

Radioligand binding assays of selected A<sub>1</sub>/A<sub>2A</sub> adenosine antagonists using Rat Membrane Cells and Chinese Hamster Cells expressing Human adenosine receptors

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# ABSTRACT

Alzheimer's and Parkinson's disease are considered to be the first and second most common neurological diseases, which contribute to mortality in the elderly population, reduces quality of life and increase socioeconomic difficulties. Despite numerous neuro-pharmacological products on the market and others that are currently being researched, A<sub>1</sub> and A<sub>2A</sub> adenosine receptor (AR) antagonists are of high interest in controlling Alzheimer's and Parkinson's disease related cognitive deficits (A<sub>1</sub> AR subtype) and Parkinson's disease motor dysfunction (A<sub>2A</sub> AR subtype).

When using animal experiments to ascertain  $A_1$  AR affinity, there are limitations translating these results to humans; as some functions observed with the rat model might not be obtained in humans. Our aim was to establish a radioligand binding assay (RBA) using Chinese hamster ovary (CHO) cells expressing human  $A_1$  ARs. In addition, known 7-azaindole derivatives were screened as novel  $A_1$  AR antagonists using both rat brain membrane cells and CHO cells expressing human  $A_1$  ARs.

An adapted method of Klotz and co-workers (1997) were used to established the A<sub>1</sub> AR radioligand binding assay with CHO cells transfected with human A<sub>1</sub> ARs. Reference compounds DPCPX (A<sub>1</sub> antagonist) and istradefylline (A<sub>2A</sub> antagonist) were used leading to successful binding competition curves, which corresponded with literature values. The 7-azaindole derivatives, previously screened for protein kinase inhibition, were found to be potential A<sub>1</sub> AR antagonists with 7-azaindole-chalcone conjugated compound **1a** showing the highest binding affinity for A<sub>1</sub> ARs in rat and human models (hA<sub>1</sub>K<sub>i</sub> = 0.92  $\mu$ M; rA<sub>1</sub>K<sub>i</sub> = 0.90  $\mu$ M). The 7-azaindole-indanone conjugated compound **1e** showed the second highest affinity (hA<sub>1</sub>K<sub>i</sub> = 1.7  $\mu$ M; rA<sub>1</sub>K<sub>i</sub> = 1.1  $\mu$ M). The 7-azaindole- $\alpha$ -tetralone fused compounds (**1b**, **1c**, **1d**) showed a two- to three-fold lower binding affinity for A<sub>1</sub> ARs in rat and human.

In conclusion, the A<sub>1</sub> AR radioligand binding assay, using transfected CHO cells, was successfully established and affinity values of reference compounds correlated with literature. Therefor reducing the challenges in ethics by minimizing the harvesting of rat brains and high costs of rat models. Further, the 7-azaindole-chalcone core was identified as a promising scaffold to explore modifications for improved A<sub>1</sub> AR affinity, which may play a therapeutic role in Alzheimer's and Parkinson's disease with regards to improved cognitive deficits associated with neurodegenerative diseases.

# Key Words:

Alzheimer's disease, Parkinson's disease,  $A_1$  adenosine receptor antagonists, 7-azaindolechalcone, Chinese hamster ovary cells

# ABBREVIATIONS

[<sup>3</sup>H]DPCPX [<sup>3</sup>H]-8-cyclopentyl-1,3-dipropylxanthine/1,3-[<sup>3</sup>H]-Dipropyl-8-

cyclopenthylxanthineα-syn alpha-synuclein

# Α

Αβ	amyloid-beta
AChEls	acetylcholinesterase inhibitor
AC	adenyl cyclase
ADA	adenosine deaminase
AR(s)	adenosine receptor(s)
ATP	adenosine 5'-triphosphate
В	
BG	basal ganglia
BBB	blood-brain barrier
BChE	butyrylcholinesterase
BDN-F	brain-derived neurotrophic factor
BPT	4-(1-benzylpiperidin-4yl) thiosemicarbazone
с	
cAMP	cyclic adenosine monophosphate
СНО	Chinese hamster ovary
CNS	central nervous system
COMT	catechol-O-methyltransferase
СРА	N6-cyclopentyladenosine
СРТ	8-cyclopentyltheophylline

CSC	8-(3-chlorostyryl)caffeine
CSF	cerebrospinal fluid
D	
DA	dopamine
DDQ	diethyl(3,-4-dihydroxyphene thylamino)quinolin-4-yl)methylphosphonate
DMEM/F12	Dulbecco's modified eagles medium with nutrient mixture F12
DMPX	3, 7-dimethyl-1-propargylxanthine
DMSO	dimethyl sulfoxide
DPCPX	8-cyclopentyl-1,3-dipropylxanthine
F	
FBS	fetal bovine serum
FDA	Food and Drug Administration
FST	forced swim test
G	
GABAB	gamma-aminobutyric acid B
GBA	glucocerebrosidase
GCS	glucosylceramide synthase
GPe	external segment of globus pallidus
GPi	internal segment of globus pallidus
Gs	stimulatory G-protein
GSTO2	glutathione S-transferase omega-2
GTP	guanosine triphosphate
н	

Н

h	human
I .	
IL-1b	interleukin 1 beta
IL-6	interleukin 6
IFN-g	interferon gamma
к	
Ki	inhibition constant
Kd	dissociation constant
КО	knockout
KW-6002	istradefylline
L	
LAMB	Laboratory for Analytical and Molecular Biology
LB	Lewy body
LPS	lipopolysaccharide
LN	Lewy neurite
L-dopa	levo-dopa/L-3,4-dihydroxyphenylalanine
М	
MAO	monoamine oxidase
MAO-A	monoamine oxidase type A
MAO-B	monoamine oxidase type B
MAPK	mitogen-activated protein kinase
MPTP	1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine
N	

Ν

Ν	nitrogen
NEAA	non-essential amino acids
NFTs	neurofibrillary tangles
NM	neuromelanin
NMDA	N-methyl-D-aspartate
NSAIDs	nonsteroidal anti-inflammatory drugs
0	
OCH3	methoxy
ОН	hydroxy
Р	
PBS	Phosphate buffered saline
PD	Parkinson's disease
Pen-Strep	Penicillin-Streptomycin
PET	positron emission tomography
PINK1	phosphatase and tensin homolog (PTEN)-induced putative kinase 1
PON1	paraoxonase 1
PSEN-1	presenilin 1
PSEN-2	presenilin 2
PTEN	phosphatase and tensin homolog -induced putative kinase 1 (PINK1)
R	
r	rat
RBA	radioligand binding assay
REM	rapid eye movement

ROS	reactive oxygen species
S	
SAR	structure activity relationship
SEM	standard error of mean
Sirt1	sirtuin 1
SNpc	substantia nigra pars compacta
SNr	substantia nigra <i>pars reticulata</i>
STN	subthalmic nucleus
т	
TNF-a	tumour necrosis factor alpha
TrkB	tyrosine kinase B

# TABLE OF CONTENTS

ACKNOWLE	DGEMENTSI
ABSTRACT	II
ABBREVIAT	IONSIV
CHAPTER 1:	INTRODUCTION1
1.1	Background1
1.2	Research Problem
1.3	Aims and Objectives5
	ALZHEIMER'S AND PARKINSON'S DISEASE AND ADENOSINE
2.1	General Background6
2.2	Neuropathology7
2.3	Aetiology
2.4	Pathogenesis and/or Mechanism of Neurodegeneration in AD and PD 8
2.4.1	Misfolding Protein9
2.4.2	Mitochondrial Dysfunction9
2.4.3	Oxidative Stress
2.4.4	Neuroinflammation9
2.5	Current Treatment Strategies in AD 10
2.5.1	Acetylcholinesterase Inhibitors11
2.5.1.1	Rivastigmine 11
2.5.1.2	Galantamine 11
2.5.1.3	Donepezil

2.5.2	NMDA Receptor Antagonists	12
2.5.2.1	Memantine	12
2.5.3	Emerging Therapies	13
2.5.3.1	Cerebrolysin	13
2.5.3.2	Targeting Misfolding Protein Aggregation	13
2.5.4	Other Treatment Approaches	14
2.5.5	Current Treatment Strategies in PD	15
2.5.5.1	Levodopa (L-Dopa)	15
2.5.5.2	Carbidopa and Benserazide	16
2.5.5.3	Dopamine Agonists	16
2.5.5.4	Catechol- O- methyltransferase (COMT) Inhibitors	17
2.5.5.5	Monoamine Oxidase (MAO) Inhibitors	18
2.5.5.6	Anticholinergic Drugs	18
2.5.5.7	Amantadine	19
2.5.5.8	Surgery	19
2.5.6	Emerging Therapies	20
2.5.6.1	Targeting $\alpha$ -syn Aggregation	20
2.5.6.2	Targeting Calcium Channel Blocking	20
2.5.6.3	A <sub>1</sub> /A <sub>2A</sub> Antagonist Treatment in AD and PD	21
2.5.6.4	A <sub>1</sub> /A <sub>2A</sub> Antagonist Therapy for AD	21
2.5.6.5	A <sub>1</sub> /A <sub>2A</sub> Antagonist Therapy in PD	22
2.6	Summary	23

CHAPTER 3:	ADENOSINE RECEPTORS	24
3.1	General Background	24
3.2	Adenosine Receptors in AD and PD	24
3.3	Adenosine Receptor Antagonists for the Treatment of AD and PD Symptoms	25
3.3.1	Cognitive Dysfunction	25
3.3.2	Neuroinflammation	26
3.3.3	Depression	27
3.3.4	Neuroprotection	27
3.3.5	Motor Function in PD	28
3.4	Potential Adenosine Receptor Antagonists	29
3.4.1	Xanthine A1 Adenosine Receptors Antagonists	29
3.4.2	Non-xanthine A1 Adenosine Receptor Antagonists	31
3.4.2.1	Monocyclic	31
3.4.2.2	Bicyclic Fused Heteroaromatic System	32
3.4.2.3	Tricyclic Fused Heteroaromatic System	33
3.4.3	Xanthine and Non-xanthine A <sub>2A</sub> AR Antagonists	34
3.4.3.1	Monocyclic Fused Heteroaromatic System	34
3.4.3.2	Bicyclic Fused Heteroaromatic System	35
3.4.3.3	Tricyclic Fused Heteroaromatic System	35
3.4.4	Dual Target A <sub>1</sub> /A <sub>2A</sub> AR Antagonists	36
3.4.4 3.4.5	Dual Target A <sub>1</sub> /A <sub>2A</sub> AR Antagonists Chalcone Based Adenosine Antagonists	

