

Amorphism and polymorphism of azithromycin

ROELF WILLEM ODENDAAL
(M.Sc, B. PHARM.)

Thesis submitted in fulfilment of the requirements for the degree

Philosophiae Doctor (PHARMACEUTICS)

at the

POTCHEFSTROOM CAMPUS OF THE NORTH-WEST UNIVERSITY

Supervisor: Prof. W. Liebenberg

Co-supervisor: Dr. M.E. Aucamp



2 KOR. 12:9

**“My genade is vir jou genoeg.
My krag kom juis tot volle werking
wanneer jy swak is.”**



Dedicated to my Heavenly Father

AND

my parents, W.T. and Antoinette Odendaal

AND

my late grandparents,

Roelf & Petronella Odendaal

Hans & Lea van Dyk

ACKNOWLEDGEMENTS

I would like to express my deepest and greatest appreciation to my **Heavenly Father**. This honour came to me by the grace of God. His mercy, love and presence were, and will always be, the foundation of my work, life, and all the success I achieve.

My parents, **W.T. and Antoinette Odendaal**. No words can describe the moral and emotional support that you gave me. I know that things got tough at times and that I wasn't the easiest to cope with, but you were the ones who continued to motivate me and you made me realise the true value of moral support.

My sister and brother-in-law, **Antonelle and J-P Combrink**. Your support is much appreciated.

My uncle and aunt, **Carl and Annelie Wilms**. Thank you for always taking an interest in my work and project; and for all the encouragement and prayers.

My supervisor, **Prof. W. Liebenberg**. All the advice, support and encouragement were of great value to me.

My co-supervisor, **Dr. M.E. Aucamp**. Thank you for your assistance and time in making this project a success.

A special thanks to **Prof. Jeanetta du Plessis**. I am very grateful for your endless support and your "open-door" policy throughout the three years of this study. You always believed in me and always had the right words to encourage me.

To **Prof. J.C. Wessels**. Thank you for all the endless support and your willingness to always be there as a colleague and friend.

To **DM Kisch Inc., Rudi van der Walt and Johann Coetzee**. Thank you for your effort and assistance during the PCT application process and every step of the way since then. It is much appreciated.

To **Belinda Venter**. Thank you for your help in analysing samples on the XRPD for this study.

To **Dr. L. Tiedt**. Thank you for your insightful eye in taking great SEM photos during this study.

To **Dr. Jan Steenekamp**. Thank you for the guidance during the formulation process and helping me to formulate tablets.

To **Prof. J.H. Hamman**. Thank you for your guidance and assistance during the permeability study.

To my **Ermelo Hospital Pharmacy family**. All your prayers can never be described in real value. Thank you so much for your support and motivation.

To all my **family and other friends**. Thank you for the support and inspiration that you gave me. You made a big difference in helping me achieve this tremendous honour.

To the **NRF**. Thank you for the financial and research support throughout the three years.

TABLE OF CONTENTS

TABLE OF CONTENTS	i
LIST OF TABLES AND FIGURES	x
LIST OF ABBREVIATIONS	xx
LIST OF EQUATIONS	xxiv
ABSTRACT	xxvi
UITTREKSEL	xxviii
STUDY OBJECTIVES	xxx

CHAPTER 1: SOLID STATE OF PHARMACEUTICAL COMPOUNDS

1.1	Introduction	1
1.2	Polymorphism	1
1.2.1	Defining polymorphism	1
1.2.2	Importance of polymorphism.....	2
1.3	Solid state	3
1.3.1	Classification of solid forms	4
1.4	Crystalline solid.....	5
1.4.1	Polymorphs.....	6
1.4.2	Solvates	7
1.4.2.1	Method for preparing solvates	7
1.4.3	Hydrates	8
1.4.3.1	Classification of hydrates based on their crystal structure	8
1.4.3.2	Conditions for hydrate formation.....	10
1.4.3.3	Method for preparing hydrates.....	10

1.4.3.4	Stability of hydrates	11
1.4.3.5	Instability of hydrates due to phase transformations.....	12
1.5	Non-crystalline / amorphous solids	14
1.5.1	Definition of the amorphous state	14
1.5.2	Structural aspects of amorphous solids	14
1.5.3	Amorphous solids	15
1.5.4	Methods for preparing amorphous solids.....	16
1.5.5	Properties of an amorphous glass	16
1.5.5.1	High potential energy	17
1.5.5.2	Structural heterogeneity.....	18
1.5.5.3	Structural relaxation.....	19
1.5.6	Stability of amorphous solids	20
1.5.6.1	Effect of temperature on stability	20
1.5.6.2	Effect of moisture on stability.....	20
1.5.7	Solubility of amorphous solids	22
1.5.7.1	Equilibrium (intrinsic) solubility <i>versus</i> metastable solubility	23
1.5.8	Effect of amorphism on the dissolution rate.....	24
1.6	Conclusion	25
1.7	References	27

CHAPTER 2: SOLID STATE CHARACTERISATION

2.1	Introduction.....	31
2.2	Materials	31
2.3	Methods for solid state characterisation	32
2.3.1	Infrared spectroscopy (IR)	32
2.3.1.1	Fourier transform infrared spectroscopy (FTIR).....	33
2.3.1.2	Diffuse reflectance infrared Fourier transform spectroscopy (DRIFTS)	33

