013, 2015). Although chemoreception in snails is well researched (Wollerman et al. 2003; Picker and Griffiths 2011; Raw et al. 2013, 2015), it was assumed, until quite recently, that competition for food and space are the main mechanisms by which *T. granifera* displace native snail populations. A study by Raw et al. (2013), however, found that three snails native to South Africa, *Assiminea* cf. *capensis* (Bartsch 1915), *Coriandria durbanensis* (Tomlin 1916) and *M. tuberculata*, responded by moving away from food sources, due to chemical cues released by *T. granifera*. In the Dominican Republic, *T. granifera* was introduced to *Biomphalaria glabrata* (Say 1818) habitats, and within 14 months *T. granifera* was displacing *B. glabrata* (Perez et al. 1991). This displacement was not due to competition for food or space, but possibly

due to chemicals secreted by the invader. These chemical cues have the potential to impact native snail behaviour, causing them to vacate optimal habitat, reducing biotic resistance and thus further facilitating invasion success (Raw et al. 2013, 2015). Although the mechanism is not fully understood, this has led to intentional introductions of *T. granifera* and *M. tuberculata* as biocontrol agents for *Schistosoma* host snails in several countries globally (Prentice 1983; Perez et al. 1991; Sodeman 1991; Pointier and Jourdan 2000; Appleton et al. 2009; Pointier et al. 2010; Hewitt and Willingham 2019). Schistosomiasis infects approximately 4.5 million people in South Africa annually (Lothe et al. 2018), and the displacement or eradication of native snails that transmit schistosomiasis by *T. granifera* may, therefore, reduce the risk of infection to people; although, this may in turn have severe ecological consequences for invaded habitats.

Tarebia granifera have reportedly caused local extinctions of native snail populations in Cuba (Pointier and Jourdan 2000; Karatayev et al. 2009), Venezuela (Pointier and Giboda 1999), Puerto Rico (Giboda et al. 1997; Chaniotis et al. 1980a, b) and South Africa (de Kock and Wolmarans 2008; Appleton et al. 2009; Miranda et al. 2011a, b; Miranda and Perissinotto 2012, 2014; Jones et al. 2017; Kesner and Kumschick 2018; Zengeya et al. 2020). Native molluscs, such as *Chambardia wahlbergi* (Krauss 1848), *Corbicula fluminalis* (Krauss 1848), *Cleopatra ferruginea* (Lea 1850), *Thiara amarula* (Linnaeus 1758), *Lanistes ovum* (Troschel 1845) and *M. tuberculata*, are considered vulnerable in South Africa (Appleton et al. 2009) and *T. granifera* poses a considerable risk to these species. It has already been reported that, once introduced, *T. granifera* rapidly outnumbers *M. tuberculata* in terms of density (Pointier et al. 1998). Further, *M. tuberculata* and *T. amarula* have become less common in South Africa in areas with higher densities of *T. granifera* (Miranda et al. 2011a).

Apart from its environmental impacts and the role as an intermediate host for medically and economically important trematodes, *T. granifera* may also pose a risk to important infrastructure (Appleton et al. 2009; Yakovenko et al. 2018; Oleh et al. 2018; Yesipova et al. 2022). The snail is abundant in reservoirs of three large industrial plants in northern KwaZulu-Natal. The shells block pipes and damage equipment and have also interfered with water circulation at a nearby fish hatchery (Appleton et al. 2009). In addition, *T. granifera* and *M. tuberculata* have been reported to threaten the hydraulic cooling structures in the Zaporizhia Nuclear Power Plant (ZNPP), Ukraine (Yakovenko et al. 2018; Oleh et al. 2018; Yesipova et al. 2022). Snails reached densities of up to 5 200 individuals m⁻² in the basins and metal pipes, and shells created obstacles which decreased effectiveness of pumping stations, meaning such infrastructure needs constant mechanical cleaning (Yesipova et al. 2022).

Feeding dynamics and food web interactions

Tarebia granifera may have severe impacts on benthic biomass that could have implications on food availability for native aquatic species and result in decreased benthic biodiversity, as well as cause disturbances

to aquatic ecosystem function (Figure 2) (March and Pringle 2003; Miranda et al. 2010, 2011a; Moslemi et al. 2012; Raw et al. 2013; Miranda and Perissinotto 2014; Hill et al. 2015; Rico-Sánchez et al. 2020; Majdi et al. 2022). This invasive snail feeds mainly on algae (mostly diatoms) and semi-decomposed organic matter, and its feeding impact can be much higher than that of other invertebrates (Miranda et al. 2011a). Miranda et al. (2011a) reported that *T. granifera* utilised up to 35% of available microphytobenthos (photosynthetic diatoms, cyanobacteria, flagellates and green algae) per day, and up to 68% of the daily primary benthic production in coastal and estuarine lakes in South Africa, with similar findings reported for zooplankton species (Perissinotto, 1992; Kibirige and Perissinotto 2003). This mollusc thus poses a significant threat to algal and zooplankton communities, potentially causing direct bottom-up and cascading effects on an ecosystem and its associated food web (Miranda et al. 2010; Miranda and Perissinotto 2014). According to Miranda et al. (2011a), the ingestion rate of *T. granifera* is comparable to that of the Golden apple snail (*Pomacea canaliculata* Lamarck 1822), among the world's most destructive invasive species (Lowe et al. 2000; Baker et al. 2010).

As invasive species disrupt trophic functioning, native species also tend to shift their dietary preference (Vander Zanden et al. 1999). This was evident in coastal lakes from KwaZulu-Natal where the native snail *Assimineia ovata* (Krauss 1848) started feeding less on microphytobenthos and more on branching algae after *T. granifera* became established (Miranda and Perissinotto 2012). Invasive *T. granifera* populations in the San Marcos River (Texas) have also been reported to consume the eggs of the threatened Fountain Darter fish (*Etheostoma fonticola* Jordan and Gilbert 1886) with potential negative consequences (Phillips et al. 2010). The widespread diet of *T. granifera* has likely contributed to its successful establishment in southern Africa (Miranda and Perissinotto 2012).

As native snail species are replaced by *T. granifera* in South Africa, native predators such as shrimps (*Macrobrachium* sp. Spence Bate 1868), crabs (*Potamonautes* sp. Macleay 1838) and dragonflies (Libellulidae) may also be impacted as many of these predators are unable to break the harder shells of *T. granifera*, increasing the likelihood of indirect bottom-up effects on invaded ecosystems (Appleton et al. 2009; Miranda et al. 2016). Hermit crabs in Tobago, (*Clibanarius tricolor* Gibbes 1850 and *Clibanarius vittatus* Bosc 1802) have started making use of *T. granifera* shells that washed downstream (Van Oosterhout et al. 2013). This may make hermit crabs less vulnerable to predators and have additional indirect bottom-up effects on aquatic food webs.

Tarebia granifera are important bioturbation agents and at high densities affect nutrient cycles of invaded aquatic ecosystems (Figure 2) (Arnott and Vanni 1996; Hall et al. 2003; Carlsson et al. 2004). Moslemi et al. (2012) found riparian cover influenced the effect of *T. granifera* on nitrogen (N) cycling in West Indian streams. This study found that snail densities were up to eight times higher, and nitrogen excretion up to nine times higher, in open-canopy habitats compared to habitats with extensive riparian

vegetation. Changes to the availability of nutrients may influence primary production and the growth and community structures of primary producers (Vanni 2002), with unknown implications for ecosystem function (Moslemi et al. 2012). Comprehensive research on the effects of *T. granifera* on aquatic food webs in invaded habitats are limited, but greatly needed, as *T. granifera* is a globally problematic invader.

***Tarebia granifera* as prey**

Alien species often have higher competitive abilities in invaded areas due to the enemy release hypothesis, e.g. resistance to predation, parasites and pathogens (Elton 1958; Tilman 1999). Although *T. granifera* can be preyed upon by birds, fish and invertebrates, it does not seem to have many successful predators in South Africa (Miranda and Perissinotto 2012; Miranda et al. 2016; Whitfield et al. 2021). Predators will often feed on the most abundant prey (known as frequency-dependent predation), but prey preference is also determined by energy gained from the prey relative to the energy cost of catching the prey (Morrison et al. 2019). *Potamonautes sidneyi* (Rathbun 1904), a crab native to South Africa, preys upon *T. granifera* in Lake Sibaya, KwaZulu-Natal (Peer et al. 2015). Miranda et al. (2016) reported that *P. sidneyi* and *P. perlatus* (Milne Edwards 1837) were, however, often unable to successfully crush the rigid shell of *T. granifera* compared to the native *M. tuberculata*. Although some large male crabs were able to crush the shells of *T. granifera*, successful attacks were less frequent compared with attacks on native *M. tuberculata*. In cases when alien prey species are harder to consume, predators will be more likely to select native species, giving the invasive species further advantage (Figure 2) (Shinen et al. 2009; López et al. 2010).

Morrison et al. (2019) suggest that Eastern musk turtles *Sternotherus odoratus* (Latreille 1802) in Texas may have shifted their diets to include invasive snails over the last three decades. *Melanoides tuberculata*, *T. granifera*, and the Giant Ramshorn Snail *Marisa cornuarietis* (Linnaeus 1758), all invasive in the USA, were the most abundant species identified from *S. odoratus* faecal samples and *T. granifera* was found in half the samples investigated during the study (Morrison et al. 2019).

Interactions with other invasive species

Ecosystems are frequently invaded by multiple invasive species, and interactions among species may amplify negative impacts on native communities and ecosystems (Johnson et al. 2009; Makherana et al. 2022b). A mesocosm study in the USA examined interactions between invasive Rusty crayfish *Orconectes rusticus* (Girard 1852) and Chinese Mystery snails *Cipangopaludina chinensis* (Gray 1834). Both invaders had weak negative impacts on one another, but both negatively affected native snail biomass and abundance (Johnson et al. 2009). These invasive species drove one native snail to local extinction and reduced a second species by more than 95%. Rusty crayfish (a snail predator) were also significantly more likely to consume native snails compared to the thicker shell and larger sized invader (Johnson et al. 2009).