3.6	Summary	39
-	RADIOLIGAND BINDING ASSAY USING CHO CELLS EXPRESSING NOSINE A₁ RECEPTORS	40
4.1	Introduction	40
4.2	Ethics	41
4.3	Cloning of Adenosine Receptor and Stable Transfection of Cells	41
4.4	Culturing of CHO A1 AR Cells	41
4.4.1	Apparatus and Equipment	42
4.4.2	Materials and Reagents	42
4.5	Procedure for Culturing	42
4.5.1	Reviving Frozen Cell Stocks	42
4.5.2	Subculturing Cells	43
4.6	Membrane Storage	44
4.6.1	Apparatus and Equipment:	44
4.6.2	Materials and Reagents	45
4.6.3	Method	45
4.7	Membrane Preparation	45
4.7.1	Apparatus and Equipment	45
4.7.2	Materials and Reagents	45
4.7.3	Method	46
4.8	Protein Concentration	46
4.9	Adenosine A1 Receptor Radioligand Binding Assays	46
4.9.1	Apparatus and Equipment	46

4.9.2	Materials and Reagents 46
4.9.3	Method 47
4.10	Statistical Data Analysis 48
CHAPTER 5:	RESULTS AND DISCUSSION
5.1	Introduction
5.2	Establishing Standard Radioligand Binding Assay for CHO Cells
	Expressing the A <sub>1</sub> AR 49
5.3	Novel 7-Azaindole-Chalcone Core Binding against A1 AR in Rat and
	Human 51
CHAPTER 6:	CONCLUSION
BIBLIOGRAP	HY56
ANNEXURE.	

# LIST OF TABLES

Table 4-1:	Summary of A1 AR radioligand binding assay using CHO cells	48
Table 5-1:	Radioligand binding assay parameters utilizing either rat or CHO $A_1$ AR	49
Table 5-2:	Inhibition constant ( $K_i$ ) values for the binding affinity of reference compounds against rat (r) and human (h) A <sub>1</sub> ARs	50
Table 5-3:	Inhibition constant ( $K_i$ ) values for the binding affinity of 7-azaindole derivatives against rat (r) and human (h) A <sub>1</sub> ARs	51

# LIST OF FIGURES

Figure 11:	Structural similarities of different chalcone based derivatives	4
Figure 4-1:	Illustrates the (A) CHO A $_1$ cells at 50-60 % confluence. (B) Clean CHO	
	A1 growth without mycoplasma contamination44	4
Figure 4-2:	Illustration of 2 mL tube with final order of addition: (1) 20 $\mu$ L radioligand	
	solution (containing [ $^{3}$ H]DPCPX) (2) 2 $\mu$ L test compound (at the desired	
	concentration ranging from 0 $\mu$ M to 100 $\mu$ M) and (3) 178 $\mu$ L membrane	
	suspension (containing CHO $A_1$ membranes)	7
Figure 5-1:	Typical dose-response curve for our in vitro model validation. The	
	average binding-response curve for compound 1a on our human $A_1$	
	determined via a radioligand binding assay using CHO cell membranes	
	expressing the human $A_1$ AR with [ <sup>3</sup> H]DPCPX as radioligand	2
Figure 5-2:	Structural relationships of 7-azaindole derivatives containing either a	
	chalcone, tetralone or indanone moiety53	3

# **CHAPTER 1: INTRODUCTION**

#### 1.1 Background

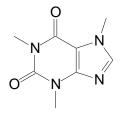
Alzheimer's disease (AD) and Parkinson's disease (PD) are two common age-related chronic neurodegenerative disorders that affect the elderly population over 65 years of age (Calne & Peppard, 1987; Nussbaum & Ellis, 2003). AD and PD are described as the first and second leading neurodegenerative diseases, respectively, and clinically present with cognitive dysfunction in AD and PD, and classical motor dysfunction in PD (Calne & Peppard, 1987, Nussbaum & Ellis, 2003). The prevalence of AD is estimated to be about 1-2% in the age of 65 years and doubles every five years to 35% or more around the age of 85 years, while the prevalence of PD is 1% over the ages of 65 years and climb to 4% at the age of 80 years (Shalash *et al.*, 2017).

AD is characterized by progressive damage of memory and other cognitive abilities leading to dementia (Nussbaum & Ellis, 2003). The cause of AD is unknown. The accumulation of extracellular amyloid plaques containing amyloid-beta (Aβ) peptide and formation of intraneuronal neurofibrillary tangles in the hippocampus, entorhinal cortex, amygdala and cerebral cortex are classical hallmarks of AD (Hauw *et al.*, 1996; Kumar & Clark, 2005). On the other hand, PD is characterized by the progressive deterioration of the basal ganglia dopaminergic nigrostriatal pathway from the substantia nigra pars compacta (SNpc) (Calne & Peppard, 1987). The dopaminergic deficit in PD arises from a loss of the neurons in the SNpc that provide innervation to the striatum (caudate and putamen) - leading to dysregulation of striatal function which account for the symptoms observed in PD; for example resting tremor, muscular rigidity and bradykinesia (Ahlskog & Muenter, 2001). Non-motor symptoms of PD also include cognitive impairment (Simola *et al.*, 2014).

Although scientists have tried to find effective symptomatic treatment, there is, at present, no cure for PD and AD (Di Stefano *et al.*, 2011; Sowell *et al.*, 2009). PD motor symptoms have been reduced by using levodopa (L-3,4-dihydroxyphenylalanine), which is a precursor to dopamine (DA) and remains the drug of choice for the treatment of PD (Ahlskog & Muenter, 2001). Administration of levodopa presents two disadvantages; namely, the onset of motor fluctuations (wearing-off and on-off phenomena) and motor complications (dyskinesias) after prolonged treatment, ultimately leading to disease progression (Ahlskog & Muenter, 2001; Obeso *et al.*, 2000). Other drug options include selective DA receptor agonists, catechol-O-methyltransferase (COMT) inhibitors (LeWitt, 2000), and monoamine oxidase-B (MAO-B) inhibitors (Tabakman *et al.*, 2004). While AD symptomatic treatment options include acetylcholinesterase inhibitor (AchEls) and N-methyl-D-asperate (NMDA) receptor antagonists (Di Stefano *et al.*, 2011), these

compounds only slow disease progression in its early stage but do not help with end stage AD (Di Stefano *et al.*, 2011).

A unique drug that targets all challenges presented by these neurodegenerative diseases is needed urgently to improve quality of life and cut the burden associated with patient care and socio-economic factors (Van Bulck *et al.*, 2019). A systematic approach where a compound targets multiple sites is more likely to succeed as a treatment option for AD and PD to alleviate symptoms (Ribeiro & Sebastião, 2010). Adenosine receptors (ARs) may be the solution to the challenge, since a number of epidemiological studies have shown an inverse relationship between coffee consumption and the prevalence of AD and PD (Lindsay *et al.*, 2002; Martyn & Gale, 2003). Caffeine is a xanthine derivative and acts as a non-selective A<sub>1</sub> and A<sub>2A</sub> AR antagonist (Schwarzschild *et al.*, 2002).



#### Caffeine

Adenosine is a neuromodulator that facilitates signalling via different neurotransmitters and receptors, which include four G-protein coupled ARs, namely A<sub>1</sub>, A<sub>2A</sub>, A<sub>2B</sub>, and A<sub>3</sub> generally found all over the human body (Burnstock, 2006). The number of A<sub>1</sub> and A<sub>2A</sub> ARs in the brain is high which increase the affinity of adenosine. Adenosine acts through inhibitory (A<sub>1</sub> and A<sub>3</sub>) or stimulatory (A<sub>2A</sub> and A<sub>2B</sub>) AR pathways (Chen *et al.*, 2014; Palmer & Stiles, 1995). Co-expression (i.e. heteromers) occurs between A<sub>1</sub> and A<sub>2A</sub> ARs in the presynaptic membranes and A<sub>2A</sub> and dopamine (D<sub>2</sub>) receptors localized around GABAergic striato-pallidal neurons (Azdad *et al.*, 2009; Trifilieff *et al.*, 2011). The activation/blockade of A<sub>1</sub> and A<sub>2A</sub> AR produce a spectrum of pathophysiological activities in the central nervous system (CNS) ranging from cognition (A<sub>1</sub> AR), locomotion (A<sub>2A</sub> AR), behaviour (A<sub>2A</sub> AR) and neurodegeneration (A<sub>2A</sub> AR) (Chen *et al.*, 2014).