2.3.2	X-ray powder diffraction (XRPD).....	34
2.3.3	Thermal methods.....	36
2.3.3.1	Differential scanning calorimetry (DSC).....	37
2.3.3.2	Thermogravimetric analysis (TGA)	38
2.3.4	Microscopy.....	38
2.3.4.1	Hot stage microscopy (HSM).....	39
2.3.4.2	Scanning electron microscopy (SEM).....	40
2.3.5	Microcalorimetry	41
2.3.6	Moisture sorption analysis	42
2.3.7	Karl Fischer titration (KFT).....	43
2.3.7.1	Volumetric KFT	43
2.3.7.2	Coulometric KFT	44
2.4	High-performance liquid chromatography (HPLC)	44
2.4.1	HPLC method development for the identification and quantification of azithromycin.....	45
2.4.2	Experimental.....	46
2.4.2.1	Materials	46
2.4.2.2	Instrumentation and software.....	46
2.4.2.3	Chromatographic conditions	46
2.4.3	Preparation of solutions	47
2.4.3.1	Calibration standard solutions.....	47
2.4.3.2	Preparation of test solutions	47
2.4.3.3	Preparation of test solutions in solubility media	48
2.4.4	Method validation.....	48
2.4.5	Results and discussion	49
2.4.5.1	Method optimisation.....	49
2.4.6	Method validation.....	50

2.4.6.1	Linearity and range	50
2.4.6.2	Accuracy	50
2.4.6.3	Specificity.....	51
2.4.6.4	Precision	52
2.4.6.5	Limit of detection and quantitation	53
2.4.7	Application of HPLC analyses in solubility and dissolution studies	54
2.4.8	Conclusion	56
2.5	Solubility	56
2.5.1	General aspects of solubility	56
2.5.1.1	Method for solubility determination	56
2.6	Stability	57
2.6.1	Method for accelerated stability study.....	58
2.6.2	Method for determining the acid stability at pH 1.2	58
2.7	Dissolution study.....	58
2.7.1	Materials and methods	58
2.8	Conclusion	59
2.9	References	61

CHAPTER 3: AZITHROMYCIN: A MACROLIDE ANTIMICROBIAL

3.1	Introduction and background	65
3.2	Classification of macrolides	65
3.3	Azithromycin	66
3.3.1	Structural aspects of azithromycin	66
3.3.2	Spectrum of pharmacological activity	68
3.3.3	Mode of action	69
3.3.4	Indications of azithromycin.....	69
3.3.5	Physical properties of azithromycin	70

3.3.5.1	Appearance	70
3.3.5.2	Solubility	70
3.3.5.3	Water content	70
3.3.5.4	Theoretical weight loss with drying	71
3.3.5.5	Melting point	71
3.3.5.6	Stability in acidic environment.....	71
3.3.5.7	Iso-electric point of azithromycin.....	72
3.3.6	Pharmacokinetic properties	72
3.3.6.1	Absorption and metabolism	72
3.3.6.2	Distribution.....	73
3.3.6.3	Elimination	74
3.3.7	Drug interactions.....	74
3.3.8	Adverse effects	74
3.3.9	Toxicity.....	75
3.3.10	Safety	75
3.3.11	Cost of treatment	75
3.3.12	Bacterial resistance to azithromycin and other macrolides	75
3.4	Conclusion	77
3.5	References	78

CHAPTER 4: POLYMORPHISM OF AZITHROMYCIN

4.1	Introduction.....	81
4.2	Recrystallisation method.....	82
4.3	Results.....	82
4.4	Discussion	83
4.5	Anhydrous azithromycin prepared from azithromycin dihydrate	83
4.5.1	Anhydrous azithromycin prepared <i>via</i> dry heat.....	83

4.5.2	Anhydrous azithromycin dried using the DSC and TGA apparatuses.....	87
4.5.3	Anhydrous azithromycin prepared with isopropanol	88
4.6	Discussion	93
4.7	Conclusion	94
4.8	References	95

CHAPTER 5: AZITHROMYCIN GLASS

5.1	Introduction	97
5.2	Method for the preparation of azithromycin glass (AZM-G)	98
5.3	Solid state characterisation of azithromycin glass (AZM-G).....	98
5.3.1	Fourier transform infrared spectroscopy (FTIR).....	98
5.3.2	X-ray powder diffraction (XRPD).....	100
5.3.3	Differential scanning calorimetry (DSC).....	102
5.3.4	Thermogravimetric analysis (TGA)	104
5.3.5	Karl Fischer titration (KFT).....	106
5.3.6	Microscopy.....	106
5.4	Solubility of AZM-G	111
5.4.1	Stability and solubility of azithromycin glass in 0.1 M HCl	111
5.4.1.1	Results of acid stability	111
5.4.1.2	Solubility in pH 1.2	113
5.4.2	Solubility in acetate buffer (pH 4.5)	114
5.4.3	Solubility in phosphate buffer (pH 6.8).....	115
5.4.4	Solubility in water	116
5.5	Stability of AZM-G.....	118
5.5.1	Results.....	118
5.5.2	Stability study.....	122
5.5.2.1	DSC analyses of AZM-G from weeks 0 – 4	123

5.5.2.2	TGA results for AZM-G during stability study	127
5.5.2.3	KFT results	129
5.5.2.4	XRPD results	130
5.5.2.5	Fourier transform infrared spectroscopy (FTIR).....	132
5.5.2.6	Microscopy.....	134
5.5.3	Stability of AZM-G in water	136
5.5.4	Stability of AZM-G coupled with an increased water fraction	138
5.6	Conclusion	142
5.7	References	143