Three molluscan species (*Corbicula fluminea* (Müller 1774), *M. tuberculata* and *T. granifera*) invasive in the

Tuxpan and Tecolutla Rivers in Mexico have negatively impacted native molluscan abundances (López-López et al. 2009). *Melanoides tuberculata* and native molluscan species in these areas have reportedly been confined to narrow habitats with a limited distribution range, and occurred in low densities, while *T. granifera* and *C. fluminea* indicated strong dominance and wider distribution. López-López et al. (2009) further indicated that *T. granifera* reached higher population densities than the other invasive species, likely by out-competing the other species and by rapidly recolonising ecosystems after pronounced depletions in snail density during the heavy rainy season. *Tarebia granifera* populations grew quickly during the dry season and were more successful than both other invasive snails and native snails (López-López et al. 2009).

Negative interactions between native and invasive species may also be due to differences in habitat use rather than direct competition (Tolley-Jordan and Owen 2008). A study conducted in Texas found that the invasive snails *M. tuberculata* and *T. granifera* co-occurred in lentic habitats while *M. tuberculata* and the native *Elimia comalensis* (Pilsbry 1890) did not co-occur (Tolley-Jordan and Owen 2008). In lentic habitats, the invaders had similar densities and even though *E. comalensis* had the highest densities of the native species, it was still 200 times less than the densities of the invaders (Tolley-Jordan and Owen 2008). Conversely, in lotic habitats *T. granifera* and *E. comalensis* distributions positively co-occurred and they had similar densities, while the density of *M. tuberculata* was ten times lower (Tolley-Jordan and Owen 2008). This is possibly because *T. granifera* is able to tolerate faster water flow than *M. tuberculata* (Dussart and Pointier 1999). The preference of *E. comalensis* for lotic habitats allowed it to occur in densities similar to that of *T. granifera* (Tolley-Jordan and Owen 2008).

Intense competitive interactions occur among exotic species with similar ecological traits. For instance, the invasion of *T. granifera* and a new morph of *M. tuberculata* (MA strain) seriously impacted other morphs of *M. tuberculata* in Guadeloupe and Martinique, Caribbean Islands (Pointier et al. 2010). These invasions led to the extinction of one morph of *M. tuberculata* (FDF strain) while others became scarce. Similar findings have been reported in the Great African Lakes where the invasion of Asian morphs of *M. tuberculata* threatens native thiarid diversity (Genner et al. 2004). In systems where endemic thiarid diversity is high, similar threats can be expected from invasive thiarid species (Pointier et al. 2005; Simone 2006; Pointier et al. 2010). Although *M. tuberculata* is native to South Africa, invasive morphs may be present due to the complex global invasion history of the snail (Genner et al. 2004; Van Bocxlaer et al. 2015; Miranda et al. 2016).

Invasive snail populations should not be regarded as genetically depauperate (Pointier et al. 2010). Introductions of *M. tuberculata* to Martinique and Guadeloupe gave rise to rare sexual events between introduced morphs, forming natural hybrids which have high levels of genetic variation and ecologically important traits (Facon et al. 2008). Different morphology and life history traits allow these hybrids to be very successful (Facon et al. 2005).

Control measures

The complete eradication of *T. granifera* from an environment is particularly difficult due to its parthenogenic life history (Miranda et al. 2010). The use of molluscicides in the natural environment is not a practical method of control as non-target species may be adversely affected (McCullough et al. 1980; Miranda et al. 2010). A 'salinity shock experiment' by Miranda et al. (2010) indicated that *T. granifera* were vulnerable to sudden increases in salinity. This led to suggestions that snails could be controlled in some estuaries by artificially breaching barriers and elevating salinity (Miranda et al. 2010). The manipulation of natural environments on such a scale may, however, also have unintended and negative consequences on non-target species and these ecosystems as a whole (Miranda et al. 2010).

The black carp *Mylopharyngodon piceus* (Richardson 1846) is a freshwater snail predator that has been effectively used as a biological control of *M. tuberculata* and *T. granifera* in several parts of the world (Ben-Ami and Heller 2001; Manh Hung et al. 2013; Yakovenko et al. 2018). The Assassin snail *Anentome helena* (von dem Busch 1847) has also been reported to be an effective biological control of *M. tuberculata* in Malaysia, Indonesia, Thailand and Laos (Schiffbauer 2009; Yakovenko et al. 2018), and a potential control agent of *T. granifera* (Yakovenko et al. 2018). The introduction of biological control agents, however, has numerous environmental risks that need to be considered, and extensive research needs to be done before attempting to use these measures in new environments (Howarth 1991; Simberloff and Stiling 1996; Hoddle 2004; Louda and Stiling 2004). As previously mentioned, trematodes play a vital role in the natural control of snail populations due to their ability to affect snail survivorship, behaviour and morphology, causing reduced fecundity and growth (Sorensen and Minchella 2001; Genner et al. 2008; Lafferty and Kuris 2009; McKoy et al. 2011). In a study conducted by McKoy et al. (2011), only 3% of *T. granifera* infected with trematodes had reproductive stages (eggs, embryos or juveniles) compared to 90% of snails that were not infected. Though potentially significant, the ecological impact of trematodes on the fecundity of *T. granifera* and the environment needs to be investigated further (McKoy et al. 2011). Unfortunately, the control of *T. granifera* with trematodes in natural environments may also have unintended negative effects on native species and the environment. Moreover, due to a lack of invasive species monitoring and management (Makherana et al. 2022a) and the large area already occupied by *T. granifera* in the eastern and northern parts of southern Africa (Appleton et al. 2009), this snail will probably continue to expand its range (Kesner and Kumschick 2018).

In the Ukraine, Yakovenko et al. (2018) found that thiarid snails were unequally distributed in the ZNPP, with an average abundance of 3.3 *T. granifera* to one *M. tuberculata*. Experimental trials using Marbled crayfish *Procambarus virginalis* (Lyko 2017), Pumpkinseed fish *Lepomis gibbosus* (Linnaeus 1758), and the loach *Botia lohachata* (Chaudhuri 1912) were found to be unsuccessful at controlling snail populations. The Assassin snail was a successful control agent but when this predatory

mollusc has more accessible food, such as organic matter, it consumes smaller quantities of the thiarid snails (Oleh et al. 2018). According to Oleh et al. (2018), the most successful method to control *T. granifera* populations in power plant intakes was using snail traps, which attract snails with bait. Snails fall into the traps as they move towards the bait and are unable to escape (Oleh et al. 2018). The authors found that traps were also effective at catching sexually mature snails, thereby removing reproductive individuals from the population. Yakovenko et al. (2018) also suggested that the power plant should stock their cooling ponds with the molluscivorous black carp and Assassin snails. This research may lead to the development of control measures to reduce accidental introductions of snails into reservoirs of importance (Yakovenko et al. 2018; Oleh et al. 2018). Laboratory investigations on biological and biomechanical methods to control snails are ongoing (Yakovenko et al. 2018; Oleh et al. 2018; Yesipova et al. 2022) but effective control of established *T. granifera* populations in a natural setting remains challenging. The most effective method to prevent snail invasions in the future, to date, is to implement strict policies that regulate the global trade of freshwater molluscs (Preston et al. 2022).

Concluding remarks

Tarebia granifera is a highly resilient and opportunistic species that is able to flourish in disturbed environments. This enables individuals to rapidly disperse and invade a wide variety of aquatic habitats in a relatively short time. The snail has been reported to cause declines and local extinctions in native snail populations on a global scale due to their ability to reproduce quickly and reach high densities.

Furthermore, high densities of *T. granifera* may have significant impacts on benthic biomass that could have implications on food availability for native aquatic species, resulting in decreased benthic biodiversity as well as changes in the food web dynamics and nutrient cycles of invaded ecosystems (Figure 2). Apart from their environmental impacts and their role as intermediate hosts for diverse and prevalent species of medically and veterinary important trematodes on a global scale, *T. granifera* have been reported to pose an important economic threat to infrastructure in Ukraine and South Africa.

Management and remediation of biological invasions in southern Africa need a more complete understanding of *T. granifera*'s impact and the mechanisms through which they occur. Previous studies have investigated the impacts and population structures of *T. granifera* in coastal lakes and estuaries in KwaZulu-Natal. However, less is known about the current distribution, population dynamics and impacts *T. granifera* have on the native biodiversity and aquatic community structures of freshwater ecosystems in southern Africa. Given that the current distribution and densities of native as well as invasive freshwater snail species are underreported, it is critical to understand impacts of this ubiquitous invader. In addition, because this invader may also act as a potential intermediate host for parasitic trematodes in southern Africa, there is a need to investigate the potential impacts of *T. granifera* on both native ecosystems and human health.

Acknowledgements — The Water Research Commission (WRC) is hereby acknowledged for providing a Postgraduate MSc Scholarship to J.J. Pearson. The National Research Foundation (NRF)–Department of Science and Innovation (DSI) Professional Development Programme is further acknowledged for the use of infrastructure provided by the NRF–SAIAB Research Platform and for funding provided to L de Necker. We also thank the two anonymous referees for their time to review the manuscript.

Funding — JJ Pearson received scholarship funding from the Water Research Commission (Project number: C2019/2020-00151) under the supervision of Dr L de Necker, Dr R Gerber and Dr W Malherbe. L de Necker received research funding from the National Research Foundation (NRF)–Department of Science and Innovation (DSI) Professional Development Programme (Grant No. 127549) and funding channelled through the NRF-SAIAB Institutional Support system. All subsequent authors declare that they have no financial interests.

Competing interests — The authors have no known competing financial interests or personal relationships to declare that are relevant to the content of this paper or that could have appeared to influence the work reported in this paper.

Author contributions — JJ Pearson: conceptualisation, data curation (literature search, investigation); writing (original draft); writing (editing). R Gerber: conceptualisation; project administration; supervision; validation; writing (review and editing). W Malherbe: conceptualisation; project administration; supervision; validation; writing (review and editing). NJ Smit: project administration; supervision; validation; writing (review and editing). L de Necker: conceptualisation; funding acquisition; project administration; supervision; writing (original draft); writing (editing).