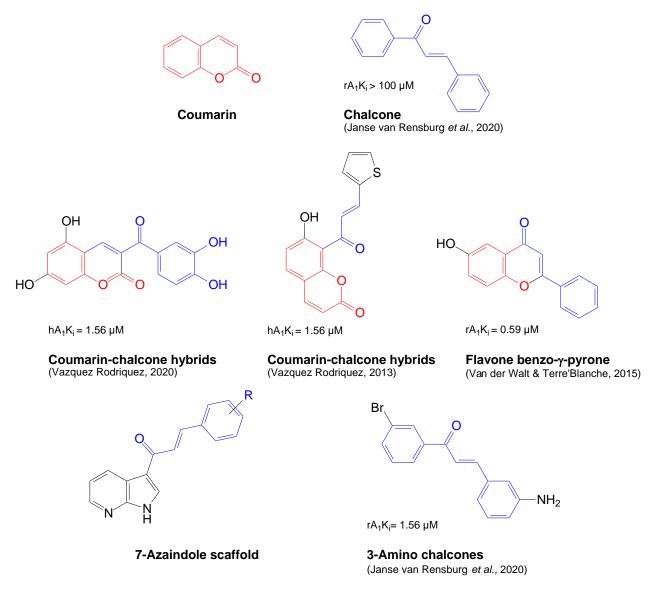
Modification of the adenosine structure has been explored to identify new AR agonists, whereas the xanthine scaffold of caffeine was used for the design of new AR antagonists. However, xanthines display low water solubility, thus limiting their *in vivo* application (Müller *et al.*, 2002). Nitrogen-free heterocyclic ring systems, such as chalcones, are considered a privileged scaffold in medicinal chemistry, seeing that they have attracted attention not only from synthetic and biosynthetic perspectives but also due to their wide-ranging biological activities, such as antimicrobial, antiviral, antifungal, antimalarial, antileishmanial, anti-inflammatory and anticancer

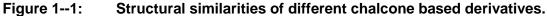
properties (Gaonkar & Vignesh, 2017; Zhuang *et al.*, 2017). Chalcones are a member of the flavonoid family which have been explored as AR antagonists (Van der Walt & Terre'Blanche, 2018), and justly, a chalcone is nothing but an open-chain flavonoid consisting of two aromatic rings linked by an aliphatic three carbon chain. In addition, several chalcone-coumarin derivatives have shown to possess AR affinity (Vazquez-Rodriguez *et al.*, 2013; Vazquez-Rodriguez *et al.*, 2020) and recently C3 amino-substituted chalcone derivatives with a bromo substitution on benzylidene ring B were reported to possess selective adenosine A<sub>1</sub> AR affinity in the micromolar range (Janse van Rensburg, *et al.*, 2020) (**Figure 1-1**).

Radioligand binding assays (RBAs) are widely used for the screening of new potential receptor ligands and are ideally suitable for structure-activity relationship analysis and molecular modelling studies. The basic concept of RBAs comprises of a radioligand that possesses high affinity for a specific receptor. Thus, the radioligands are used to measure the affinity that a test compound possesses for a specific receptor. 1,3-[<sup>3</sup>H]-Dipropyl-8-cyclopenthylxanthine ([<sup>3</sup>H]DPCPX) is an example of a radioligand used to determine a compound's affinity for the adenosine A<sub>1</sub> AR subtype. The adenosine A<sub>1</sub> RBAs are performed with rat whole brain membranes that express the A<sub>1</sub> AR subtype (Van der Walt & Terre'Blanche, 2015). The above-mentioned RBA is frequently performed using rat membranes. However, using rat membranes are costly and the use of animals in scientific and therapeutic research has been a subject of hot discussion for many years. There are strong opposing opinions among enthusiasts and sceptics about the relevance of animal data for humans and the likelihood of successful cross-species translation.

Prompted by the above discussion, the current pilot study undertook to establish the A<sub>1</sub> AR RBA using Chinese hamster ovary (CHO) cells expressing the human A<sub>1</sub> AR. In addition, reference compounds DPCPX and istradefylline (KW-6002) were included for comparison with literature values. Further several known 7-azaindole hybrids, previously evaluated as potential protein kinase inhibitors (Qhobosheane *et al.*, 2020b), were chosen to be screened as novel adenosine A<sub>1</sub> AR antagonists. **Figure 1-1** represents the structural similarity of the 7-azaindole scaffold compared to the chalcone moiety.

Firstly, this chapter provides the background, rationale, research problem and aims and objectives of the current pilot study. Chapter 2 and Chapter 3 contain a literature review of PD, AD and ARs. Chapter 4 describes the experimental procedures and the results and discussion are presented in Chapter 5. Lastly, Chapter 6 summarises the present study and suggests future research.





## 1.2 Research Problem

Understanding how humans respond to drugs is an essential question in drug research. Often this question cannot be addressed because it is not possible to perform some experiments in human volunteers. An alternative can be the generation of responses in animal models and "translating' those results to humans. The limitations are far from clear and further assessment of the potential to extrapolate animal data to humans are needed (Jacobson & Gao, 2006). Several A<sub>1</sub> AR antagonists were screened by Maemoto and co-workers (1997), showing that there were no major differences in inhibition constant ( $K_i$ ) values between human and rat tissue using [<sup>3</sup>H]DPCPX as ligand. Contradictory, the adenosine A<sub>1</sub> antagonist, DPCPX, has a  $K_i$  value of 3 nM in human compared to a value of 0.5 nM in rat using [<sup>3</sup>H]DPCPX as ligand (Alnouri *et al.*, 2015; Maemoto *et al.*, 1997), whereas no such species difference was found for theophylline (Klotz *et al.*, 1997). Supporting these findings Fredholm and co-workers (2001) compared

literature values of antagonists showing the following: DPCPX (human 3.9 nM vs rat 0.3 nM), XAC (human 29 nM vs rat 2.8 nM) and ZM 241385 (human 260 nM vs rat 2 nM).

It is clear that some of the functions observed with one animal model might not be obtained in other animal models and in humans. Cross-species testing is thus important to validate the receptor function or effects of agonists and antagonists. Therefore, the question arises whether the compounds in our library which show high affinity and selectivity at rat  $A_1$  AR will exhibit similar affinity and selectivity for their human orthologues.

# 1.3 Aims and Objectives

Since there is a contradiction in literature between rat and human adenosine receptor affinity data together with the costs and ethics involved in using rats, the aim of this pilot study is to establish the RBA using CHO cells expressing human  $A_1$  ARs. In addition, the affinities of known 7-azaindole-chalcone hybrids will be tested as novel adenosine  $A_1$  AR antagonists using CHO cells expressing human  $A_1$  AR.

In short, the objectives of this study are:

- Growing and harvesting of transfected CHO cells expressing human A1 AR
- Establishing RBA using expressed CHO cells
- Comparing affinity values of reference compounds DPCPX and istradefylline in both rat brain membrane cells and CHO cells expressing the human A<sub>1</sub> AR
- Full RBA using CHO cells and promising 7-azaindole-chalcones previously screened in rat RBA (unpublished results)
- To ascertain structure activity relationships of 7-azaindole-chalcones which govern A1 AR affinity

# CHAPTER 2: ALZHEIMER'S AND PARKINSON'S DISEASE AND ADENOSINE RECEPTORS

#### 2.1 General Background

Alzheimer's disease (AD) and Parkinson's disease (PD) are known as the most progressive neurodegenerative disorders, holding the first and second titles, respectively (Xie *et al.*, 2014). These neurological conditions both present a spectrum of clinical features and neuropathological findings.

In 1907 Dr Alois Alzheimer discovered and described "presenile dementia" (Jarvik & Greenson, 1987), Alzheimer described the neurohistopathology and distinguished the pathological hallmark, namely "plaque-only". This hallmark entailed insoluble amyloid-beta (Aβ) protein in the brain parenchyma, the cerebral blood vessels and neurofibrillary tangles (NFTs) consisting of precipitates of hyperphosphorylated forms of microtubules-associated protein tau. The cognitive impairment in patients with AD is associated with synaptic loss in the neocortex and limbic system (Ittner & Götz, 2011). AD is the most common cause of dementia in the elderly, which presents as learning and memory impairment leading to executive dysfunction and, ultimately, interferes with daily life activities (Scheltens *et al.*, 2016).

On the other hand, PD was first described by James Parkinson in 1817 in an essay entitled: "An essay on the shaking palsy" (Parkinson, 2002). The onset of PD is characterized by the degeneration of dopamine neurons in the substantia nigra pars compacta (SNpc) within the basal ganglia (BG), as well as Lewy neurite intracellular inclusions composed of  $\alpha$ -synuclein ( $\alpha$ -syn) proteins (Xie *et al.*, 2014). Classically known as a movement disorder, PD presents with motor symptoms such as tremor, rigidity, bradykinesia and imbalance (Jankovic, 2008). Non-motor symptoms include sleep disorders, psychiatric symptoms, olfactory dysfunction, gastrointestinal and cognitive dysfunction (Simola *et al.*, 2014).

Both AD and PD present common features such as protein aggregation, oxidative stress, progressive neuronal degeneration, systemic- and neuroinflammation. Additionally, AD and PD boast an array of treatment options which do not modify disease progression but only treat symptoms (Van Bulck *et al.*, 2019).

The prevalence of AD and PD continue to increase because there is, at present, no cure (Prince *et al.*, 2015; Rocca, 2018). This assertion is substantiated in the literature, and AD is commonly known as the most prevalent and PD the second most prevalent neurodegenerative disorders in the world (Alam *et al.*, 2016). In 2015, there were 46.8 million AD patients worldwide with direct

and indirect costs to society of 81,800 million USD (Prince *et al.*, 2015), and in 2016 there were 6.1 million individuals with PD worldwide (Rocca, 2018). The incidence of AD and PD throughout the world is expected to triple by 2050 (Weiner *et al.*, 2015).

#### 2.2 Neuropathology

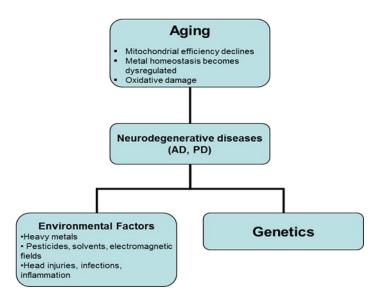
AD is the most common form of dementia in the elderly and is characterized by the presence of A $\beta$  in senile (diffuse and neuritic) plaques, NFTs in the brain and a major loss of synaptic connections (Duyckaerts & Dickson, 2011). Senile plaques that are found in the cerebral grey matter are made of A $\beta$  deposition, surrounded by abnormal arranged neurites (Ferrer *et al.*, 2004; Perl, 2010). NFTs consist of abnormal accumulation of hyperphosphorylated tau proteins found within the perikaryal cytoplasm, including the entorhinal cortex, subiculum, hippocampus, amygdala, Meynert nucleus and neocortex (Ferrer *et al.*, 2004). The presence of dementia and major neurocognitive disorders are highly linked to intermediate and high level neuropathological changes in AD. Clinical diagnosis of symptomatic AD in living patients depend on signs of dementia and positive biomarkers of A $\beta$  from positron emission tomography (PET) and A $\beta$  or tau protein levels from cerebrospinal fluid (CSF) (McKhann *et al.*, 2011), while a definite diagnosis of AD may only be made after death (Šimić *et al.*, 2017).