CHAPTER 6: PRODUCT DEVELOPMENT

6.1	Introduction	146
6.2	Solid dosage form design	146
6.2.1	Particle size, shape and density	148
6.2.2	Characterisation of powder flow properties.....	148
6.2.2.1	Angle of repose.....	149
6.2.2.2	Compressibility	149
6.2.2.3	Flow through a hopper.....	151
6.2.3	Methods for improving the flow of AZM-G powder.....	151
6.2.3.1	Particle size	151
6.2.3.2	Excipients	153
6.2.4	Granulation methods	155
6.2.4.1	Dry granulation	155
6.2.4.2	Wet granulation.....	156
6.3	Direct compression of AZM-G powder.....	156
6.4	Wet granulation of AZM-G powder	157
6.4.1	Bulk production of AZM-G tablets (Formulation 5).....	158

6.4.2.	Bulk production of AZM-DH tablets.....	162
6.5	Dissolution study of AZM-DH and AZM-G in different media	164
6.5.1	Results.....	164
6.5.1.1	Dissolution profiles in pH 4.5 acetate buffer.....	164
6.5.1.2	Dissolution profiles in pH 6.8 phosphate buffer.....	165
6.5.1.3	Dissolution profiles in water	166
6.5.1.4	Dissolution profiles in pH 1.2 HCl	167
6.5.2	Discussion	167
6.6	Stability of AZM-DH and AZM-G tablets	168
6.6.1	Materials and methods	168
6.6.2	Results.....	169
6.6.2.1	Fourier transform infrared spectroscopy (FTIR).....	169
6.6.2.2	Dissolution	171
6.6.3	Discussion	172
6.7	Conclusion	173
6.8	References	175

CHAPTER 7: PERMEABILITY

7.1	Introduction.....	177
7.2	Membrane permeability	178
7.2.1	Transcellular and paracellular passive diffusion	179
7.2.2	Endocytosis	180
7.2.3	Active transport.....	180
7.3	Effect of membrane permeability on bioavailability	181
7.4	Materials and methods	181
7.4.1	Preparation of azithromycin solutions	182
7.4.2	Preparation of tissue for <i>in vitro</i> transport.....	183

7.5	Data analysis	187
7.6	Results.....	187
7.6.1	Azithromycin transport at pH 7.2.....	187
7.6.2	Azithromycin transport at pH 6.8.....	189
7.6.3	Azithromycin transport at pH 4.5.....	191
7.6.4	Azithromycin transport in water.....	192
7.7	Discussion	193
7.8	Conclusion	195
7.9	References	197

CHAPTER 8: CONCLUSION 199

ANNEXURE A..... 202

ANNEXURE B..... 206

ANNEXURE C..... 208

LIST OF TABLES AND FIGURES

CHAPTER 1: SOLID STATE

Figure 1.1	Diagrammatic representation of the classification of solid state forms, according to Cui (2007:5).....	5
-------------------	--	---

CHAPTER 2: SOLID STATE CHARACTERISATION

Table 2.1	XRPD settings for the measurement of azithromycin samples	36
------------------	---	----

Table 2.2	Equations for regression obtained during method validation to compare linearity at 205 nm and 210 nm	50
------------------	--	----

Table 2.3	Accuracy as a result of azithromycin recovered from the 100 % calibration solution	51
------------------	--	----

Table 2.4	Summary of intermediate validation results at 205 nm and 210 nm, as obtained by the second analyst	52
------------------	--	----

Table 2.5	DL and QL values achieved by the HPLC method at 205 nm and 210 nm	53
------------------	---	----

Figure 2.1	HPLC chromatograms of AZM-DH reference material dissolved in mobile phase	47
-------------------	---	----

Figure 2.2	HPLC chromatogram of AZM-DH dissolved in mobile phase (1 mg/mL) at 205 nm. Mobile phase: phosphate buffer adjusted with phosphoric acid (pH 3.0)/acetonitrile (600/400), Flow rate 1.0 mL/min, Injection volume 15 µL. UV detection 205 nm	49
-------------------	--	----

Figure 2.3	HPLC Chromatogram of AZM-DH dissolved in mobile phase (1 mg/mL), at 205 nm. Mobile phase: phosphate buffer adjusted with 1.0 M sodium hydroxide (pH 8.4)/acetonitrile (500/500), Flow rate 1.0 mL/min, Injection volume 15 µL. UV detection 205 nm	50
-------------------	--	----