Data availability — Data sharing is not applicable as no datasets were generated or analysed as part of this paper.

References

- Abbott RT. 1952. A study of an intermediate snail host (*Thiara granifera*) of the oriental lung fluke (*Paragonimus*). *Proceedings of the United States National Museum* 102: 71–116. <https://doi.org/10.5479/si.00963801.102-3292.71>.
- Acosta AA, Netherlands EC, Retief F, de Necker L, du Preez L, et al. 2020. Conserving freshwater biodiversity in an African subtropical wetland: South Africa's lower Phongolo river and floodplain. In: Kideghesho JR (ed) *Managing wildlife in a changing world*. Intechopen, London, pp 11–39. <https://doi.org/10.5772/intechopen.93752>.
- Albarrán-Mélzer NC, Rangel Ruiz LJ, Benítez HA, Lagos ME. 2020. Can temperature shift morphological changes of invasive species? A morphometric approach on the shells of two tropical freshwater snail species. *Hydrobiologia* 847: 151–160. <https://doi.org/10.1007/s10750-019-04078-z>.
- Almeida PRDS, Nascimento SLD, Viana GFS. 2018)+. Effects of invasive species snails in continental aquatic bodies of Pernambuco semi-arid. *Acta Limnologica Brasiliensia* 30: e103. <https://doi.org/10.1590/s2179-975x10616>.
- Appleton CC. 2002. Mollusca. In: Day JA and de Moor IJ (ed) *Guides to the freshwater invertebrates of Southern Africa*. Water Research Commission, Pretoria. pp 42–125.
- Appleton CC. 2003. Alien and invasive freshwater Gastropoda in South Africa. *African Journal of Aquatic Science* 28: 69–81. <https://doi.org/10.2989/16085914.2003.9626602>.
- Appleton CC, Forbes AT, Demetriades NT. 2009. The occurrence, bionomics and potential impacts of the invasive freshwater snail

- Tarebia granifera* (Lamarck, 1822) (Gastropoda: Thiaridae) in South Africa. *Zoologische Mededelingen* 83: 525–536.
- Appleton CC, Miranda NAF. 2015a. Two Asian freshwater snails newly introduced into South Africa and an analysis of alien species reported to date. *African Invertebrates* 56: 1–17. <https://doi.org/10.5733/afin.056.0102>.
- Appleton CC, Miranda N. 2015b. Locating bilharzia transmission sites in South Africa: guidelines for public health personnel. *Southern African Journal of Infectious Diseases* 30: 95–102. <https://doi.org/10.1080/23120053.2015.1074438>.
- Appleton CC, Nadasan DS. 2002. First report of *Tarebia granifera* (Lamarck, 1816) (Gastropoda: Thiaridae) from Africa. *Journal of Molluscan Studies* 68: 399–402. <https://doi.org/10.1093/mollus/68.4.399>.
- Arnott DL, Vanni MJ. 1996. Nitrogen and phosphorus recycling by the zebra mussel (*Dreissena polymorpha*) in the western basin of Lake Erie. *Canadian Journal of Fisheries and Aquatic Sciences* 53: 646–659. <https://doi.org/10.1139/f95-214>.
- Baker P, Zimmanck F, Baker SM. 2010. Feeding rates of an introduced freshwater gastropod *Pomacea insularum* on native and nonindigenous aquatic plants in Florida. *Journal of Molluscan Studies* 76: 138–143. <https://doi.org/10.1093/mollus/eyp050>.
- Ben-Ami F, Heller J. 2001. Biological control of aquatic pest snails by the black carp *Mylopharyngodon piceus*. *Biological Control* 22: 131–138. <https://doi.org/10.1006/bcon.2001.0967>.
- Bogan AE. 2007. Global diversity of freshwater mussels (Mollusca, Bivalvia) in freshwater. In: Balian EV, Lévêque C, Segers H, Martens K (ed) *Freshwater animal diversity assessment. Developments in hydrobiology*. Springer, Dordrecht. 198: 139–147. https://doi.org/10.1007/978-1-4020-8259-7_16.
- Bosma NJ. 1934. The life history of the trematode *Alaria mustelae*, Bosma, 1931. *Transactions of the American Microscopical Society* 53: 116–153. <https://doi.org/10.2307/3222088>.
- Carlsson NO, Brönmark C, Hansson LA. 2004. Invading herbivory: the golden apple snail alters ecosystem functioning in Asian wetlands. *Ecology* 85: 1575–1580. <https://doi.org/10.1890/03-3146>.
- Carlton JT, Thompson JK, Schemel LE, Nichols FH. 1990. Remarkable invasion of San Francisco Bay (California, USA) by the Asian clam *Potamocorbula amurensis*. I. Introduction and dispersal. *Marine Ecology Progress Series* 66: 81–94. <https://doi.org/10.3354/meps066081>.
- Chai JY, Shin EH, Lee SH, Rim HJ. 2009. Foodborne intestinal flukes in Southeast Asia. *The Korean Journal of Parasitology* 47: S69–S102. <https://doi.org/10.3347/kjp.2009.47.S.S69>.
- Chalkowski K, Lepczyk CA, Zohdy S. 2018. Parasite ecology of invasive species: conceptual framework and new hypotheses. *Trends in Parasitology* 34: 655–663. <https://doi.org/10.1016/j.pt.2018.05.008>.
- Chalkowski K, Morgan A, Lepczyk CA, Zohdy S. 2021. Spread of an avian eye fluke, *Philophthalmus gralli*, through biological invasion of an intermediate host. *The Journal of Parasitology* 107: 336–348. <https://doi.org/10.1645/20-72>.
- Chaniotis BN, Butler JM, Ferguson FF, Jobin WR. 1980a. Bionomics of *Tarebia granifera* (Gastropoda: Thiaridae) in Puerto Rico, an Asiatic vector of *paragonimiasis westermani*. *Caribbean Journal of Science* 16: 81–90.
- Chaniotis BN, Butler JM, Ferguson F, Jobin WR. 1980b. Presence of males in Puerto Rican *Thiara (Tarebia) granifera* (Gastropoda: Thiaridae), a snail thought to be parthenogenetic. *Caribbean Journal of Science* 16: 95–97.
- Chaniotis BN, Butler JM, Ferguson FF, Jobin WR. 1980c. Thermal limits, desiccation tolerance, and humidity reactions of *Thiara (Tarebia) granifera muiensis* (Gastropoda: Thiaridae) host of the Asiatic lung fluke disease. *Caribbean Journal of Science* 16: 91–93.
- Chen KJ. 2003. A preliminary study on the reproductive ecology of the freshwater snail *Tarebia granifera* (Lamarck, 1822) (Prosobranchia: Thiaridae) in Jinlun River, South-eastern Taiwan. Thesis, National Sun Yat-Sen University.
- Chontanarath T, Wongsawad C. 2010. Prevalence of *Haplorchis taichui* in field-collected snails: a molecular approach. *The Korean Journal of Parasitology* 48: 343–346. <https://doi.org/10.3347/kjp.2010.48.4.343>.
- Chontanarath T, Wongsawad C. 2013. Prevalence of *Haplorchis taichui* infection in snails from Mae Taeng River Basin, Chiang Mai Province, by techniques of morphology and molecular biology. *Journal of Yala Rajabhat University* 8: 9–21.
- Chuboon S, Wongsawad C, Wongsawad P. 2013. Molecular identification of trematode, *Haplorchis taichui* cercariae (Trematoda: Heterophyidae) in *Tarebia granifera* snail using ITS2 sequences. *Journal of Yala Rajabhat University* 8: 22–30.
- Collen B, Whitton F, Dyer EE, Baillie JE, Cumberlidge N, et al. 2014. Global patterns of freshwater species diversity, threats and endemism. *Global Ecology and Biogeography* 23: 40–51. <https://doi.org/10.1111/geb.12096>.
- Contreras-Arquieta A, Contreras-Balderas S. 1999. Description, biology, and ecological impacts of the screw snail, *Thiara tuberculata* (Müller, 1774) (Gastropoda: Thiaridae) in Mexico. In: Claudi R, Leach J H (ed) *Nonindigenous Freshwater Organisms. Vectors, Biology, and Impacts*. Lewis Publishers, Boca Raton, FL. pp 151–160.
- Cooke SJ, Allison EH, Beard TD, Arlinghaus R, Arthington AH, et al. 2016. On the sustainability of inland fisheries: Finding a future for the forgotten. *Ambio* 45: 753–764. <https://doi.org/10.1007/s13280-016-0787-4>.
- Cowie RH. 1998. Patterns of introduction of non-indigenous non-marine snails and slugs in the Hawaiian Islands. *Biodiversity and Conservation* 7: 349–368. <https://doi.org/10.1023/A:1008881712635>.
- Daszak P, Cunningham AA, Hyatt AD. 2000. Emerging infectious diseases of wildlife—threats to biodiversity and human health. *Science* 287: 443–449. <https://doi.org/10.1126/science.287.5452.443>.
- de Kock KN, Wolmarans CT. 2007. Distribution and habitats of the alien invader freshwater snail *Physa acuta* in South Africa. *Water SA* 33: 717–722. <https://doi.org/10.4314/wsa.v33i5.184093>.
- de Kock KN, Wolmarans CT. 2008. Invasive alien freshwater snail species in the Kruger National Park, South Africa. *Koedoe: African Protected Area Conservation and Science* 50: 49–53. <https://doi.org/10.4102/koedoe.v50i1.126>.
- de Necker L, Brendonck L, van Vuren J, Wepener V, Smit NJ. 2021. Aquatic invertebrate community resilience and recovery in response to a supra-seasonal drought in an ecologically important naturally saline lake. *Water* 13: 948. <https://doi.org/10.3390/w13070948>.
- Dechruksa W, Krailas D, Ukong S, Inkapananakul W, Koonchornboon T. 2007. Trematode infections of the freshwater snail family Thiaridae in the Khek river, Thailand. *The Southeast Asian Journal of Tropical Medicine and Public Health* 38: 1016–1028.
- Di Marco M, Chapman S, Althor G, Kearney S, Besancon C, et al. 2017. Changing trends and persisting biases in three decades of conservation science. *Global Ecology and Conservation* 10: 32–42. <https://doi.org/10.1016/j.gecco.2017.01.008>.
- Ditrich O, Na V, Scholz T, Giboda M. 1992. Larval stages of medically important flukes (Trematoda) from Vientiane province, Laos. Part II. Cercariae. *Annals of Human and Comparative Parasitology* 67: 75–81. <https://doi.org/10.1051/parasite/199267375>.
- Doanh PN, Van Hien H, Dung BT, Loan HT. 2019. Infection status and molecular identification of digenetic cercariae in snails in Kim Son district, Ninh Binh Province and Ba Vi district, Ha Noi. *Academia Journal of Biology* 41: 31–38. <https://doi.org/10.15625/2615-9023/v41n3.13893>.
- Dube T, de Necker L, van Vuren JH, Wepener V, Smit NJ, Brendonck L. 2017. Spatial and temporal variation of invertebrate