PD pathology is underlined by degeneration and loss of nigrostriatal dopaminergic neurons, with the presence of Lewy bodies (LB) (Braak *et al.*, 2006). LB or Lewy neurites (LN) are intracytoplasmic neuronal inclusions of the SNpc and largely composed of α-syn (Xie *et al.*, 2014). Additionally, pale staining inclusions can also be found in the amygdala and neocortex (Dickson, 2018). This neurodegeneration affects the noradrenergic locus coeruleus (oral parts) and motor vagal nucleus, the dopaminergic mesocorticolimbic system, the serotonergic raphe nuclei, the cholinergic nucleus basalis of Meynert, pedunculopontine nucleus, Westphal-Edinger nucleus, and many peptidergic brainstem nuclei containing cholecystokinin, met enkephalin, substance P, somatostatin and neuropeptide Y (De Erausquin *et al.*, 1994; Jenner *et al.*, 1992). Braak and coworkers (2006) found that PD pathology extend beyond the substantia nigra where PD starts in the medulla and then moves to the olfactory bulb, pons, substantia nigra, basal forebrain, amygdala, medial temporal lobe and lastly to the cortical area (Di Stefano *et al.*, 2011; Dickson, 2018).

## 2.3 Aetiology

According to literature the aetiology factors of AD and PD are not known or are unclear (Di Stefano *et al.*, 2011; Dickson, 2018), while other studies indicate that environmental factors and genetics play a major role in the cause of AD and PD (**Figure 2-1**) (Chin-Chan *et al.*, 2015). AD and PD,

are both related to old-age and low-grade inflammatory status of the immune system, commonly referred as inflammaging (Franceschi *et al.*, 2000), which might be the most important aetiology factor of these neurodegenerative diseases. These diseases come in two forms: namely familial (genetic cause) and idiopathic (unknown cause). The familial form of PD (various genetic mutations before the age of 65 years) accounts for 5% and the idiopathic form accounts for 95% of all AD and PD cases (Chakrabarti *et al.*, 2015; Grünblatt *et al.*, 2018). Shared common genetic determinants have been hypothesized to be involved in both AD and PD. Xie and colleagues (2014) reported that common genetic mutations of paraoxonase 1 (PON1), glutathione S-transferase omega-2 (GSTO2) and NEDD9 genes are implicated in and shared between AD and PD (Allen *et al.*, 2012; Chapuis *et al.*, 2008). The use of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) to induce neurotoxicity can also cause a syndrome closely related to PD (Langston *et al.*, 1983). Pesticides and herbicides are implicated to cause PD, while exposure to rotenone (a broad-spectrum insecticide, piscicide and pesticide) are also connected to the development of PD in humans (Dauer & Przedborski, 2003).



# Figure 2-1: Etiology of AD and PD neurodegeneration

#### 2.4 Pathogenesis and/or Mechanism of Neurodegeneration in AD and PD

The pathogenic mechanisms of these two neurodegenerative diseases are interrelated with no claim of sole responsibility for disease progression in AD and PD. The pathogenic mechanisms may be attributed to misfolding and aggregation of proteins, mitochondrial dysfunction, oxidative stress, inflammation and apoptosis among others see **Figure 2-2** (Van Bulck *et al.*, 2019).

#### 2.4.1 Misfolding Protein

In AD A $\beta$  aggregates around postsynaptic compartments and produce toxic species including dimers, oligomers and fibrils, which block complex 4-dependent respiration leading to mitochondrial impairment (Ittner & Götz, 2011). Secondly, hyperphosphorylation of tau interferes with neuronal function by impairing mitochondrial complex 1 of the respiratory chain. In PD, LB are largely composed of  $\alpha$ -syn which accumulate in the neurogenic region where adult neurogenesis occur, and impairs olfactory bulb and hippocampal neurogenesis (Horgusluoglu *et al.*, 2017).

#### 2.4.2 Mitochondrial Dysfunction

Mitochondria participate in neurodegenerative disorders by different mechanisms for example misfolding and aggregation of proteins which directly damage mitochondrial DNA, impairment to organelle trafficking and dynamics, bioenergetics/mitophagy processes, apoptosis, oxidative stress, cell metabolism and mitochondria-dependent cell death (Fiorito *et al.*, 2018). PD-related gene mutations are also implicated in mitochondrial quality control through phosphatase and tensin homolog (PTEN)-induced putative kinase 1 (PINK1) and Parkin proteins (Truban *et al.*, 2017). AD-related gene mutations alter mitochondrial activity through PSEN-1 and PSEN-2 proteins (Sarasija & Norman, 2018)

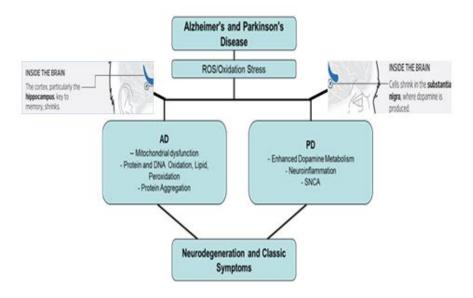
## 2.4.3 Oxidative Stress

It is well reported in literature how AD and PD are associated with oxidative stress via mitochondrial dysfunction and neuroinflammation. At a molecular level, oxidative stress causes peroxidation of lipids and formation of 4-hydroxynonenal, which lead to mutation and destabilization of nucleic acid (Cheignon *et al.*, 2018). These biochemical changes result in a loss of membrane integrity, metabolism, mutation of DNA, a decrease of ATP levels, altered mitochondrial function and activation of pro-apoptotic pathways leading to neuronal death (Hroudov *et al.*, 2014).

#### 2.4.4 Neuroinflammation

Inflammation in AD and PD progression is defined by the accumulation of activated glial cells (astrocytes and microglia) in damaged regions of the brain (Alam *et al.*, 2016), leading to chronic inflammation. Chronic inflammation stimulation may lead to undesired injury which may result in continuous eroding of surrounding tissues causing neuronal death (Akiyama *et al.*, 2000). With aging, prolonged and impaired activation of macrophage and microglia can induce reactive oxygen species (ROS) production (Chinta *et al.*, 2015).

In AD, an autoimmune reaction in the brain activates microglial cells which have both positive and negative effects on AD pathogenesis, where the negative outweighs the positive. Cytokines including **tumour necrosis factor alpha** (TNF-a), interleukin 1 beta (IL-1b) and interleukin 6 (IL-6) act directly on cholinergic neurons and induce their apoptosis (Liang *et al.*, 2017). In PD, autoreactive neuromelanin (NM) specific T cells participate in dopamine neuronal damage and activation of microglia (Koutsilieri *et al.*, 2013). Intraneuronal aggregation of  $\alpha$ -syn protein promotes activation of microglia and astrocytes which produce cytokines TNF-a, IL-1b, IL-6, interferon gamma (IFN-g) and ROS (Hirsch *et al.*, 2012; Liang *et al.*, 2017).



# Figure 2-2: Pathogenesis and mechanism of neurodegeneration of Alzheimer's (AD) and Parkinson's (PD) diseases. Adapted from Tan (2018).

## 2.5 Current Treatment Strategies in AD

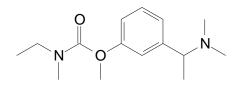
The therapeutic options that are usually selected for AD exploits a symptomatic attack using acetylcholinesterase inhibitors (AChEIs) and an N-methyl-D-aspartate (NMDA) receptor antagonist (Di Stefano *et al.*, 2011). Only four drugs are approved by the United States Food and Drug Administration (FDA); rivastigmine, galantamine and donepezil (AChEIs) and memantine (a NMDA receptor antagonist) (Di Stefano *et al.*, 2011). Cumming and colleagues reported recently that 132 agents were in clinical trials for the treatment of AD (Cummings *et al.*, 2019). Despite the robust development of drugs for the treatment of AD, only symptoms of AD can be addressed by presently approved drugs which intend to improve cognitive function via two different mechanisms: firstly, agonism of the cholinergic system and secondly, antagonism of the NMDA receptor (Di Stefano *et al.*, 2011). The degree of cholinergic loss due to acetylcholine-synthesizing enzyme, namely choline acetyltransferase, in the hippocampus and cortex is related to the level of cognitive impairment and density of amyloid plaques (Bassil & Grossberg, 2009; Perry *et al.*,

1978). During the first year of treatment, AChEIs have the tendency to stabilize cognitive performance and daily functioning, but after the first year the disease, unfortunately, progresses (Scheltens *et al.*, 2016). Just like other AChEIs, donepezil, and most 4-(1-benzylpiperidin-4yl) thiosemicarbazone (BPT) analogues, have the potential to act as moderate AChEIs (Van Bulck *et al.*, 2019). These pyridoxal BPT analogues were tested on five major hallmarks associated with AD and proved to be most effective (Van Bulck *et al.*, 2019). These analogues have shown the ability to inhibit Cu<sup>2+</sup> mediated A $\beta_{(1-40)}$  and A $\beta_{(1-42)}$  aggregation and cellular iron uptake (Amor *et al.*, 2014).