Figure 2.4:	HPLC chromatogram of AZM-DH tablet (5 mg/mL) dissolved in mobile phase at 205 nm and 210 nm. Mobile phase: phosphate buffer (adjusted with 1.0 M sodium hydroxide to pH 6.0)/acetonitrile (700/300), Flow rate 1.0 mL/min, Injection volume 15 μ L. UV detection 205 nm and 210 nm	52
Figure 2.5	HPLC chromatogram of AZM-DH dissolved in water (0.1 mg/mL) at 205 nm. Mobile phase: phosphate buffer (adjusted with 1.0 M sodium hydroxide to pH 6.0)/acetonitrile (700/300), Flow rate 1.0 mL/min, Injection volume 15 μ L. UV detection 205 nm	54
Figure 2.6:	HPLC chromatogram of AZM-DH dissolved in pH 4.5 acetate buffer (1.0 mg/mL) at 205 nm. Mobile phase: phosphate buffer (adjusted with 1.0 M sodium hydroxide to pH 6.0)/acetonitrile (700/300), Flow rate 1.0 mL/min, Injection volume 15 μ L. UV detection 205 nm	54
Figure 2.7:	HPLC chromatogram of AZM-DH dissolved in pH 6.8 phosphate buffer (0.5 mg/mL) at 205 nm. Mobile phase: phosphate buffer (adjusted with 1.0 M sodium hydroxide to pH 6.0)/acetonitrile (700/300), Flow rate 1.0 mL/min, Injection volume 15 μ L. UV detection 205 nm.....	55
Figure 2.8:	HPLC chromatogram of AZM-DH dissolved in 0.1 M HCl (pH 1.2) (1.0 mg/mL) at 205 nm. Mobile phase: phosphate buffer (adjusted with 1.0 M sodium hydroxide to pH 6.0)/acetonitrile (700/300), Flow rate 1.0 mL/min, Injection volume 15 μ L. UV detection 205 nm	55

CHAPTER 3: AZITHROMYCIN: A MACROLIDE ANTIMICROBIAL

Figure 3.1	Chemical structure of azithromycin base (anhydrous form).....	67
Figure 3.2	Chemical structure of erythromycin.....	68
Figure 3.3	Chemical structure of azithromycin dihydrate.....	68

CHAPTER 4: POLYMORPHISM OF AZITHROMYCIN

Figure 4.1	Overlay of DSC traces of AZM-DH (red) and AZM (green), dried in an oven for 60 minutes at 100°C, and of anhydrous AZM (magenta), dried in a DSC prior to analysis	84
-------------------	---	----

Figure 4.2	TGA thermogram indicating a 2.79 % weight loss for AZM, dried in an oven for 60 minutes at 100°C prior to analysis.....	85
Figure 4.3	FTIR spectra of AZM-DH (red), and AZM (green), dried in an oven for 60 minutes at 100°C prior to analysis.....	86
Figure 4.4	XRPD patterns of AZM (blue) and AZM-DH (red), dried in an oven for 60 minutes at 100°C prior to analysis.....	86
Figure 4.5	TGA thermogram indicating a 0.89 % weight loss for anhydrous AZM, dried in the furnace of a TGA prior to analysis.....	88
Figure 4.6	DSC trace of anhydrous AZM, prepared with isopropanol.....	89
Figure 4.7	Enhanced DSC trace ranging between 49°C and 53°C of anhydrous AZM, prepared with isopropanol.....	89
Figure 4.8	TGA thermogram indicating a weight loss of 14.39 % for anhydrous AZM, prepared with isopropanol.....	90
Figure 4.9	FTIR spectra of anhydrous AZM, prepared with isopropanol (red) and of AZM-DH (black).....	91
Figure 4.10	XRPD pattern of anhydrous AZM, illustrating the halo pattern typical of amorphous solids.....	92
Figure 4.11	HSM images (taken at increasing temperatures) of anhydrous AZM, prepared with isopropanol. (a) The anhydrous AZM morphologically presents as an amorphous glass at room temperature, (b) The transition from solid to liquid occurs at 52°C, (c) Crystals start to form from the liquid at 62°C as indicated by the red squares, (d) Crystals resulting from the continuous growth at increasing temperatures up to 101°C, the point at which the crystals start to melt	93

CHAPTER 5: AZITHROMYCIN GLASS

Table 5.1	Peak differences in FTIR patterns to distinguish between AZM-DH and AZM-G	100
------------------	---	-----

Table 5.2	Summary of the percentage water content of AZM-DH and AZM-G as determined with KFT and TGA.....	106
Table 5.3	Solubility data of AZM in 0.1 M HCl	113
Table 5.4	Solubility data of AZM in acetate buffer (pH 4.5).....	115
Table 5.5	Solubility data of AZM in phosphate buffer (pH 6.8).....	116
Table 5.6	Solubility data of AZM in water.....	117
Table 5.7	Summary of the screening test outcomes of AZM-G during the four weeks stability study at 40°C and 75 % RH	123
Table 5.8	T_g values of AZM-G according to various formulae	124
Table 5.9	HSM images of AZM-G (unmilled) over the course of the stability study	135
Table 5.10	T_g values of AZM-G with increased water fraction	139
<hr/>		
Figure 5.1	FTIR spectrum of AZM-DH	99
Figure 5.2	FTIR spectrum of AZM-G.....	99
Figure 5.3	XRPD pattern of AZM-DH.....	101
Figure 5.4	XRPD pattern of AZM-G	102
Figure 5.5	DSC trace of AZM-DH showing dehydration at 76.35°C and 86.45°C and the subsequent melting endotherm at 119.04°C	103
Figure 5.6	DSC trace of AZM-G with T_g (enhanced trace) at 106.65°C	104
Figure 5.7	TGA thermogram of AZM-DH indicating a weight loss of 4.40 %	105
Figure 5.8	TGA thermogram of AZM-G indicating a weight loss of 0.61 %.....	105