- community structure in flood-controlled tropical floodplain wetlands. *Journal of Freshwater Ecology* 32: 1–15. <https://doi.org/10.1080/02705060.2016.1230562>.
- Dudgeon D, Arthington AH, Gessner MO, Kawabata ZI, Knowler DJ, et al. 2006. Freshwater biodiversity: importance, threats, status and conservation challenges. *Biological Reviews* 81: 163–182. <https://doi.org/10.1017/S1464793105006950>.
- Dussart GBJ, Pointier JP. 1999. A comparative study of the hydrodynamic performance of shells of *Tarebia granifera* (Lamarck, 1822) and *Melanooides tuberculata* (Muller, 1774). *Malacological Review (Supplement)* 8: 19–29.
- Ebert D. 1994. Virulence and local adaptation of a horizontally transmitted parasite. *Science* 265: 1084–1086. <https://doi.org/10.1126/science.265.5175.1084>.
- Ebert D. 1998. Experimental evolution of parasites. *Science* 282: 1432–1436. <https://doi.org/10.1126/science.282.5393.1432>.
- Elton C. 1958. *The ecology of invasions by animals and plants* 4th ed. University of Chicago press, Chicago. <https://doi.org/10.1007/978-1-4899-7214-9>.
- Facon B, Genton BJ, Shykoff J, Jarne P, Estoup A, David P. 2006. A general eco-evolutionary framework for understanding bioinvasions. *Trends in Ecology and Evolution* 21: 130–135. <https://doi.org/10.1016/j.tree.2005.10.012>.
- Facon B, Jarne P, Pointier JP, David P. 2005. Hybridization and invasiveness in the freshwater snail *Melanooides tuberculata*: hybrid vigour is more important than increase in genetic variance. *Journal of Evolutionary Biology* 18: 524–535. <https://doi.org/10.1111/j.1420-9101.2005.00887.x>.
- Facon B, Machline E, Pointier JP, David P. 2004. Variation in desiccation tolerance in freshwater snails and its consequences for invasion ability. *Biological Invasions* 6: 283–293. <https://doi.org/10.1023/B:BINV.0000034588.63264.4e>.
- Facon B, Pointier JP, Jarne P, Sarda V, David P. 2008. High genetic variance in life-history strategies within invasive populations by way of multiple introductions. *Current Biology* 18: 363–367. <https://doi.org/10.1016/j.cub.2008.01.063>.
- Faulkner KT, Burness A, Byrne MJ, Kumschick S, Peters K, et al. 2020. South Africa's pathways of introduction and dispersal and how they have changed over time. In: van Wilgen BW, Measey J, Richardson DM, Wilson JR, Zengeya TA (ed) *Biological invasions in South Africa*. Springer, Berlin, pp 313–354. https://doi.org/10.1007/978-3-030-32394-3_12.
- Faust EC. 1921. Notes on South African larval trematodes. *The Journal of Parasitology* 8: 11–21. <https://doi.org/10.2307/3270937>.
- Fromme AE, Dybdahl MF. 2006. Resistance in introduced populations of a freshwater snail to native range parasites. *Journal of Evolutionary Biology* 19: 1948–1955. <https://doi.org/10.1111/j.1420-9101.2006.01149.x>.
- Gallardo B, Clavero M, Sánchez MI, Vilà M. 2016. Global ecological impacts of invasive species in aquatic ecosystems. *Global Change Biology* 22: 151–163. <https://doi.org/10.1111/gcb.13004>.
- GBIF. 2021. Global Biodiversity Information Facility. GBIF occurrence download. <https://doi.org/10.15468/dl.e2dytc>. [Accessed 7 September 2021]
- Genner MJ, Michel E, Erpenbeck D, De Voogd N, Witte F, Pointier JP. 2004. Camouflaged invasion of Lake Malawi by an Oriental gastropod. *Molecular Ecology* 13: 2135–2141. <https://doi.org/10.1111/j.1365-294X.2004.02222.x>.
- Genner MJ, Michel E, Todd JA. 2008. Resistance of an invasive gastropod to an indigenous trematode parasite in Lake Malawi. *Biological Invasions* 10: 41–49. <https://doi.org/10.1007/s10530-007-9105-1>.
- Giboda M, Ditrich O, Scholz T, Viengsay T, Bouaphanh S. 1991. Human *Opisthorchis* and *Haplorchis* infections in Laos. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 85: 538–540. [https://doi.org/10.1016/0035-9203\(91\)90248-W](https://doi.org/10.1016/0035-9203(91)90248-W).
- Giboda M, Malek EA, Correa R. 1997. Human schistosomiasis in Puerto Rico: reduced prevalence rate and absence of *Biomphalaria glabrata*. *The American Journal of Tropical Medicine and Hygiene* 57: 564–568. <https://doi.org/10.4269/ajtmh.1997.57.564>.
- Grabda B. 1960. Life cycle of *Haematoloechus similis* (Looss, 1899) (Trematoda-Plagiorchiidae). *Acta Parasitologica Polonica* 8: 357–367.
- Gurevitch J, Padilla DK. 2004. Are invasive species a major cause of extinctions? *Trends in Ecology and Evolution* 19: 470–474. <https://doi.org/10.1016/j.tree.2004.07.005>.
- Hall RO, Tank JL, Dybdahl MF. 2003. Exotic snails dominate nitrogen and carbon cycling in a highly productive stream. *Frontiers in Ecology and the Environment* 1: 407–411. [https://doi.org/10.1890/1540-9295\(2003\)001\[0407:ESDNAC\]2.0.CO;2](https://doi.org/10.1890/1540-9295(2003)001[0407:ESDNAC]2.0.CO;2).
- Halse SA. 1984. Food intake, digestive efficiency and retention time in spur-winged geese *Plectropterus gambensis*. *South African Journal of Wildlife Research* 14: 106–110.
- Havel JE, Kovalenko KE, Thomaz SM, Amalfitano S, Kats LB. 2015. Aquatic invasive species: challenges for the future. *Hydrobiologia* 750: 147–170. <https://doi.org/10.1007/s10750-014-2166-0>.
- Heneberg P, Rojas A, Bizos J, Kocková L, Malá M, Rojas D. 2014. Focal *Philophthalmus gralli* infection possibly persists in *Melanooides tuberculata* over two years following the definitive hosts' removal. *Parasitology International* 63: 802–807. <https://doi.org/10.1016/j.parint.2014.07.012>.
- Hewitt R, Willingham AL. 2019. Status of schistosomiasis elimination in the Caribbean region. *Tropical Medicine and Infectious Disease* 4: 24–41. <https://doi.org/10.3390/tropicalmed4010024>.
- Hill JM, Jones RW, Hill MP, Weyl OLF. 2015. Comparisons of isotopic niche widths of some invasive and indigenous fauna in a South African river. *Freshwater Biology* 60: 893–902. <https://doi.org/10.1111/fwb.12542>.
- Hillebrand H, Kahlert M, Haglund AL, Berninger UG, Nagel S, Wickham S. 2002. Control of microbenthic communities by grazing and nutrient supply. *Ecology* 83: 2205–2219. [https://doi.org/10.1890/0012-9658\(2002\)083\[2205:COMCBG\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2002)083[2205:COMCBG]2.0.CO;2).
- Hoberg EP, Kutz SJ, Nagy J, Jenkins E, Elkin B, Branigan M, Cooley D. 2002. *Protostrongylus stilesi* (Nematoda: Protostrongylidae): Ecological isolation and putative host-switching between Dall's sheep and muskoxen in a contact zone. *Comparative Parasitology* 69: 1–9. [https://doi.org/10.1654/1525-2647\(2002\)069\[0001:PSNPEI\]2.0.CO;2](https://doi.org/10.1654/1525-2647(2002)069[0001:PSNPEI]2.0.CO;2).
- Hockey PAR, Dean WRJ, Ryan PG, Maree S. 2005. *Roberts' birds of southern Africa*, 7th ed. John Voelcker Bird Book Fund, Cape Town.
- Hodde MS. 2004. Restoring balance: using exotic species to control invasive exotic species. *Conservation Biology* 18: 38–49. <https://doi.org/10.1111/j.1523-1739.2004.00249.x>.
- Howarth FG. 1991. Environmental impacts of classical biological control. *Annual Review of Entomology* 36: 485–509. <https://doi.org/10.1146/annurev.en.36.010191.002413>.
- Huston DC, Worsham MD, Huffman DG, Ostrand KG. 2014. Infection of fishes, including threatened and endangered species by the trematode parasite *Haplorchis pumilio* (Looss, 1896) (Trematoda: Heterophyidae). *BioInvasions Record* 3: 189–194. <https://doi.org/10.3391/bir.2014.3.3.09>.
- Iglesias R, García-Estévez JM, Ayres C, Acuña A, Cordero-Rivera A. 2015. First reported outbreak of severe spirorchidiasis in *Emys orbicularis*, probably resulting from a parasite spillover event. *Diseases of Aquatic Organisms* 113: 75–80. <https://doi.org/10.3354/dao02812>.
- Johnson PT, Olden JD, Solomon CT, Vander Zanden MJ. 2009. Interactions among invaders: community and ecosystem