#### 2.5.1 Acetylcholinesterase Inhibitors

#### 2.5.1.1 Rivastigmine

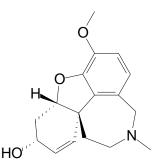
Rivastigmine's inhibitory effect on acetylcholinesterase is termed 'pseudo-irreversible' due to its persistent action after plasma levels of the drug has decreased, while it also inhibits butyrylcholinesterase (Polinsky, 1998). Early clinical trials have shown that rivastigmine exhibits a dose dependent effect – with a higher dose range improving cognition and functionality (Herrmann *et al.*, 2011). Some of the advantages of rivastigmine are less gastrointestinal cholinergic side effects (Massoud *et al.*, 2011).



#### Rivastigmine

#### 2.5.1.2 Galantamine

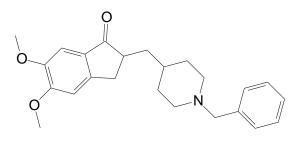
Galantamine is a tertiary alkaloid that is a reversible and competitive cholinesterase inhibitor and nicotinic acetylcholine receptor modulator that improves nicotine transmission (Robinson & Plosker, 2006). In comparison to other AChEIs, galantamine has an inconsistent dose dependence effect, but still remains statistically significant for the improvement of cognition, functionality and behaviour (Herrmann *et al.*, 2011; Massoud *et al.*, 2011).



Galantamine

#### 2.5.1.3 Donepezil

Donepezil binds to acetylcholinesterase in a reversible and non-competitive manner and is hydrolysed instead of acetylcholine (Herrmann *et al.*, 2011). Research has shown that donepezil can protect cortical neurons against glutamate toxicity, prevent apoptotic cell death, increase expression of nicotinic receptors and decrease A $\beta$ -induced toxicity (Herrmann *et al.*, 2011; Mangialasche *et al.*, 2010). Clinical trials have shown that donepezil-treated patients have increased livelihood, but its effects on behaviour were inconsistent (Herrmann *et al.*, 2011; Holmes *et al.*, 2004).



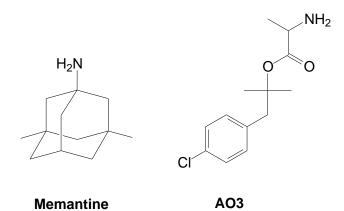
Donepezil

#### 2.5.2 NMDA Receptor Antagonists

#### 2.5.2.1 Memantine

Memantine works as a non-competitive antagonist with moderate affinity for NMDA receptors that target glutamatergic dysfunction (Lipton, 2006). Inhibition of the NMDA receptor is believed to inhibit excitotoxicity of Ca<sup>2+</sup> influx and cell death, while other researchers say the efficacy of memantine is inconclusive (Lipton, 2006). The voltage-dependency and rapid kinetics of memantine allow the reduction in receptor blocking at regular levels which increase blocking at pathological levels and this improves both synaptic function, memory, and cognition (Wenk, 2006). Overall, in clinical trials memantine has shown positive effects on patients' cognition and function, but the effect on the patient's quality of life is unclear (Wilkinson, 2012).

Another non-competitive NMDA receptor antagonist, A03, has a similar effect as memantine on reducing excitotoxicity with additional benefit on sirtuin 1 (Sirt1) expression reduction (Van Bulck *et al.*, 2019). Treatment with A03 increases Sirt1 expression which plays a major role in tau pathology a risk factor for AD (Campagna *et al.*, 2018; Van Bulck *et al.*, 2019).



#### 2.5.3 Emerging Therapies

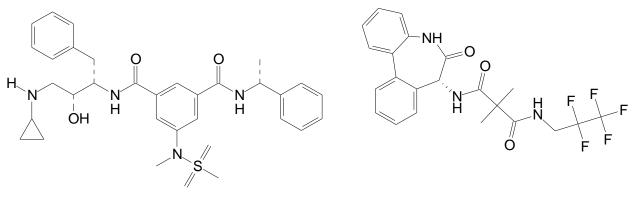
#### 2.5.3.1 Cerebrolysin

Cerebrolysin is a non-cholinergic treatment, which is manufactured using purified brain proteins (Herrmann *et al.*, 2011). It has neurotrophic effects, similar to endogenous nerve growth factors (Veinbergs *et al.*, 2000) and acts as a preserver of neural function and structure under conditions of stress and promotes neuronal plasticity and differentiation (Rockenstein *et al.*, 2007; Veinbergs *et al.*, 2000). The suggestions provided by studies highlight interactions with inhibitory neurotransmitter receptors such as gamma-aminobutyric acid B (GABAB) and adenosine A<sub>1</sub> receptors (Herrmann *et al.*, 2011; Xiong *et al.*, 1996; Xiong *et al.*, 1995). In clinical trials cerebrolysin has shown to have similar efficacy as donepezil, and combination therapy of these drugs possess synergistic properties (Herrmann *et al.*, 2011).

#### 2.5.3.2 Targeting Misfolding Protein Aggregation

IRL-1620 is a synthetic analogue that activates endothelin B receptors which are highly expressed in the central nervous system (CNS) and which play a role in synaptogenesis and neurogenesis (Van Bulck *et al.*, 2019). The clearance of both A $\beta$  toxicity as well as increased cerebral blood flow is stimulated by IRL-1620 which can also increase neural growth factors and Synapsin I expression (Briyal *et al.*, 2015; Gulati *et al.*, 2018).

Further research has shown firstly, that reduction of A $\beta$  production can be stimulated by  $\beta$ -secretase inhibitors,  $\gamma$ -secretase inhibitors and  $\alpha$ -secretase enhancers (Van Bulck *et al.*, 2019). Secondly, decreasing A $\beta$  aggregation by suppressing the accumulation and reducing the stability of A $\beta$  oligomers was achieved by using a nutraceutical (Vivimind<sup>TM</sup>) (Haas, 2012). This nutraceutical has since been marketed, despite no changes in cognitive function (Van Bulck *et al.*, 2019). Lastly, facilitating A $\beta$  clearance is shown to be induced by two strategies involving antibody mediation: active immunization (which include A $\beta$  peptide) and passive immunization (by inserting immunoglobulins) (Van Bulck *et al.*, 2019).

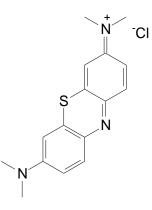


**Beta-Secretase** 

Gamma-secretase inhibitor

#### 2.5.4 Other Treatment Approaches

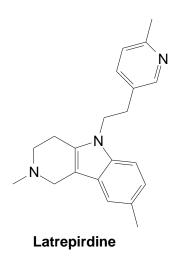
Treatment that targets the pathology of tau, include lithium and valproate which have been used as inhibitors of glycogen synthase kinase-3 and both of these drugs are currently undergoing phase 2 and phase 3 clinical trials, respectively (Tariot & Aisen, 2009). On the other hand methylthioninium chloride has been used to dissolve tau filaments in phase 3 clinical trials (Van Bulck *et al.*, 2019).



#### Methylthioninium chloride

Nonsteroidal anti-inflammatory drugs (NSAIDs) have been related to a reduction in the incidence of AD (Szekely *et al.*, 2004). However a number of neuroinflammatory drugs used in clinical trials have shown no significant improvement of cognition in AD patients (Herrmann *et al.*, 2011).

Mitochondrial dysfunction can be targeted by the use of latrepirdine (also known as dimebon), prescribed in Russia as an anti-histamine. Latrepiridine showed significant cognitive improvement in phase 2 clinical trials, but in a phase 3 multi-national connection trial no improvement in cognition was shown (Doody *et al.*, 2008; Herrmann *et al.*, 2011). Another mitochondrial intervention compound was developed with neuroprotective effects, namely diethyl(3,-4-dihydroxyphene thylamino)quinolin-4-yl) methylphosphonate (DDQ). DDQ reduced A $\beta_{(1-42)}$  levels, and increased A $\beta_{(1-40)}$  levels, mitochondrial ATP, and cytochrome oxidase activity. Further it also reduced free radicals and oxidative stress in AD patients (Kuruva & Reddy, 2017).



# 2.5.5 Current Treatment Strategies in PD

PD is a devastating neurodegenerative disorder, which requires immediate treatment that will prolong the health of neurons while retaining neuronal functions. The aim of the current PD treatment is to alleviate symptomatic presentation by amplification of dopamine signalling in the striatum by mechanisms that (Díaz-Cabiale *et al.*, 2002; Xu *et al.*, 2005):

- increase formation of dopamine (DA) (by the DA precursor levodopa)
- stimulate DA receptors (via DA receptor agonists)
- block the degradation of DA (with MAO-B Inhibitors, and COMT inhibitors)

The common goals of these drugs are to restore the equilibrium of DA in the brain that is affected by dopaminergic cell loss (Borovac, 2016). The antiparkinsonian drugs pose challenges due to debilitating side effects that reduce the patient's quality of life.

#### 2.5.5.1 Levodopa (L-Dopa)

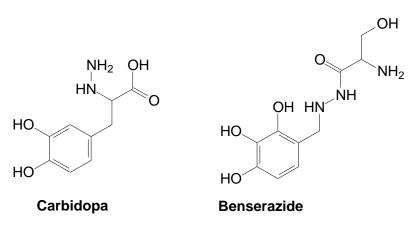
The presence and regional distribution of DA in the human brain as well as it's deficiency in the striatum of PD patients were described in 1959 and 1960 (Nagatsu & Sawada, 2009). PD results as the level of DA decreases in the striatum of the brain; DA is synthesized from L-3,4-dihydroxyphenylalanine (L-dopa).

After prolonged treatment with L-dopa, many patients developed motor complications such as motor fluctuations and abnormal involuntary movement (dyskinesia) (Fabbrini *et al.*, 2007; Pahwa *et al.*, 2006).