Figure 5.9	HSM image of AZM-DH (Temperature of 25°C, Magnification at 10x, Sample covered with silicon oil).....	107
Figure 5.10	SEM images (a & b) of AZM-DH on a scale of 500 µm (a) and 200 µm (b).....	107
Figure 5.11	(a) HSM and (b-d) SEM images of unmilled AZM-G viewed on a scale of 500 µm (b-c) and 200 µm (d) respectively	108
Figure 5.12	SEM image of milled AZM-G on a scale of 50 µm.....	109
Figure 5.13	Series of HSM images (a-f) of AZM-G illustrating the morphological changes during a glass transition from a solid glass into a liquid state. (a) HSM image taken at 80°C illustrating that AZM-G is a glassy solid, (b) HSM image at 90°C illustrating that AZM-G still exists as a glassy solid, (c) HSM image taken at 95°C illustrating that the glassy solid is starting to show signs of reduced viscosity as the molecules become more mobile, (d) HSM image of AZM-G at 99°C illustrating a further decrease in viscosity of the supercooled liquid (glass), (e) HSM image taken at 104°C with the glass transition from the solid into the liquid being visible, (f) HSM image at 106°C, illustrating the complete transition from the glassy solid into the liquid phase.....	110
Figure 5.14	Percentage degradation of AZM-DH and AZM-G in 0.1 M HCl (pH 1.2).....	112
Figure 5.15	Histogram to compare the solubility values (mg/mL) obtained for AZM-DH, AZM-G and milled AZM-G in 0.1 M HCl (pH 1.2)	114
Figure 5.16	Histogram to compare the solubility values (mg/mL) obtained for AZM-DH, AZM-G and milled AZM-G in acetate buffer (pH 4.5)	115
Figure 5.17	Histogram to compare the solubility values (mg/mL) obtained for AZM-DH, AZM-G and milled AZM-G in phosphate buffer (pH 6.8).....	116
Figure 5.18	Histogram to compare the solubility values (mg/mL) obtained for AZM-DH, AZM-G and milled AZM-G in distilled water	117
Figure 5.19	Comparative histogram of the solubility of AZM-DH (blue), unmilled AZM-G (maroon), and milled AZM-G (green) in different aqueous media having varying pH values	118

Figure 5.20	Water vapour adsorption (solid line) and desorption (dotted line) isotherms of AZM-DH (x-axis representing the % RH, y-axis representing the % weight)	120
Figure 5.21	Water vapour adsorption (solid line) and desorption (dotted line) isotherms of unmilled AZM-G.....	121
Figure 5.22	Water vapour adsorption (solid line) and desorption (dotted line) isotherms of milled AZM-G.....	121
Figure 5.23	XRPD pattern of AZM-G after exposure to 100 % RH for a period of 3 days	122
Figure 5.24	Overlay of DSC traces of AZM-DH generated during the stability (Initial (blue), Week 2 (green), Week 4 (red))	124
Figure 5.25	Overlay of DSC traces of AZM-G generated during the stability study (Initial (green), Week 1 (red), Week 2 (blue), Week 3 (magenta), Week 4 (black))	126
Figure 5.26	Overlay of TGA thermograms of AZM-DH generated during the stability study (Initial (blue), Week 2 (red), Week 4 (green))	128
Figure 5.27	Overlay of TGA thermograms of AZM-G generated during the stability study (Initial (green), Week 1 (red), Week 2 (blue), Week 3 (magenta), Week 4 (black)).	129
Figure 5.28	XRPD pattern for AZM-DH serving as reference during the stability study.....	131
Figure 5.29	Overlay of XRPD patterns of AZM-G (unmilled) generated during the stability study (Initial (red), Week 1 (blue), Week 2 (green), Week 3 (grey), Week 4 (brown))	131
Figure 5.30	FTIR spectrum of AZM-DH that was used as the reference during the stability study.....	133
Figure 5.31	Overlay of FTIR spectra of AZM-G generated during the stability study (Initial (maroon), Week 1 (grey), Week 2 (blue), Week 3 (red), Week 4 (green))	133

Figure 5.32	(a) HSM image of AZM-DH and (b) SEM image of AZM-DH at Week 0	134
Figure 5.33	HSM images of AZM-DH at (a) Week 2 and (b) Week 4	134
Figure 5.34	Overlay of the FTIR spectra of AZM-DH (blue), unmilled AZM-G (red) and milled AZM-G (green) after 20 days in water	137
Figure 5.35	DSC trace of AZM-G indicating an endothermic event at 109.13°C after 20 days in water	138
Figure 5.36	Overlay of DSC traces of AZM-G with increased water fraction, ranging from 1 - 50 %. (1 % (brown); 3 % (yellow); 5 % (light blue); 7 % (black); 15 % (magenta); 30 % (green); 40 % (red); 50 % (blue))	140
Figure 5.37	Overlay of the experimental T_g (yellow) and the T_g according to the linear- (orange), Gordon-Taylor- (blue) and the Fox equations (red).....	141
Figure 5.38	FTIR spectra of AZM-DH (black), AZM-G with 0 % water (blue), and AZM-G with an added 50 % water fraction (grey)	141

CHAPTER 6: PRODUCT DEVELOPMENT

Table 6.1	Trial formulation of a 700 mg tablet containing 500 mg of AZM-G.....	157
Table 6.2	Summary of excipients included in tablet formulations prepared by using wet granulation to improve powder flow properties of formulations.....	158
Table 6.3	Final formulation of AZM-DH tablets	163
Figure 6.1	Particle size distribution of AZM-DH.....	152
Figure 6.2	Particle size distribution of AZM-DH granules	152
Figure 6.3	Particle size distribution of AZM-G	153
Figure 6.4	Particle size distribution of AZM-G granules	153