- effects of multiple invasive species in an experimental aquatic system. *Oecologia* 159: 161–170. <https://doi.org/10.1007/s00442-008-1176-x>.
- Johnson PD, Bogan AE, Brown KM, Burkhead NM, Cordeiro JR, et al. 2013. Conservation status of freshwater gastropods of Canada and the United States. *Fisheries* 38: 247–282. <https://doi.org/10.1080/03632415.2013.785396>.
- Jones S, Carrasco NK, Perissinotto R, Fox C. 2020. Abiotic and biotic responses to the 2016/2017 restoration project at the St Lucia Estuary mouth, South Africa. *African Journal of Aquatic Science* 45: 153–166. <https://doi.org/10.2989/16085914.2019.1680340>.
- Jones RW, Hill JM, Coetzee JA, Avery TS, Weyl OLF, Hill MP. 2017. The abundance of an invasive freshwater snail *Tarebia granifera* (Lamarck, 1822) in the Nseleni River, South Africa. *African Journal of Aquatic Science* 42: 75–81. <https://doi.org/10.2989/16085914.2017.1298984>.
- Jones PE, Tummers JS, Galib SM, Woodford DJ, Hume JB, et al. 2021. The use of barriers to limit the spread of aquatic invasive animal species: a global review. *Frontiers in Ecology and Evolution* 9: 611631. <https://doi.org/10.3389/fevo.2021.611631>.
- Karatayev AY, Burlakova LE, Karatayev VA, Padilla DK. 2009. Introduction, distribution, spread, and impacts of exotic freshwater gastropods in Texas. *Hydrobiologia* 619: 181–194. <https://doi.org/10.1007/s10750-008-9639-y>.
- Keane RM, Crawley MJ. 2002. Exotic plant invasions and the enemy release hypothesis. *Trends in Ecology and Evolution* 17: 164–170. [https://doi.org/10.1016/S0169-5347\(02\)02499-0](https://doi.org/10.1016/S0169-5347(02)02499-0).
- Keller RP, Drake JM, Lodge DM. 2007. Fecundity as a basis for risk assessment of nonindigenous freshwater molluscs. *Conservation Biology* 21: 191–200. <https://doi.org/10.1111/j.1523-1739.2006.00563.x>.
- Kelly DW, Paterson RA, Townsend CR, Poulin R, Tompkins DM. 2009. Parasite spillback: a neglected concept in invasion ecology? *Ecology* 90: 2047–2056. <https://doi.org/10.1890/08-1085.1>.
- Kesner D, Kumschick S. 2018. Gastropods alien to South Africa cause severe environmental harm in their global alien ranges across habitats. *Ecology and Evolution* 8: 8273–8285. <https://doi.org/10.1002/ece3.4385>.
- Kibirige I, Perissinotto R. 2003. In situ feeding rates and grazing impact of zooplankton in a South African temporarily open estuary. *Marine Biology* 142: 357–367. <https://doi.org/10.1007/s00227-002-0963-x>.
- Krailas D, Namchote S, Rattanathai P. 2011. Human intestinal flukes *Haplorchis taichui* and *Haplorchis pumilio* in their intermediate hosts, freshwater snails of the families Thiariidae and Pachychilidae, in southern Thailand. *Zoosystematics and Evolution* 87: 349–360. <https://doi.org/10.1002/zoos.201100012>.
- Krailas D, Namchote S, Koonchornboon T, Dechruksa W, Boonmekam D. 2014. Trematodes obtained from the thiarid freshwater snail *Melanoides tuberculata* (Müller, 1774) as vector of human infections in Thailand. *Zoosystematics and Evolution* 90: 57–86. <https://doi.org/10.3897/zse.90.7306>.
- Lafferty KD, Kuris AM. 2009. Parasitic castration: the evolution and ecology of body snatchers. *Trends in Parasitology* 25: 564–572. <https://doi.org/10.1016/j.pt.2009.09.003>.
- Le TH, Nguyen KT, Nguyen NTB, Doan HTT, Blair D. 2017. The ribosomal transcription units of *Haplorchis pumilio* and *H. taichui* and the use of 28S rDNA sequences for phylogenetic identification of common heterophyids in Vietnam. *Parasites and Vectors* 10: 17. <https://doi.org/10.1186/s13071-017-1968-0>.
- Letelier VS, Ramos AM, Huaquín LG. 2007. Exotic freshwater mollusks in Chile. *Revista Mexicana De Biodiversidad* 78: 9S-13S. <https://doi.org/10.22201/ib.20078706e.2007.002.301>.
- Leuven RS, van der Velde G, Baijens I, Snijders J, van der Zwart C, et al. 2009. The River Rhine: a global highway for dispersal of aquatic invasive species. *Biological Invasions* 11: 1989-2008. <https://doi.org/10.1007/s10530-009-9491-7>.
- Lopes AS, Pulido-Murillo EA, Melo AL, Pinto HA. 2020. *Haplorchis pumilio* (Trematoda: Heterophyidae) as a new fish-borne zoonotic agent transmitted by *Melanoides tuberculata* (Mollusca: Thiariidae) in Brazil: a morphological and molecular study. *Infection, Genetics and Evolution* 85: 104495. <https://doi.org/10.1016/j.meegid.2020.104495>.
- Lopes-Lima M, Riccardi N, Urbanska M, Köhler F, Vinarski M, et al. 2021. Major shortfalls impairing knowledge and conservation of freshwater molluscs. *Hydrobiologia* 848: 2831–2867. <https://doi.org/10.1007/s10750-021-04622-w>.
- López MS, Coutinho R, Ferreira CE, Rilov G. 2010. Predator-prey interactions in a bioinvasion scenario: differential predation by native predators on two exotic rocky intertidal bivalves. *Marine Ecology Progress Series* 403: 101–112. <https://doi.org/10.3354/meps08409>.
- López-Altarrriba E, Garrido-Olvera L, Benavides-González F, Blanco-Martínez Z, Pérez-Castañeda R, et al. 2019. New records of invasive mollusks *Corbicula fluminea* (Müller, 1774), *Melanoides tuberculata* (Müller, 1774) and *Tarebia granifera* (Lamarck, 1816) in the Vicente Guerrero reservoir (NE Mexico). *BiolInvasions Records* 8: 640–652. <https://doi.org/10.3391/bir.2019.8.3.21>.
- López-López E, Sedeño-Díaz JE, Tapia-Vega P, Oliveros E. 2009. Invasive mollusks *Tarebia granifera* Lamarck, 1822 and *Corbicula fluminea* Müller, 1774 in the Tuxpam and Tecolutla rivers, Mexico: spatial and seasonal distribution patterns. *Aquatic Invasions* 4: 435–450. <https://doi.org/10.3391/ai.2009.4.3.2>.
- Lothe A, Zulu N, Øyhus AO, Kjetland EF, Taylor M. 2018. Treating schistosomiasis among South African high school pupils in an endemic area, a qualitative study. *BMC Infectious Diseases* 18: 239. <https://doi.org/10.1186/s12879-018-3102-0>.
- Louda SM, Stiling P. 2004. The double-edged sword of biological control in conservation and restoration. *Conservation Biology* 18: 50–53. <https://doi.org/10.1111/j.1523-1739.2004.00070.x>.
- Lovell SJ, Stone SF, Fernandez L. 2005. The economic impacts of aquatic invasive species: a review of the literature. *Agricultural and Resource Economics Review* 35: 195–208. <https://doi.org/10.1017/S1068280500010157>.
- Lowe S, Browne M, Boudjelas S, De Poorter M. 2000. 100 of the world's worst invasive alien species: a selection from the global invasive species database. The Invasive Species Specialist Group, Auckland. pp 1–12.
- Lymbery AJ, Morine M, Kanani HG, Beatty SJ, Morgan DL. 2014. Co-invaders: the effects of alien parasites on native hosts. *International Journal for Parasitology: Parasites and Wildlife* 3: 171–177. <https://doi.org/10.1016/j.ijppaw.2014.04.002>.
- Madhyastha A, Dutta J. 2012. *Tarebia granifera*. *The IUCN red list of threatened species 2012*: e.T165813A1102513. <https://doi.org/10.2305/IUCN.UK.2012-1.RLTS.T165813A1102513.en>.
- Madsen H, Frandsen F. 1989. The spread of freshwater snails including those of medical and veterinary importance. *Acta Tropica* 46: 139–146. [https://doi.org/10.1016/0001-706X\(89\)90030-2](https://doi.org/10.1016/0001-706X(89)90030-2).
- Majdi N, de Necker L, Fourie H, Loggenberg A, Netherlands EC, et al. 2022. Diversity and distribution of benthic invertebrates dwelling rivers of the Kruger National Park, South Africa. *Koedoe* 64: 1–18. <https://doi.org/10.4102/koedoe.v64i1.1702>.
- Makherana F, Cuthbert RN, Dondofema F, Wasserman RJ, Chauke GM, et al. 2022a. Distribution, drivers and population structure of the invasive alien snail *Tarebia granifera* in the Luvuvhu system, South Africa. *River Research and Applications* 38: 1362-1373. <https://doi.org/10.1002/rra.3937>.
- Makherana F, Cuthbert RN, Monaco CJ, Dondofema F, Wasserman RJ, et al. 2022b. Informing spread predictions of two alien snails using movement traits. *Science of the Total Environment* 811: 152364. <https://doi.org/10.1016/j.scitotenv.2021.152364>.
- Malatji MP, Myende N, Mukaratirwa S. 2021. Are freshwater snails, *Melanoides* sp. and invasive *Tarebia granifera* (Gastropoda:

- Thiaridae), suitable intermediate hosts for *Calicophoron microbothrium* (Trematoda: Paramphistomoidea)? An experimental study. *Frontiers in Veterinary Science* 8 <https://doi.org/10.3389/fvets.2021.705954>.
- Malherbe W. 2018. Ramsar wetlands in South Africa: historic and current aquatic research. *South African Journal of Science and Technology* 38: 1–13.
- Manh Hung NM. 2022. Trematode larval infections in snails collected from aquaculture ponds in Ha Noi and Yen Bai, Vietnam. *Academia Journal of Biology*, 44: 43–52. <https://doi.org/10.15625/2615-9023/16202>.
- Manh Hung NM, Stauffer JR, Madsen H. 2013. Prey species and size choice of the molluscivorous fish, black carp (*Mylopharyngodon piceus*). *Journal of Freshwater Ecology* 28: 547–560. <https://doi.org/10.1080/02705060.2013.800826>.
- March JG, Pringle CM. 2003. Food web structure and basal resource utilization along a tropical island stream continuum, Puerto Rico. *Biotropica* 35: 84–93. <https://doi.org/10.1111/j.1744-7429.2003.tb00265.x>.
- McCullough FS, Gayral PH, Duncan J, Christie JD. 1980. Molluscicides in schistosomiasis control. *Bulletin of the World Health Organization* 58: 681–689.
- McKoy SA, Hyslop EJ, Robinson RD. 2011. Associations between two trematode parasites, an ectosymbiotic annelid, and *Thiara (Tarebia) granifera* (Gastropoda) in Jamaica. *Journal of Parasitology* 97: 828–832. <https://doi.org/10.1645/GE-2494.1>.
- Meyerson LA, Carlton JT, Simberloff D, Lodge DM. 2019. The growing peril of biological invasions. *Frontiers in Ecology and the Environment* 17: 191. <https://doi.org/10.1002/fee.2036>.
- Mimori T, Hirai H, Kifune T, Inada K. 1982. *Philophthalmus* sp. (Trematoda) in a human eye. *American Journal of Tropical Medicine and Hygiene* 31: 859–861. <https://doi.org/10.4269/ajtmh.1982.31.859>.
- Miranda NA, Measey GJ, Peer N, Raw JL, Perissinotto R, Appleton CC. 2016. Shell crushing resistance of alien and native thiarid gastropods to predatory crabs in South Africa. *Aquatic Invasions* 11: 303–311. <https://doi.org/10.3391/ai.2016.11.3.08>.
- Miranda NA, Perissinotto R. 2012. Stable isotope evidence for dietary overlap between alien and native gastropods in coastal lakes of northern KwaZulu-Natal, South Africa. *PLoS One* 7: e31897. <https://doi.org/10.1371/journal.pone.0031897>.
- Miranda NA, Perissinotto R. 2014. Benthic assemblages of wetlands invaded by *Tarebia granifera* (Lamarck, 1822) (Caenogastropoda: Thiaridae) in the iSimangaliso Wetland Park, South Africa. *Molluscan Research* 34: 40–48. <https://doi.org/10.1080/13235818.2013.866177>.
- Miranda NA, Perissinotto R, Appleton CC. 2010. Salinity and temperature tolerance of the invasive freshwater gastropod *Tarebia granifera*. *South African Journal of Science* 106: 1–7. <https://doi.org/10.4102/sajs.v106i3/4.156>.
- Miranda NA, Perissinotto R, Appleton CC. 2011a. Feeding dynamics of the invasive gastropod *Tarebia granifera* in coastal and estuarine lakes of northern KwaZulu-Natal, South Africa. *Estuarine, Coastal and Shelf Science* 91: 442–449. <https://doi.org/10.1016/j.ecss.2010.11.007>.
- Miranda NA, Perissinotto R, Appleton CC. 2011b. Population structure of an invasive parthenogenetic gastropod in coastal lakes and estuaries of northern KwaZulu-Natal, South Africa. *PLoS One* 6: e24337. <https://doi.org/10.1371/journal.pone.0024337>.
- Miranda NA, Taylor SJ, Cwewe Y, Appleton CC. 2022. First record of the Asian freshwater snail *Sinotaia cf. quadrata* (Benson, 1842) from Africa. *BiolInvasions Record* 11: 676–685. <https://doi.org/10.3391/bir.2022.11.3.10>.
- Mitchell AJ, Salmon MJ, Huffman DG, Goodwin AE, Brandt TM. 2000. Prevalence and pathogenicity of a heterophyid trematode infecting the gills of an endangered fish, the fountain darter, in two central Texas spring-fed rivers. *Journal of Aquatic Animal Health* 12: 283–289. [https://doi.org/10.1577/1548-8667\(2000\)0122.0.CO;2](https://doi.org/10.1577/1548-8667(2000)0122.0.CO;2).
- Moodley D, Geerts S, Richardson DM, Wilson JR. 2013. Different traits determine introduction, naturalization and invasion success in woody plants: Proteaceae as a test case. *PLoS One* 8: e75078. <https://doi.org/10.1371/journal.pone.0075078>.
- Morrison MB, Butterfield BP, Ross SG, Collins C, Walde A, et al. 2019. The diet of the Eastern Musk Turtle (*Stemotherus odoratus*) as it pertains to invasive snail consumption in a freshwater spring habitat in Texas. *Herpetology Notes* 12:1133–1139.
- Moslemi JM, Snider SB, MacNeill K, Gilliam JF, Flecker AS. 2012. Impacts of an invasive snail (*Tarebia granifera*) on nutrient cycling in tropical streams: the role of riparian deforestation in Trinidad, West Indies. *PLoS One* 7: e38806.
- Moustafa AS, Badawy AMM, Hussien EHM. 2021. First record of an intermediate thiarid snail host; *Tarebia granifera* (Lamarck, 1822) of the lung trematod (*Paragonimus westermani* Kerbert) in Gena Province, Upper Egypt. *Egyptian Journal of Aquatic Biology and Fisheries* 25: 759–772. <https://doi.org/10.21608/ejabf.2021.181341>.
- Mujiono N, Isnainingsih NR. 2021. Diversity of land and freshwater snails (Mollusca: Gastropoda) of Laiwangi Wanggameti National Park, Sumba Island, Indonesia. *Journal Biodjati* 6: 162–173. <https://doi.org/10.15575/biodjati.v6i2.13521>.
- Murray HD. 1964. *Tarebia granifera* and *Melanoides tuberculata* in Texas. *Annual Report to the American Malacological Union* 53: 15–16.
- Nash MA, Hoffmann AA. 2012. Effective invertebrate pest management in dryland cropping in southern Australia: the challenge of marginality. *Crop Protection* 42: 289–304. <https://doi.org/10.1016/j.cropro.2012.06.017>.
- Nentwig W, Kühnel E, Bacher S. 2010. A generic impact-scoring system applied to alien mammals in Europe. *Conservation Biology* 24: 302–311. <https://doi.org/10.1111/j.1523-1739.2009.01289.x>.
- Nguyen HM, Van HH, Ho LT, Tatonova YV, Madsen H. 2021. Are *Melanoides tuberculata* and *Tarebia granifera* (Gastropoda, Thiaridae), suitable first intermediate hosts of *Clonorchis sinensis* in Vietnam? *PLoS Neglected Tropical Diseases* 15: e0009093. <https://doi.org/10.1371/journal.pntd.0009093>.
- Nollen PM, Murray HD. 1978. *Philophthalmus gralli*: identification, growth characteristics, and treatment of an oriental eyefluke of birds introduced into the continental United States. *The Journal of Parasitology* 64: 178–180. <https://doi.org/10.2307/3279646>.
- Novoa A, Kumschick S, Richardson DM, Rouget M, Wilson JR. 2016. Native range size and growth form in Cactaceae predict invasiveness and impact. *Neobiota* 30: 75–90. <https://doi.org/10.3897/neobiota.30.7253>.
- Nwoko OE, Kalinda C, Manyangadze T, Chimbari MJ. 2022. Species diversity, distribution, and abundance of freshwater snails in KwaZulu-Natal, South Africa. *Water* 14: 2267. <https://doi.org/10.3390/w14142267>.
- Oetama D, Purnama MF. 2022. Freshwater gastropod community in South Konawe District, Southeast Sulawesi, Indonesia. *Biodiversitas Journal of Biological Diversity* 23: 3364–3372. <https://dx.doi.org/https://doi.org/10.13057/biodiv/d230707>.
- Oleh M, Kyrlyo B, Olena K. 2018. Biological and bio-mechanical principles of the controlling molluscs *Melanoides tuberculata* (Müller 1774) and *Tarebia granifera* (Lamarck, 1822) in reservoirs of strategic importance. *World Scientific News* 99: 71–83.
- Oliveira JLD, Miyahira IC, Gonçalves ICB, Ximenes RF, Lacerda LEMD, et al. 2020. Non-marine invasive gastropods on Ilha Grande (Angra dos Reis, Rio de Janeiro, Brazil): distribution and implications for conservation. *Biota Neotropica* 20. <https://doi.org/10.1590/1676-0611-bn-2020-1060>.
- Paller VG, Samudio JA, Patagnan KL, Santamaria L, Tolentino AK, et al. 2021. *Paragonimus westermani* infection of freshwater crab