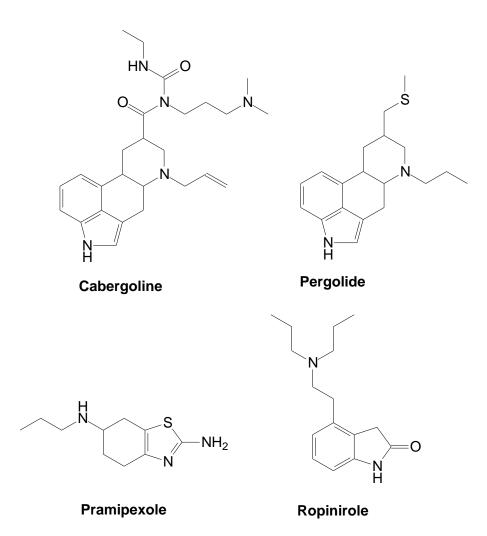
#### 2.5.5.2 Carbidopa and Benserazide

L-dopa is generally combined with benserazide or carbidopa; which are aromatic amino acid decarboxylases inhibitors which do not cross the blood-brain barrier (BBB) but prevent the conversion of L-dopa to dopamine peripherally (Münchau & Bhatia, 2000). Consequently, adverse effects are minimized, central delivery is improved and the dosage of L-dopa can be reduced (Almeida *et al.*, 2004). Other than L-dopa's adverse effects, a major concern is that L-dopa could be neurotoxic, since L-dopa is metabolised to toxic metabolites and free radicals, both possible mechanisms of neurodegeneration in PD (Basma *et al.*, 1995; Graham *et al.*, 1978).



## 2.5.5.3 Dopamine Agonists

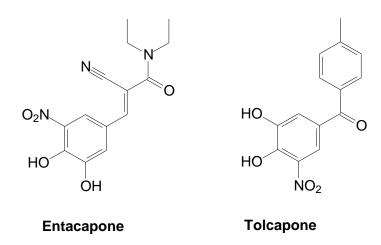
A number of reports argue that the treatment of PD should start with dopamine agonists, as Ldopa leads to the onset of dyskinesia (Montastruc *et al.*, 1999). Two types of drugs are commonly used as dopamine agonists: ergoline and non-ergoline derivative agonists (Borovac, 2016). Ergoline derivatives are first generation DA receptor agonists and also have interactions with receptors other than the D<sub>2</sub> family (Kvernmo *et al.*, 2006); Examples of ergoline agonists are: bromocriptine, cabergoline, pergolide and lisuride (Borovac, 2016). The two most used DA receptor agonists are cabergoline and pergolide which can be used as monotherapy in the early stages of PD (Borovac, 2016). Non-ergoline agonists bind to  $D_2$  and  $D_3$  receptors with high affinity (Borovac, 2016; Frampton, 2014). The most commonly prescribed non-ergoline drugs in the USA are pramipexole and ropinirole (Borovac, 2016). Ergoline agonists have diverse side effects compared to non-ergoline agonists. The side effects of ergoline agonists include constipation, nausea, headache, heart failure, daytime sleepiness and risk of cancer (Pagano *et al.*, 2015; Tholfsen *et al.*, 2015).



2.5.5.4 Catechol- O- methyltransferase (COMT) Inhibitors

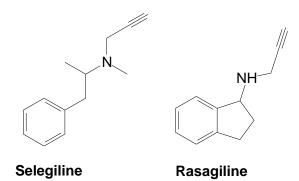
The physiological substrates of COMT include L-dopa, catecholamines (DA, norepinephrine and epinephrine), their hydroxylated metabolites, and catechol estrogen (Ball & Knuppen, 1980). The general function of COMT is to eliminate the biologically active or toxic catechols and other hydroxylated metabolites. The regulation of active DA and norepinephrine in various parts of the brain are associated with COMT and apparently it is associated with moods and other mental

processes (Mannisto, 1999). Studies done on rats have shown that the combination of L-dopa and COMT inhibitors provide increased DA formation and release in the brain (Mannisto, 1999). Entacapone and tolcapone are commonly used COMT inhibitors which prolong the duration of L-dopa effects, thereby increasing DA delivery to the brain. Unfortunately these drugs cause the onset of nausea, anorexia, vomiting, orthostatic hypertension, sleep disorder and hallucination (Kaakkola, 2000).



#### 2.5.5.5 Monoamine Oxidase (MAO) Inhibitors

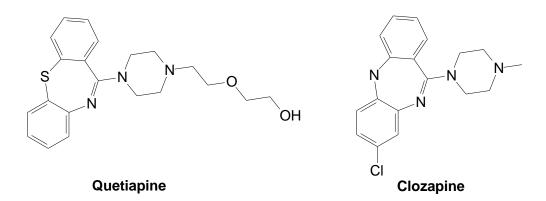
MAO inhibitors prevent the breakdown of biogenic amine neurotransmitters such as DA and thus increases the concentration of DA in the synaptic cleft and postsynaptic receptor sites. MAO-B inhibitors like selegiline and rasagiline are used to treat akinesia and motor fluctuations in PD as monotherapy or in combination with L-dopa and decarboxylase inhibitors. Selegiline causes sleepiness, nausea, vomiting and dizziness in 2-5% of patients, while rasagiline's side effects are nausea, vomiting, orthostatic hypotension, somnolence, hallucination and dyskinesia (Riederer & Laux, 2011).



# 2.5.5.6 Anticholinergic Drugs

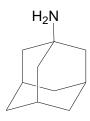
Anticholinergic drugs provide an alternative mechanism of action which alleviates some of the troublesome symptoms experienced in PD, in particular the involuntary resting tremor. Clozapine

and quetiapine are common anticholinergic drugs which can be used as monotherapy in the early stage of PD, and synergistically with L-dopa in more advanced stages of PD (Brocks, 1999). These drugs have the potential to delay the need of L-dopa use, thus reducing the dose of L-dopa required, and further extend the use of L-dopa. Furthermore, anticholinergic drugs decrease extrapyramidal side effects associated with the use of antipsychotic agents (Brocks, 1999). Some reports have shown the relationship between anticholinergic drugs use and cognitive decline (Ehrt *et al.*, 2010) and mental confusion (de Smet *et al.*, 1982).



#### 2.5.5.7 Amantadine

Results from clinical trials with amantadine and a placebo group showed more benefits in relieving akinesia and rigidity than tremors (Dallos *et al.*, 1970). L-dopa induced dyskinesias can be counteracted by amantadine (anti-dyskinetic), however it was reported that the effect wears off after 9 months of treatment (Wolf *et al.*, 2010). Amantadine is initially prescribed as mild symptomatic therapy either as monotherapy or in combination with L-dopa. It enhances the release of DA, the reuptake of DA, anticholinergic effects and blockade of NMDA glutamate receptors (Lees, 2002).



## Amantadine

## 2.5.5.8 Surgery

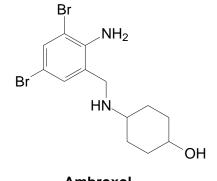
Implantation of electrodes into the bilateral subthalamic nucleus of the globus pallidus for deep brain stimulation in patients with advanced PD have shown significant motor benefits (Deep-Brain Stimulation for Parkinson's Disease Study Group, *et al.*, 2001; Hertel *et al.*, 2006). This procedure

reduced dyskinesia and motor fluctuations in patients, but stimulation has shown to mirror the effects of a destructive lesion whereby high risk of intracranial bleeding occurs (Deep-Brain Stimulation for Parkinson's Disease Study Group; J.A. Obeso *et al.*, 2001).

#### 2.5.6 Emerging Therapies

#### 2.5.6.1 Targeting $\alpha$ -syn Aggregation

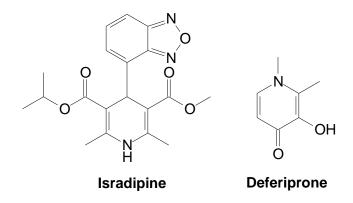
As mentioned earlier  $\alpha$ -syn aggregation is one of the hallmarks of PD and there are a number of mechanisms to avoid the detrimental effects of  $\alpha$ -syn (Van Bulck *et al.*, 2019). Protein accumulation can be avoided by increasing its degradation. A decrease in the glucocerebrosidase (GBA) protein activity leads to accumulation of  $\alpha$ -syn. GBA enhancer drugs (e.g. ambroxol - currently in phase 2 clinical trials) can be used to increase GBA's activity. Alternatively, the inhibition of glucosylceramide synthase (GCS), a substrate of GBA, can be inhibited by GZ/SAR40261 which is in phase 2 clinical trials (Wong & Krainc, 2017).



Ambroxol

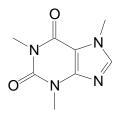
# 2.5.6.2 Targeting Calcium Channel Blocking

Cytosolic Ca<sup>2+</sup> overload may lead to oxidative stress and alleviating increased Ca<sup>2+</sup> load in DA cells can be achieved by L-type voltage gated calcium channel blockers. For instance, isradipidine which specifically blocks voltage gated calcium 1.3 channels and is currently in phase 3 clinical trials (NCT02168842). Some developing drugs that target metal ion homeostasis alteration in PD are also used in AD (Van Bulck *et al.*, 2019). Deferiprone, which is an iron chelator in phase 3 clinical trials is used as possible disease-modifying drug against PD (Belaidi & Bush, 2016).



2.5.6.3 A<sub>1</sub>/A<sub>2A</sub> Antagonist Treatment in AD and PD

The first observation of AR signalling, dates back to 1927 after intravenous adenosine caused temporary heart block (Drury & Szent-Györgyi, 1929). Then only after cloning of purinergic (adenosine) receptors in the 1990's the therapeutic potential of A<sub>1</sub>, A<sub>2A</sub>, A<sub>2B</sub> and A<sub>3</sub> ARs were explored (Ralevic & Burnstock, 1998), with the emphasis on A<sub>1</sub> and A<sub>2A</sub> ARs as drug targets for neurological diseases (Burnstock, 2017). Antagonism of both A<sub>1</sub> and A<sub>2A</sub> ARs are reported to improve cognitive and memory decline in AD (Agostinho *et al.*, 2010; Fredholm *et al.*, 1999), as well as cognitive and motor impairments in PD (Mihara *et al.*, 2007). Case control and prospective studies have associated the consumption of the non-selective A<sub>1</sub>/A<sub>2A</sub> AR antagonist caffeine with a decreased risk of AD (Lindsay, 2002; van Boxtel *et al.*, 2003) and PD (Ascherio *et al.*, 2001; Ross, 2000).