Figure 6.5	Images of the wet granulation process during which dry contents are mixed in a planetary mixer (left) to produce a wet mass of granules (right)	159
Figure 6.6	Images of the dried mass of granules (left) and the mesh with which the dry mass granules were sieved (right)	159
Figure 6.7	Images of the resulting dry granules after being sieved (the left) and the rotating mixer (Turbula) shaking the glass jar containing the granulated powder and excipients (right)	160
Figure 6.8	Images of the CADMACH [®] single punch tablet press (left) and the manufactured tablets (950 mg) containing 500 mg of AZM-G (right)	161
Figure 6.9	SEM micrographs of AZM-G powder (left, scale of 100 μm) and dry granules before tableting (right, scale of 50 μm).....	162
Figure 6.10	SEM micrograph of a dry granule at a higher magnification (scale: 20 μm).....	162
Figure 6.11	SEM micrographs of AZM-DH (left, scale: 200 μm) and dry granules before tableting (right, scale: 50 μm).....	164
Figure 6.12	Dissolution profiles (percentage AZM dissolved as a function of time) of AZM-DH and AZM-G tablets in pH 4.5 acetate buffer	165
Figure 6.13	Dissolution profiles (percentage AZM dissolved as a function of time) of AZM-DH and AZM-G tablets in pH 6.8 phosphate buffer	166
Figure 6.14	Dissolution profiles (percentage AZM dissolved as a function of time) of AZM-DH and AZM-G tablets in water	167
Figure 6.15	Overlay of FTIR spectra for AZM-DH tablets during the stability study. AZM-DH Initial (green), Month 1 (blue), Month 2 (grey), Month 3 (red)	170
Figure 6.16	Overlay of FTIR spectra for AZM-G tablets during the stability study. AZM-G Initial (blue), Month 1 (grey), Month 2 (red), Month 3 (green)	171
Figure 6.17	Dissolution profiles of AZM-DH tablets in pH 6.8 phosphate buffer	172
Figure 6.18	Dissolution profiles of AZM-DH tablets in pH 6.8 phosphate buffer	172

CHAPTER 7: PERMEABILITY

Table 7.1	Concentrations of AZM-DH and AZM-G in different transport media.....	182
Table 7.2	Results of statistical analyses for AZM-DH and AZM-G in different media at different pH values.....	194
Table 7.3	Correlation between the improved solubility of AZM-G and the permeability of AZM at variable pH values.....	195
<hr/>		
Figure 7.1	Graphic illustration of the different mechanisms of intestinal absorption. (A) Paracellular diffusion; (B) Paracellular diffusion enhanced by a modulator of tight junctions; (C) Transcellular passive diffusion; (C* Intracellular metabolism); (D) Carrier-mediated active transcellular transport; (E) Transcellular diffusion coupled with an efflux mechanism; (F) Transcellular endocytosis (Taken from Hunter & Hirst, 1997:131)	179
Figure 7.2	Isolated section of pig intestinal tissue after being washed with cold Krebs Ringer bicarbonate buffer	184
Figure 7.3	Isolated intestinal mucosa sheet after removal of the serosal layer, prior to cutting it into smaller pieces.....	185
Figure 7.4	Diffusion apparatus with the six sets of Ussing chambers.....	185
Figure 7.5	Diffusion apparatus and HPLC vials used during this study	186
Figure 7.6	Overlay of the concentrations of AZM-DH and AZM-G being transported across the intestinal mucosa, as a function of time (pH 7.2 buffer).....	189
Figure 7.7	Overlay of the concentrations of AZM-DH and AZM-G being transported across the intestinal mucosa, as a function of time (pH 6.8 buffer)	191
Figure 7.8	Overlay of the concentrations of AZM-DH and AZM-G being transported across the intestinal mucosa, as a function of time (pH 4.5 buffer).....	192
Figure 7.9	P_{app} values of AZM-DH and AZM-G in different media at different pH values..	194

LIST OF ABBREVIATIONS

> - greater than

≥ - greater than or equal to

≤ - less than or equal to

< - less than

° - degrees

°C - degrees Celsius

Ω - ohm

% - percentage

% WL - percentage weight loss

θ - Theta

μg - microgram

μL - micro litre

μm - micro metre

A - surface area

API - active pharmaceutical ingredient

ATP - adenosine triphosphate

AVG - average

AZM - azithromycin

AZM-DH - azithromycin dihydrate

AZM-G - azithromycin glass

BET - Brunauer-Emmett-Teller theory

BP - British Pharmacopeia

C - carbon

C₀ - initial concentration

cm - centimetre

conc. - concentration

DCM - dichloromethane

DL - limit of detection

DRIFTS - diffuse reflectance infrared Fourier transform spectroscopy

DSC - differential scanning calorimetry

e.g. - *exempli gratia* (for example)

EP - European Pharmacopeia

FTIR - Fourier transform infra-red spectroscopy

g - gram

h - hour

H - hydrogen

H₂O - water

HCl - hydrochloric acid

HPLC - high performance liquid chromatography

HSM - hot stage microscopy

ICH - International Conference on Harmonisation

i.e. - *id est* (that is)