- Sundathelphusa philippina* and melaniid snails in Cadacan River in Irosin, Sorsogon, Philippines. *Journal of Parasitic Diseases* 45: 634–642. <https://doi.org/10.1007/s12639-020-01340-3>.
- Peer N, Perissinotto R, Miranda NA, Raw JL. 2015. A stable isotopic study of the diet of *Potamonautes sidneyi* (Brachyura: Potamonautidae) in two coastal lakes of the iSimangaliso Wetland Park, South Africa. *Water SA* 41: 549–558. <https://doi.org/10.4314/wsa.v41i4.15>.
- Perez JG, Vargas M, Malek EA. 1991. Displacement of *Biomphalaria glabrata* by *Thiara granifera* under natural conditions in the Dominican Republic. *Memórias do Instituto Oswaldo Cruz* 86: 341–347. <https://doi.org/10.1590/S0074-02761991000300008>.
- Perissinotto R. 1992. Mesozooplankton size-selectivity and grazing impact on the phytoplankton community of the Prince Edward Archipelago (Southern Ocean). *Marine Ecology Progress Series* 79: 243–258. <https://doi.org/10.3354/meps079243>.
- Phillips CT, Alexander ML, Howard R. 2010. Consumption of eggs of the endangered fountain darter (*Etheostoma fonticola*) by native and nonnative snails. *The Southwestern Naturalist* 55: 115–117. <https://doi.org/10.1894/JS-26.1>.
- Picker M, Griffiths C. 2011. *Alien and invasive animals - a South African perspective*. Cape Town: Struik Nature. 240 pp.
- Pillay D, Perissinotto R. 2008. The benthic macrofauna of the St. Lucia Estuary during the 2005 drought year. *Estuarine, Coastal and Shelf Science* 77: 35–46. <https://doi.org/10.1016/j.ecss.2007.09.004>.
- Pimentel D, Zuniga R, Morrison D. 2005. Update on the environmental and economic costs associated with alien-invasive species in the United States. *Ecological Economics* 52: 273–288. <https://doi.org/10.1016/j.ecolecon.2004.10.002>.
- Pinto HA, de Melo AL. 2011. A checklist of trematodes (Platyhelminthes) transmitted by *Melanoides tuberculata* (Mollusca: Thiaridae). *Zootaxa* 2799: 15–28. <https://doi.org/10.11646/zootaxa.2799.1.2>.
- Pointier JP. 1999. Invading freshwater gastropods: some conflicting aspects for public health. *Malacologia* 41: 403–411.
- Pointier JP, David P, Jarne P. 2010. The biological control of the snail hosts of schistosomes: the role of competitor snails and biological invasions. In: Toledo R, Fried B (ed) *Biomphalaria snails and larval trematodes*. Springer, New York, pp 215–238. https://doi.org/10.1007/978-1-4419-7028-2_9.
- Pointier JP, de Jong RJ, Tchuenté LT, Kristensen TK, Loker ES. 2005. A neotropical snail host of *Schistosoma mansoni* introduced into Africa and consequences for the schistosomiasis transmission: *Biomphalaria tenagophila* in Kinshasa (Democratic Republic of Congo). *Acta Tropica* 93: 191–199. <https://doi.org/10.1016/j.actatropica.2004.11.003>.
- Pointier JP, Facon B, Jarne P, David P. 2003. Thiarid snails, invading gastropods of tropical freshwater habitats. *Xenophora* 104: 14–20.
- Pointier JP, Giboda M. 1999. The case for biological control of snail intermediate hosts of *Schistosoma mansoni*. *Parasitology Today* 15: 395–397. [https://doi.org/10.1016/S0169-4758\(99\)01517-3](https://doi.org/10.1016/S0169-4758(99)01517-3).
- Pointier JP, Jourdan J. 2000. Biological control of the snail hosts of schistosomiasis in areas of low transmission: the example of the Caribbean area. *Acta Tropica* 77: 53–60. [https://doi.org/10.1016/S0001-706X\(00\)00123-6](https://doi.org/10.1016/S0001-706X(00)00123-6).
- Pointier JP, Samadi S, Jarne P, Delay B. 1998. Introduction and spread of *Thiara granifera* (Lamarck, 1822) in Martinique, French West Indies. *Biodiversity and Conservation* 7: 1277–1290. <https://doi.org/10.1023/A:1008887531605>.
- Porter A. 1940. The larval trematodes found in certain South African molluscs with special reference to schistosomiasis. *JAMA* 114: 682–683. <https://doi.org/10.1001/jama.1940.02810080054030>.
- Prenter J, MacNeil C, Dick JT, Dunn AM. 2004. Roles of parasites in animal invasions. *Trends in Ecology and Evolution* 19: 385–390. <https://doi.org/10.1016/j.tree.2004.05.002>.
- Prentice MA. 1983. Displacement of *Biomphalaria glabrata* by the snail *Thiara granifera* in field habitats in St. Lucia, West Indies. *Annals of Tropical Medicine and Parasitology* 77: 51–59. <https://doi.org/10.1080/00034983.1983.11811672>.
- Prentis PJ, Wilson JR, Dormontt EE, Richardson DM, Lowe AJ. 2008. Adaptive evolution in invasive species. *Trends in Plant Science* 13: 288–294. <https://doi.org/10.1016/j.tplants.2008.03.004>.
- Preston DL, Crone ER, Miller-ter Kuile A, Lewis CD, Sauer EL, Trovillion DC. 2022. Non-native freshwater snails: a global synthesis of invasion status, mechanisms of introduction, and interactions with natural enemies. *Freshwater Biology* 67: 227–239. <https://doi.org/10.1111/fwb.13848>.
- Pulido-Murillo EA, Furtado LfV, Melo AL, Rabelo ÉM, Pinto HA. 2018. Fishborne zoonotic trematodes transmitted by *Melanoides tuberculata* snails, Peru. *Emerging Infectious Diseases* 24: 606–608. <https://doi.org/10.3201/eid2403.172056>.
- Purnama MF, Sari SF, Admaja AK. 2020. Spatial distribution of invasive alien species *Tarebia granifera* in Southeast Sulawesi, Indonesia. *AACL Bioflux* 13: 1355–1365.
- Purnama MF, Sari SF, Oetama D, Sirza LO, Admaja AK, et al. 2021. Specific characteristics of niche and spatial distribution of invasive alien species *Tarebia granifera* in Buton Island, Indonesia. *Aquaculture, Aquarium, Conservation and Legislation* 14: 233–248.
- Purnama MF, Sari SF, Salwiyah S, Haslianti H, Abdullah A, et al. 2022. Diversity report of freshwater gastropods in Buton Island, Indonesia. *Biodiversitas Journal of Biological Diversity* 23: 1938–1949. <https://doi.org/10.13057/biodiv/d230428>.
- Rajapakse RDK, Wijerathne KMTN, Wijesundera M. 2009. Ocular infection with an avian trematode (*Philophthalmus* sp). *Ceylon Medical Journal* 54: 128–129. <https://doi.org/10.4038/cmj.v54i4.1454>.
- Raphahlelo ME, Addo-Bediako A, Luus-Powell WJ. 2022. Distribution and diversity of benthic macroinvertebrates in the Mhlapitsi River, South Africa. *Journal of Freshwater Ecology* 37: 145–160. <https://doi.org/10.1080/02705060.2021.2023054>.
- Raw JL, Miranda NA, Perissinotto R. 2013. Chemical cues released by an alien invasive aquatic gastropod drive its invasion success. *PLoS One* 8: e64071. <https://doi.org/10.1371/journal.pone.0064071>.
- Raw JL, Miranda NA, Perissinotto R. 2015. Chemical cues released by heterospecific competitors: behavioural responses of native and alien invasive aquatic gastropods. *Aquatic Sciences* 77: 655–666. <https://doi.org/10.1007/s00027-015-0409-4>.
- Reid AJ, Carlson AK, Creed IF, Eliason EJ, Gell PA, et al. 2019. Emerging threats and persistent conservation challenges for freshwater biodiversity. *Biological Reviews* 94: 849–873. <https://doi.org/10.1111/brv.12480>.
- Ricciardi A, MacIsaac HJ. 2000. Recent mass invasion of the North American Great Lakes by Ponto-Caspian species. *Trends in Ecology and Evolution* 15: 62–65. [https://doi.org/10.1016/S0169-5347\(99\)01745-0](https://doi.org/10.1016/S0169-5347(99)01745-0).
- Richardson DM, Pyšek P. 2006. Plant invasions: merging the concepts of species invasiveness and community invasibility. *Progress in Physical Geography* 30: 409–431. <https://doi.org/10.1191/0309133306pp490pr>.
- Rico-Sánchez AE, Sundermann A, López-López E, Torres-Olvera MJ, Mueller SA, Haubrock PJ. 2020. Biological diversity in protected areas: not yet known but already threatened. *Global Ecology and Conservation* 22: e01006. <https://doi.org/10.1016/j.gecco.2020.e01006>.
- Rumi A, Sánchez J, Ferrando NS. 2010. *Theba pisana* (Müller, 1774) (Gastropoda, Helicidae) and other alien land molluscs species in Argentina. *Biological Invasions* 12: 2985–2990. <https://doi.org/10.1007/s10530-010-9715-x>.