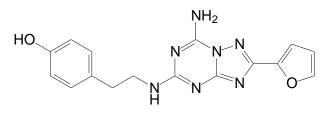


Caffeine

### 2.5.6.4 A<sub>1</sub>/A<sub>2A</sub> Antagonist Therapy for AD

Angulo and colleagues (2003) clearly showed that AD patients has a change in the pattern of expression and redistribution of ARs compared with samples from controls. Therefore, the location of  $A_1$  ARs in neurodegenerative structures such as neurofibrillary tangles and of  $A_{2A}$  ARs in the microglia of patient's hippocampus, suggest that ARs can play a role in regulating the events along the development of AD.

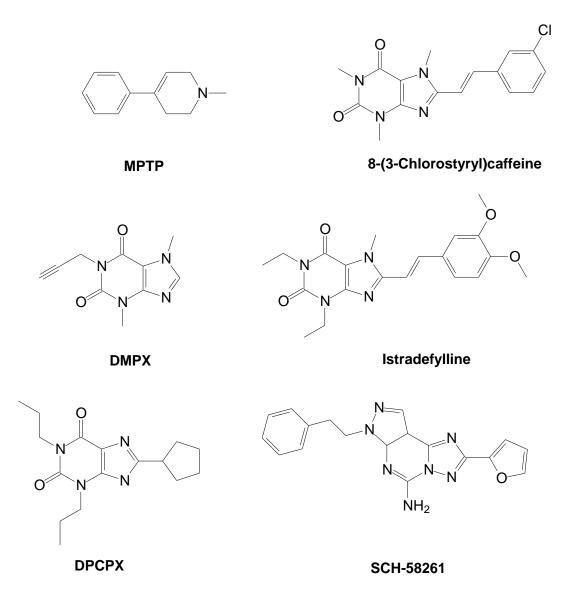
A novel, potent and selective  $A_1$  AR antagonist FR194921 exerts both cognitive-enhancement and anxiolytic activity (Maemoto *et al.*, 2004). Further investigations on rodents were done by intraperitoneal injection of caffeine, an  $A_1$  and  $A_{2A}$  AR antagonist, and ZM 241385, a selective  $A_{2A}$  AR antagonist. Both caffeine and ZM 241385 prevented learning and memory impairment by attenuating neuronal damage caused by A $\beta$  toxicity in AD (Dall'Igna *et al.*, 2003).



ZM241385

## 2.5.6.5 A<sub>1</sub>/A<sub>2A</sub> Antagonist Therapy in PD

Reports have shown that PD patients with dyskinesia have an increased number of A<sub>2A</sub> ARs in their putamen, postulating that L-dopa induced dyskinesia is linked to the A<sub>2A</sub> AR (Chen et al., 2013; Mishina et al., 2011; Ramlackhansingh et al., 2011). Studies of induced toxicity of MPTP, showed either xanthine based compounds such as 8-(3-chlorostyryl)caffeine (CSC), 3, 7dimethyl-1-propargylxanthine (DMPX) or istradefylline (KW-6002) or non-xanthine like SCH-58261, to attenuate dopamine-induced neurotoxicity (Field et al., 2011; Lopes et al., 2011). Further results showed the neuroprotective abilities of A2A AR antagonists, while A1 AR antagonists did not show any neuroprotection after multiple doses with 8-cyclopentyl-1,3dipropylxanthine (DPCPX) (Lopes et al., 2011). A different study by Mihara and colleagues (2007), showed that dual A<sub>1</sub>/A<sub>2A</sub> AR antagonist ASP-5854 reversed memory loss induced by scopolamine in the rat model, whereas specific A<sub>2A</sub> AR antagonist, KW-6002, did not. In addition, ASP-5854 improved haloperidol-induced catalepsy in rats and therefore it was concluded that ASP-5854 improves motor deficits and is neuroprotective through A<sub>2A</sub> AR antagonism, and enhances cognitive function through A1 AR antagonism. KW-6002 is the only A2A AR antagonist treatment that has been approved in Japan since March 2013, while the FDA still needs statistically significant efficacy in the decrease of symptoms of the disease before approving it in the USA (Mishina et al., 2011; Lopes et al., 2011); however, the FDA recently approved KW-6002 as adjunctive treatment for parkinsonian motor symptoms.



#### 2.6 Summary

Tremendous efforts have been made to combat neurodegenerative diseases such as AD and PD through different treatment strategies. As discussed in this chapter, PD and AD can also share the same aetiology mechanism, while presenting different symptoms. The challenges concerning different treatments were briefly discussed, while it is important to note that the current treatments only alleviate symptoms with added side effects and no cure. Research has shown that dual  $A_1/A_{2A}$  AR antagonism may be used for the pharmacological treatment of AD and PD symptoms with the addition of  $A_{2A}$  AR antagonists possessing neuroprotective properties. A multi-drug target approach involving manipulation of AR signalling in combination with L-dopa has been postulated, which reduces the risk of developing dyskinesia presented by L-dopa in PD treatment (Lopes *et al.*, 2011). A detailed discussion of ARs and their antagonists follows in Chapter 3.

# **CHAPTER 3: ADENOSINE RECEPTORS**

#### 3.1 General Background

Adenosine is a nucleoside in the brain which is released by neurons and astrocytes into extracellular spaces to act as a neuromodulator, whereby it facilitates signalling by various neurotransmitters and receptors (Chen *et al.*, 2014). The four G-protein coupled adenosine receptors (ARs), namely A<sub>1</sub>, A<sub>2A</sub>, A<sub>2B</sub> and A<sub>3</sub> are generally found throughout the body, A<sub>1</sub> and A<sub>2A</sub> ARs are predominantly expressed in the brain and both possess a high affinity for adenosine (Chen *et al.*, 2014). These AR subtypes usually modulate cAMP formation: the A<sub>1</sub> and A<sub>3</sub> ARs inhibit adenyl cyclase and the A<sub>2A</sub> and A<sub>2B</sub> ARs stimulate adenyl cyclase (Londos *et al.*, 1980, van Calker *et al.*, 1979)

A<sub>1</sub> ARs are distributed throughout the brain, and are most abundant in the limbic and neocortical regions and present in the basal ganglia and cerebellum, and in most nuclei of diencephalons of the brain stem (Yuzlenko & Kiec-Kononowicz, 2006). A<sub>2A</sub> ARs are predominantly distributed in the basal ganglia as well as limbic and neocortical regions in the brain (Yuzlenko & Kiec-Kononowicz, 2006). Co-expression (heteromers) of A<sub>1</sub> and A<sub>2A</sub> (A<sub>1</sub>/A<sub>2A</sub>) ARs is usually found in presynaptic membranes, while A<sub>2A</sub> and dopamine D<sub>2</sub> receptor (A<sub>2A</sub>/D<sub>2</sub>) heteromers are selectively localized on GABAergic striato-pallidal neurons (Azdad *et al.*, 2009, Ciruela *et al.*, 2006, Trifilieff *et al.*, 2011). Locomotive control is facilitated by regulation of dopamine (D<sub>1</sub> and D<sub>2</sub>) neurotransmission, which either face motor depression or activation as result of A<sub>1</sub> and A<sub>2A</sub> (agonist or antagonist) effect (Popoli *et al.*, 1996; Armentero *et al.*, 2011). As a substrate of activation both A<sub>1</sub> and A<sub>2A</sub> ARs display a spectrum of pathophysiological activities in the central nervous system which may lead to memory and cognitive decline, depression, motor impairment, neuroinflammation and neurodegeneration in Alzheimer's disease (AD) and Parkinson's disease (PD) (Gomes *et al.*, 2011).

#### 3.2 Adenosine Receptors in AD and PD

Stimulation of  $A_1$  and  $A_{2A}$  ARs by endogenous adenosine enables dilapidation of intercorporate cognition and memory, as experienced in AD (Cunha & Agostinho, 2010). As mentioned above  $A_1$  and  $A_{2A}$  ARs are mainly located around synaptic terminals where these receptors control the release of acetylcholine and glutamate neurotransmitters, involved in memory and other cognitive functions (Cunha, 2005, Ribeiro *et al.*, 2002).

A commonly used therapeutic strategy in AD is through inhibition of acetylcholinesterase (AChE) and N-methyl-D-aspartate (NMDA) (Van Bulck *et al.*, 2019). An A<sub>1</sub> AR antagonist can block the release of AChE thereby acting as a cognitive enhancer (Ribeiro & Sebastiao, 2010). Additionally,

the A<sub>2A</sub>-AR antagonists SCH58261 and ZM 241385 have shown to enhance social memory and halt A $\beta$  induced synaptic loss in animal studies of AD (Kopf *et al.*, 1999, Prediger *et al.*, 2005). Experimental studies have shown that caffeine improves memory and cognitive abilities, reduce A $\beta$  production, stabilize blood-brain barrier (BBB) integrity, due to its anti-inflammatory effects that stimulate the pro-survival cascade and inhibit the pro-apoptotic pathway (Chen *et al.*, 2010, Rivera-Oliver & Díaz-Ríos, 2014). Caffeine is a non-selective A<sub>1</sub>/A<sub>2A</sub> AR antagonist and may be advantageous for AD as it leads to reduced cognitive impairment while minimizing A $\beta$ accumulation (Arendash *et al.*, 2009, Dall'Igna *et al.*, 2007). However, some conflicting reports state that prolonged caffeine exposure can cause desensitization as well as inverse effects that resemble AR agonists (Jacobson *et al.*, 1993).