IR - infrared spectroscopy

IUPAC - International Union of Pure and Applied Chemistry

J - Joule

K - Kelvin

KBr - potassium bromide

KCl - potassium chloride

kg - kilogram

KFT - Karl Fischer titration

L - litre

M - molar

m - metre

MAC - *Mycobacterium avium* complex

MDCK - Madin-Darby canine kidney

mg - milligram

MIC - minimum inhibitory concentration

min - minutes

mL - millilitre

mΩ - milli-ohm
MM - molecular mass
mm - millimetre
N - nitrogen
N - newton
NaOH - sodium hydroxide
nm - nanometre
O - oxygen
PAMPA - Parallel artificial membrane permeability assay
 P_{app} - apparent permeability coefficient
P-gp - P-glycoprotein
ppm - parts per million
QL - limit of quantification
 r^2 - correlation coefficient
RH - relative humidity
RNA - ribonucleic acid
rpm - revolutions per minute
RSD - relative standard deviation
s - second
SA - South Africa
SEM - scanning electron microscopy
STDEV (SD) - standard deviation
TAM - Thermal Activity Monitor
TEER - Trans-epithelial electrical resistance
 T_g - glass transition temperature
TGA - thermogravimetric analysis
 T_m - melting temperature (point)
UFLC - Ultra fast liquid chromatography
USA - United States of America
USP - United States Pharmacopeia
UV - ultraviolet

vs - versus

w/w - weight per weight

XRPD - X-ray powder diffraction

LIST OF EQUATIONS

CHAPTER 5: AZITHROMYCIN GLASS

$$R_L = 1 / (1 + b_L \cdot C_0) \dots \dots \dots (5.1)$$

$$T_{gmix} = w_1 T_{g1} + w_2 T_{g2} \dots \dots \dots (5.2)$$

$$T_{gmix} = \frac{(w_1 T_{g1} + k w_2 T_{g2})}{(w_1 + k w_2)} \dots \dots \dots (5.3)$$

Where $k = \frac{(\rho_1 T_{g1})}{(\rho_2 T_{g2})}$

$$\frac{1}{T_{gmix}} = \frac{w_1}{T_{g1}} + \frac{w_2}{T_{g2}} \dots \dots \dots (5.4)$$

CHAPTER 6: PRODUCT DEVELOPMENT

$$P_B = w/v \dots \dots \dots (6.1)$$

$$P_T = w/v \dots \dots \dots (6.2)$$

$$\% \text{ compressibility} = ((P_T - P_B) / P_T) \times 100 \dots \dots \dots (6.3)$$

$$\text{Hausner ratio} = P_T / P_B \dots \dots \dots (6.4)$$

CHAPTER 7: PERMEABILITY

$$P_{app} = (dC/dt) \times (1/A \cdot C_0 \cdot 60) \dots\dots\dots (7.1)$$

$$\%_{AZM-G} = (AZM-G P_{app} / AZM-DH P_{app}) \times 100 \%_{AZM-DH} \dots\dots\dots (7.2)$$

$$\% \text{ improvement} = \%_{AZM-G} - 100 \dots\dots\dots (7.3)$$

ABSTRACT

Azithromycin, an azalide and member of the macrolide group, is a broad spectrum antimicrobial, representing one of the bestselling antimicrobials worldwide. It is derived from erythromycin and exhibits improved acidic stability as a result of its structural modifications. The stable solid form of azithromycin is its dihydrate, although it also naturally occurs in its metastable forms, i.e. the monohydrate and anhydrate. Because azithromycin is poorly soluble in water, its absorption from the gastro-intestinal tract is negatively influenced, which ultimately affects its bioavailability following oral administration (37 %).

Polymorphic (monohydrates and dihydrates) and anhydrous forms of azithromycin were screened and investigated. One anhydrous form also proved to be amorphous, which shifted the focus of this study from polymorphism to amorphism. An amorphous glassy azithromycin was subsequently prepared and fully characterised to present its solid state profile.

The stability of this amorphous glassy form was established at a high temperature and relative humidity over a period of four weeks. Exposure to increased relative humidity (up to 95 %) and increased water content (up to 50 %) also served as stability indicating tests. Its solubility in various aqueous media was determined. A solid dosage form (tablet), containing the azithromycin glass, was prepared, whereafter these tablets were subjected to dissolution studies in different aqueous media. The stability of azithromycin glass in tablet form was determined over a period of three months. The permeability of azithromycin glass across excised pig intestinal tissue was further established at various pH values.

This amorphous glassy form of azithromycin (AZM-G) proved to be very stable at high temperature and relative humidity, whilst also remaining stable after prolonged exposure to 95 % of relative humidity, as it only adsorbed moisture onto its surface. Water content (up to 50 %) had no plasticising effect on azithromycin glass. It demonstrated a significantly higher water solubility (339 % improvement) in comparison with the commercially available azithromycin dihydrate and was it also 39 % more soluble in phosphate buffer (pH 6.8) than its dihydrate counterpart.

The prepared azithromycin glass tablets showed a promising dissolution profile in water, due to the improved water solubility of this glass form. The transport of azithromycin glass at higher pH values (6.8 and 7.2) across the membrane proved to be significantly higher than that of azithromycin dihydrate, thus also illustrating its pH dependence for its transport across pig intestinal tissue.

The improved water solubility of the azithromycin glass, together with its faster dissolution rate, its superior stability and its increased permeability, may ultimately result in a higher azithromycin bioavailability following oral administration.