- Scharler UM, Lechman K, Radebe T, Jerling HL. 2020. Effects of prolonged mouth closure in a temporarily open/closed estuary: a summary of the responses of invertebrate communities in the uMdloti Estuary, South Africa. *African Journal of Aquatic Science* 45: 121–130. <https://doi.org/10.2989/16085914.2019.1689911>.
- Schiffbauer J. 2009. *Anentome helena* (Meder, 1847) - the assassin snail. *Arthropoda* 17: 60–65.
- Shinen JS, Morgan SG, Chan AL. 2009. Invasion resistance on rocky shores: direct and indirect effects of three native predators on an exotic and a native prey species. *Marine Ecology Progress Series* 378: 47–54. <https://doi.org/10.3354/meps07870>.
- Simberloff D, Stiling P. 1996. How risky is biological control? *Ecology* 77: 1965–1974. <https://doi.org/10.2307/2265693>.
- Simone LRLD. 2006. *Land and freshwater molluscs of Brazil: an illustrated inventory on the Brazilian malacofauna, including neighbour regions of the South America, respect to the terrestrial and freshwater ecosystems*. 18th ed. EGB, FAPESP, São Paulo, Brazil.
- Sirza LJ, Purnama MF, Anwa K. 2020. Invasive status of *Tarebia granifera* based on density of population in river of Gunung Sejuk Village, South Buton Regency. *Aquasains* 9: 875–880. <https://doi.org/10.23960/aqs.v9i1.p875-880>.
- Skirnisson K, Galaktionov KV, Kozminsky EV. 2004. Factors influencing the distribution of digenetic trematode infections in a mudsnail (*Hydrobia ventrosa*) population inhabiting salt marsh ponds in Iceland. *Journal of Parasitology* 90: 50–59. <https://doi.org/10.1645/GE-118R>.
- Snider SB, Gilliam JF. 2008. Movement ecology: size-specific behavioural response of an invasive snail to food availability. *Ecology* 89: 1961–1971. <https://doi.org/10.1890/07-0715.1>.
- Snoeks J, Harrison IJ, Stiassny MLJ. 2011. The status and distribution of freshwater fishes. In: Darwall W, Smith K, Allen D, Holland R, Harrison I, Brooks E (ed) *The diversity of life in African freshwaters: underwater, under threat. An analysis of the status and distribution of freshwater species throughout mainland Africa*. IUCN, Gland, Switzerland. pp 42–91.
- Sodeman WA. 1991. *Thiara (Tarebia) granifera* (Lamarck): an agent for biological control of *Biomphalaria*. In: National Research Council (ed) *Aquaculture and schistosomiasis, proceedings of the Network Meeting, Manila, Philippines*. National Academy Press, Washington DC, USA. pp 6–10.
- Sommerville C. 1982. The pathology of *Haplorchis pumilio* (Looss, 1896) infections in cultured tilapias. *Journal of Fish Diseases* 5: 243–250. <https://doi.org/10.1111/j.1365-2761.1982.tb00479.x>.
- Sorensen RE, Minchella DJ. 2001. Snail-trematode life history interactions: past trends and future directions. *Parasitology* 123: S3–S18. <https://doi.org/10.1017/S0031182001007843>.
- Strayer DL. 2010. Alien species in fresh waters: ecological effects, interactions with other stressors, and prospects for the future. *Freshwater Biology* 55: 152–174. <https://doi.org/10.1111/j.1365-2427.2009.02380.x>.
- Sullivan KT, Littrell BM. 2020. Spatiotemporal variation in *Elimia comalensis* (Gastropoda: Pleuroceridae) density and interspecific associations with exotic thiarid snails in the upper San Marcos River, Texas. *Malacologia* 63: 1–9. <https://doi.org/10.4002/040.063.0101>.
- Thomaz SM, Kovalenko KE, Havel JE, Kats LB. 2015. Aquatic invasive species: general trends in the literature and introduction to the special issue. *Hydrobiologia* 746: 1–12. <https://doi.org/10.1007/s10750-014-2150-8>.
- Tilman D. 1999. The ecological consequences of changes in biodiversity: a search for general principles. *Ecology* 80: 1455–1474. <https://doi.org/10.2307/176540>.
- Tolley-Jordan LR, Chadwick MA. 2019. Effects of parasite infection and host body size on habitat associations of invasive aquatic snails: implications for environmental monitoring. *Journal of Aquatic Animal Health* 31: 121–128. <https://doi.org/10.1002/aah.10059>.
- Tolley-Jordan LR, Owen JM. 2008. Habitat influences snail community structure and trematode infection levels in a spring-fed river, Texas, USA. *Hydrobiologia* 600: 29–40. <https://doi.org/10.1007/s10750-007-9173-3>.
- Torchin ME, Lafferty KD, Dobson AP, McKenzie VJ, Kuris AM. 2003. Introduced species and their missing parasites. *Nature* 421: 628–630. <https://doi.org/10.1038/nature01346>.
- Ukong S, Krailas D, Dangprasert T, Changgarm P. 2007. Studies on the morphology of cercariae obtained from freshwater snails at Erawan Waterfall, Erawan National Park, Thailand. *Southeast Asian Journal of Tropical Medicine and Public Health* 38: 302–312.
- Umadevi K, Madhavi R. 2006. The life cycle of *Haplorchis pumilio* (Trematoda: Heterophyidae) from the Indian region. *Journal of Helminthology* 80: 327–332. <https://doi.org/10.1017/JOH2006359>.
- Van Bocxlaer B, Clewing C, Etimosundja JPM, Kankonda A, Ndeo OW, Albrecht C. 2015. Recurrent camouflaged invasions and dispersal of an Asian freshwater gastropod in tropical Africa. *BMC Evolutionary Biology* 15: 33. <https://doi.org/10.1186/s12862-015-0296-2>.
- Van Leeuwen CH, Huig N, Van der Velde G, Van Alen TA, Wagemaker CA, et al. 2013. How did this snail get here? Several dispersal vectors inferred for an aquatic invasive species. *Freshwater Biology* 58: 88–99. <https://doi.org/10.1111/fwb.12041>.
- Van Oosterhout C, Mohammed R, Xavier R, Stephenson J, Archard GA, et al. 2013. Invasive freshwater snails provide resource for native marine hermit crabs. *Aquatic Invasions* 8: 185–191. <https://doi.org/10.3391/ai.2013.8.2.06>.
- Vander Zanden MJ, Casselman JM, Rasmussen JB. 1999. Stable isotope evidence for the food web consequences of species invasions in lakes. *Nature* 401: 464–467. <https://doi.org/10.1038/46762>.
- Vanni MJ. 2002. Nutrient cycling by animals in freshwater ecosystems. *Annual Review of Ecology and Systematics* 33: 341–370. <https://doi.org/10.1146/annurev.ecolsys.33.010802.150519>.
- Vanschoenwinkel B, Waterkeyn A, Vandecaetsbeek T, Pineau O, Grillas P, Brendonck LUC. 2008. Dispersal of freshwater invertebrates by large terrestrial mammals: a case study with wild boar (*Sus scrofa*) in Mediterranean wetlands. *Freshwater Biology* 53: 2264–2273. <https://doi.org/10.1111/j.1365-2427.2008.02071.x>.
- Vanschoenwinkel B, Waterkeyn A, Nihwatiwa T, Pinceel T, Spooren E, Geerts A, Clegg B, Brendonck LUC. 2011. Passive external transport of freshwater invertebrates by elephant and other mud-wallowing mammals in an African savannah habitat. *Freshwater Biology* 56: 1606–1619. <https://doi.org/10.1111/j.1365-2427.2011.02600.x>.
- Veeravechskij N, Namchote S, Neiber MT, Glaubrecht M, Krailas D. 2018a. Exploring the evolutionary potential of parasites: larval stages of pathogen digenetic trematodes in their thiarid snail host *Tarebia granifera* in Thailand. *Zoosystematics and Evolution* 94: 425–460. <https://doi.org/10.3897/zse.94.28793>.
- Veeravechskij N, Krailas D, Namchote S, Wiggnering B, Neiber MT, Glaubrecht M. (2018b) Molecular phylogeography and reproductive biology of the freshwater snail *Tarebia granifera* in Thailand and Timor (Cerithioidea, Thiaridae): morphological disparity versus genetic diversity. *Zoosystematics and Evolution* 94: 461–493. <https://doi.org/10.3897/zse.94.28981>.
- Walker JC. 1978. The finding of *Biomphalaria straminea* amongst fish imported into Australia. World Health Organization (WHO) document. WHO/SCHISTO/78: 46. WHO, Geneva, Switzerland.

- Wang JJ, Chung LY, Lee JD, Chang EE, Chen ER, et al. 2002. *Haplorchis* infections in intermediate hosts from a clonorchiasis endemic area in Meinung, Taiwan, Republic of China. *Journal of Helminthology* 76: 185–188. <https://doi.org/10.1079/JOH2002114>.
- Waterkeyn A, Vanschoenwinkel B, Elsen S, Anton-Pardo M, Grillas Brendonck L. 2010. Unintentional dispersal of aquatic invertebrates via footwear and motor vehicles in a Mediterranean wetland area. *Aquatic Conservation: Marine and Freshwater Ecosystems* 20: 580–587. <https://doi.org/10.1002/aqc.1122>.
- Weyl OLF, Ellender BR, Wassermann RJ, Truter M, Dalu T, et al. 2020. Alien freshwater fauna in South Africa. In: van Wilgen BW, Measey J, Richardson DM, Wilson JR, Zengeya TA (ed) *Biological Invasions in South Africa*. Springer Nature, Cham, Switzerland. pp 153–183. https://doi.org/10.1007/978-3-030-32394-3_6.
- Whitfield AK, Adams JB, Harrison TD, Lamberth SJ, Lemley DA, et al. 2021. Possible impacts of non-native plant, pathogen, invertebrate and fish taxa on the indigenous ichthyofauna in South African estuaries: a preliminary review. *Biological Invasions* 23: 2729–2747. <https://doi.org/10.1007/s10530-021-02541-4>.
- Wollerman L, Duva M, Ferrier MD. 2003. Responses of *Littoraria irrorata* (Mollusca: Gastropoda) to water-borne chemicals: a comparison of chemical sources and orientation mechanisms. *Marine and Freshwater Behaviour and Physiology* 36: 129–142. <https://doi.org/10.1080/10236240310001603756>.
- Wolmarans CT, de Kock KN. 2006. The current status of freshwater molluscs in the Kruger National Park. *Koedoe* 49: 39–44. <https://doi.org/10.4102/koedoe.v49i2.122>.
- WWF (World Wide Fund for Nature). 2020. Living Planet Report 2020 - Bending the curve of biodiversity loss. In: Almond REA, Grooten M, Petersen T (ed). *World Wildlife Fund for Nature* (WWF), Gland, Switzerland. pp 1–162.
- Yakovenko V, Fedonenko O, Klimenko O, Petrovsky O. 2018. Biological control of the invasive snail species *Melanooides tuberculata* and *Tarebia granifera* in Zaporizka Nuclear Power Plant cooling pond. *Ukrainian Journal of Ecology* 8: 975–982. https://doi.org/10.15421/2018_301.
- Yesipova N, Marenkov O, Sharamok T, Nesterenko O, Kurchenko V. 2022. Development of the regulation of hydrobiological monitoring in circulation cooling system of the Zaporizhzhia Nuclear Power Plant. *Eastern-European Journal of Enterprise Technologies* 2: 6–17. <https://doi.org/10.15587/1729-4061.2022.255537>.
- Yin N, Zhao S, Huang XC, Ouyang S, Wu XP. 2022. Complete mitochondrial genome of the freshwater snail *Tarebia granifera* (Lamarck, 1816) (Gastropoda: Cerithioidea: Thiaridae). *Mitochondrial DNA Part B* 7: 259–261. <https://doi.org/10.1080/23802359.2022.2026832>.
- Zengeya TA, Kumschick S, Weyl OLF, van Wilgen BW. 202.) An evaluation of the impacts of alien species on biodiversity in South Africa using different assessment methods. In: van Wilgen BW, Measey J, Richardson DM, Wilson JR, Zengeya TA (ed) *Biological Invasions in South Africa*. Springer Nature, Cham, Switzerland. pp 489–512. https://doi.org/10.1007/978-3-030-32394-3_17.