These research outcomes hence give rise to the need for investigating the effect of administering lower dosages of azithromycin and to determine whether the same antimicrobial efficacy would possibly be achieved, due to maintaining the same tissue concentration levels at these lower dosages.

Key words: Macrolide; Azithromycin; Amorphous; Solubility; Stability; Dissolution; Permeability.

UITTREKSEL

Asitromisien, 'n asalied wat deel van die makroliedgroep vorm, is 'n breë spektrum antimikrobe geneesmiddel, wat wêreldwyd as een van die topverkopers van antimikrobiese geneesmiddels beskou word. Asitromisien is 'n derivaat van eritromisien en toon verbeterde stabiliteit in suuromgewings, as gevolg van sy strukturele modifikasies. Die dihidraat word as die stabiele soliede vorm van asitromisien beskou, alhoewel dit ook natuurlik in sy monohidraat- en anhidraat vorms voorkom. Omdat asitromisien swak oplosbaar is in water, word sy absorpsie vanuit die gastroïntestinale kanaal negatief beïnvloed, wat uiteindelik weer sy biobeskikbaarheid na orale toediening affekteer (37 %).

Polimorfiese (monohidrate en dihidrate) en anhidriese vorme van asitromisien is oorsigtelik tydens hierdie studie ondersoek. Een anhidraat het ook amorfiese eienskappe getoon, wat daartoe aanleiding gegee het dat die fokus van hierdie studie vanaf polimorfisme na amorfisme verskuif het. 'n Amorfiese glas van asitromisien is vervolgens berei en volledig in terme van vaste vorm eienskappe gekarakteriseer .

Die stabiliteit van die bereide amorfiese asitromisien was by toestande van hoë temperatuur en relatiewe humiditeit oor 'n tydperk van vier weke bepaal. Blootstelling aan verhoogde relatiewe humiditeit (tot en met 95 %) en verhoogde waterinhoud het voorts as stabiliteitsaanduidende toetse gedien. Die oplosbaarheid van die glas is ook in verskeie waterige mediums bepaal. Amorfiese asitromisien is vervolgens in 'n tabletvorm geformuleer, waarna dissolusie studies op die tablette in verskeie waterige mediums uitgevoer is. Die stabiliteit van amorfiese asitromisien in tabletvorm is oor 'n tydperk van drie maande bepaal. Die deurlaatbaarheid van amorfiese asitromisien oor uitgesnyde intestinale varkweefsel by verskillende pH-waardes is verder ook bepaal.

Hierdie amorfiese, glasagtige vorm van asitromisien het bewys gelever dat dit baie stabiel was, ten spyte van blootstelling aan beide hoë temperatuur en relatiewe humiditeit. Verlengde blootstelling aan 95 % relatiewe humiditeit het ook geen effek op die stabiliteit van die glas getoon nie, aangesien dit die vog slegs op die oppervlak adsorbeer het. Toenemende waterinhoud (tot en met 50 %) het geen plastiserende effek op amorfiese asitromisien gehad nie. Die glas het ongeveer 339 % beter wateroplosbaarheid in

teenstelling met die dihidraat van asitromisien aangetoon. Dit was voorts ook 39 % meer oplosbaar in fosfaatbuffer (pH 6.8).

Die dissolusie profiel van die bereide asitromisien (amorfe vorm) tablette in water was baie belowend, weens die verhoogde oplosbaarheid van hierdie vorm. Hoër asitromisien konsentrasies is vinniger verkry in vergelyking met die tablette wat asitromisien dihidraat-bevat het. Die deurlaatbaarheid van amorfe asitromisien oor die intestinale membraan by hoë pH-waardes (6.8 en 7.2) was beduidend beter as die van die dihidraat. Die pH-afhanklikheid van die deurlaatbaarheid van die asitromisien glas oor die intestinale weefsel is hierdeur beklemtoon.

Die verbeterde wateroplosbaarheid van amorfe asitromisien, tesame met die vinniger dissolusie-tempo, die stabiliteit en verhoogde deurlaatbaarheid mag uiteindelik tot verhoogde biobeskikbaarheid na orale toediening lei.

Hierdie navorsingsuitkomstes het dus die behoefte laat ontstaan om die impak van die toediening van laer dosisse te ondersoek, ten einde te bepaal of dieselfde antimikrobiese effektiwiteit moontlik bereik kan word, weens die handhawing van identiese weefselkonsentrasies ten spyte van hierdie laer dosisse.

Sleutelwoorde: Makrolied; Asitromisien; Amorf; Oplosbaarheid; Stabiliteit; Dissolusie; Deurlaatbaarheid.

STUDY OBJECTIVES

Main objectives of this study

HPLC method development

- ✓ Developing a robust HPLC method for identification and quantification of azithromycin

Polymorphism of azithromycin

- ✓ Screening of different solid state forms produced through recrystallisation
- ✓ Preparation of anhydrous azithromycin
- ✓ Characterisation of anhydrous azithromycin

Amorphism of azithromycin

- ✓ Preparation of amorphous azithromycin glass
- ✓ Characterisation of azithromycin glass
- ✓ Solubility and stability of azithromycin glass

Product development

- ✓ Formulating azithromycin glass in a tablet
- ✓ Dissolution profiles of prepared tablets in various aqueous media
- ✓ Stability of azithromycin glass in tablet formulation

Permeability of azithromycin glass at various pH values