The involvement of ARs in the development of PD may be as follows: (1) A<sub>1</sub> ARs are commonly found in presynaptic striatal neurons where their stimulation inhibits glutamate, dopamine, and acetylcholine release which contribute to motor and non-motor dysfunctions experienced in PD (Ambrósio *et al.*, 1997, Fredholm & Dunwiddie, 1988). (2) Several studies have highlighted the role of A<sub>2A</sub> AR in the pathophysiology of PD namely: a). the physiological role in motor control; b) ability to regulate glutamatergic transmission; c) increased mediated activity in PD; d) eventual involvement in neuroinflammation in substantia nigra pars compacta (SNpc); e) ability to control metabolism and mitochondrial functioning (Gomes *et al.*, 2011).

Although phase 3 clinical trials revealed that caffeine provides no clinical improvement of motor manifestation in PD (Postuma *et al.*, 2017), there is alarming evidence of prospective and case-control studies that show caffeine intake is related to a reduction in the risk of PD (Palacios *et al.*, 2012). Currently, the first line pharmaco-therapeutic strategy in PD targets restoring dopamine levels and/or effects by the use of a dopamine precursor, dopamine agonists and inhibitors of enzymatic degradation of dopamine. These drugs mainly alleviate motor symptoms and limitations include long-term side effects (in particular, motor disability, including dyskinesia) and inability to stop the ongoing degenerative process (Gomes *et al.*, 2011).

### 3.3 Adenosine Receptor Antagonists for the Treatment of AD and PD Symptoms

## 3.3.1 Cognitive Dysfunction

Caffeine is related to the regulation of neurotransmitter release and increased synaptic plasticity through A<sub>1</sub> and A<sub>2A</sub> AR antagonism that affects neuronal processes associated with mood and cognition (Gomes *et al.*, 2011). Human studies demonstrated the effects of caffeine in preventing and delaying the onset of old-aged cognitive decline (Eskelinen *et al.*, 2009, Jarvis, 1993, Maia & de Mendonça, 2002).

Various pre-clinical studies support the use of A<sub>1</sub> AR antagonists for the treatment of cognitive dysfunction. As shown with the administration of the A<sub>1</sub> AR antagonist rolofylline (KW-3902) to pro-aggregant-tau transgenic mice which restored the spatial memory deficits and normalized basic (synaptic transmission) neuronal activity (Dennissen *et al.*, 2016). The A<sub>1</sub> AR antagonist 8-cyclopentyl-1,3-dipropylxanthine (DPCPX) seemed to prevent morphine-induced impairment in regaining partial reference memory and further enhanced learning and memory in animal models (Lu *et al.*, 2010, Maemoto *et al.*, 2004). Further scopolamine-induced memory loss was alleviated by orally active FR19492, an A<sub>1</sub> AR antagonist, which also displayed anxiolytic effects in the elevated plus-maze test (Maemoto *et al.*, 2004). In animal studies, both acute and chronic caffeine intake has shown to improve social recognition, olfactory memory, and emotional behaviour; which could be due to suppression of α- and γ-secretase in old rats (Panza *et al.*, 2015, Prediger *et al.*, 2005, Sallaberry *et al.*, 2013).

Growing evidence also supports the  $A_{2A}$  AR as a major player in memory performance as could be seen in both spatial water maze and radial arm maze test of  $A_{2A}$  AR knockout (KO) mice (Gimenez-Llort *et al.*, 2007, Zhou *et al.*, 2009). Various brain insults that resulted in memory impairment has been shown to be reversed by blocking  $A_{2A}$  ARs, while alleviating deposition of A $\beta$  aggregates (Dai *et al.*, 2010, Gelber *et al.*, 2011). The effect of  $A_{2A}$  AR antagonism through processes such as reduction of glutamate overdrive release, increase of brain-derived neurotrophic factor (BDN-F) and tyrosine kinase B (TrkB), also improved synaptic plasticity (Armentero *et al.*, 2011, Sallaberry *et al.*, 2013).

#### 3.3.2 Neuroinflammation

Caffeine and A<sub>2A</sub> AR antagonists can inhibit hippocampal lipopolysaccharide (LPS) induced neuroinflammation in aged mice (Brothers *et al.*, 2010, Rebola *et al.*, 2011) by acting as an anti-inflammatory, anti-rheumatic and immunosuppressive drug. Different studies have shown that consumption of caffeine down-regulate neuroinflammatory responses and nitric oxide production (Salvemini & Mollace., 2013, Tsutsui *et al.*, 2004, Yaday *et al.*, 2012).

A<sub>1</sub> ARs, found around non-neuronal spaces, are able to regulate both for astrocytic and microgliarelated functions in brain tissue (Cupino & Zabel, 2013, Haselkorn *et al.*, 2010, Tsutsui *et al.*, 2004), which explain the evacuation of A $\beta$  accumulation caused by A<sub>1</sub> AR antagonism in and around cerebral blood vessels (Cupino & Zabel, 2013). Additionally, studies on A<sub>1</sub> AR KO mice indicate the anti-inflammatory function of these receptors (Wei *et al.*, 2011).

Pre-treatment of MPTP mice with KW-6002 (A<sub>2A</sub> AR antagonist), present low microglia activation, which further decreased neurotoxic factors, noxious astrogliosis and mitogen-activated protein

26

kinase (MAPK) p38 in the SNpc and striatum (Ikeda *et al.*, 2002, Pinna *et al.*, 2010, Yasuda *et al.*, 2011). The blockade of the A<sub>2A</sub> AR shows a pattern of neuroprotection that consistently link decrease of inflammatory events in AD and PD patients (Armentero *et al.*, 2011, Brambilla *et al.*, 2003, Yu *et al.*, 2008)

#### 3.3.3 Depression

Different studies have shown that consumption of coffee in both men and women decrease depression at doses similar to dementia protection (Lucas *et al.*, 2011, Pham *et al.*, 2013, Ruusunen *et al.*, 2010). The implication of the adenosine system in the pathophysiology of depression was shown by the antidepressant action of A<sub>1</sub> (DPCPX) and A<sub>2A</sub> (ZM 241385) AR antagonists and, also, caffeine which reduced the depressant effects of Zinc in rodents (Lobato *et al.*, 2008). Moreover, A<sub>1</sub> and A<sub>2A</sub> AR KO mice showed anxiogenic-like behaviour (Gimenez-Llort *et al.*, 2002, Johansson *et al.*, 2001).

Adenosine has been shown to produce antidepressant like effects in the forced swim test (FST) in mice through the A<sub>1</sub> AR (Kaster *et al.*, 2004, Lobato *et al.*, 2008), which is contradictory to other studies that have shown high immobility times in the FST when mice were treated with adenosine (Porsolt *et al.*, 1977). Endogenous adenosine, has limited long-term depression effects through the activation of A<sub>1</sub> ARs, while the inhibition of the A<sub>1</sub> AR present anxiolytic properties (de Mendonça *et al.*, 1997, Maemoto *et al.*, 2004).

The A<sub>2A</sub> AR antagonist istradefylline (KW-6002), is not only effective in the treatment of the motor symptoms of PD, but may also induce antidepressant-like effects (Preti *et al.*, 2015). KW-6002 alone or co-administered with currently available antidepressants may be useful for the treatment of depression as well as motor symptoms of PD (Yamada *et al.*, 2013). Various studies have shown that the antidepressant effect of A<sub>2A</sub> AR antagonists such as CSC, SCH-58261, KW-6002, ZM 241385 and caffeine reduced immobility time in the FST and tail suspension test in animal models of depression (El Yacoubi *et al.*, 2003, Huang *et al.*, 2004, Pechlivanova *et al.*, 2012).

### 3.3.4 Neuroprotection

The chronic consumption of caffeine provides neuroprotection by reversing excitotoxicity caused by induced hypoxia and ischemia in animal models. (de Mendonça *et al.*, 2000), and furthermore, protects BBB disruption in both AD and PD (Xuesong *et al.*, 2010). Another mechanism of caffeine-induced neuroprotection results from the increase of pro-BDNF (increasing neurotrophin ratio) which increase neuronal survival and reduction of glutamate overdrive (Armentero *et al.*, 2011, Chen & Chern, 2011).

Various research evidently show that chronic administration of caffeine displayed neuroprotective effects in rat cerebellar neurons where it reduces A $\beta$ -induced neurotoxicity via A<sub>2A</sub> AR antagonists (Arendash *et al.*, 2009, Cao *et al.*, 2009). The selective A<sub>2A</sub> antagonist ZM 241385 showed similar neuroprotection effects to that of caffeine, while the selective A<sub>1</sub> AR antagonist 8-cyclopentyltheohylline (CPT) did not show any neuroprotection (Dall'Igna *et al.*, 2003). The direct injection of the A<sub>2A</sub> AR antagonist ZM 241385 into the hippocampus of male Wistar rats, reduced kainate-induced neuronal damage, while direct injection of A<sub>2A</sub> AR agonist CGS21680 into the hippocampus failed to provide protection (Jones *et al.*, 1998).

Based on various studies A<sub>2A</sub> AR antagonists can conversely protect the brain from damage after ischemia (Chen *et al.*, 1999), excitotoxicity (Popoli *et al.*, 2000), traumatic brain injury (Dai *et al.*, 2010) and PD (Chen *et al.*, 2001, Lopes *et al.* 1999). Caffeine and A<sub>2A</sub> AR antagonists mediate the excessive release of neuroinflammatory responses and promote neuroprotection (Ikeda *et al.*, 2002a). Moreover, A<sub>2A</sub> AR antagonists provide broad-spectrum neuroprotection via its regulation of glutamate neurotransmission and mitochondrial toxicity (Chen *et al.*, 2001)

## 3.3.5 Motor Function in PD

The gradual loss of dopamine in the ventral midbrain and subsequent loss of dopamine input to the forebrain (striatal) motor structures is the major cause of motor impairment in PD (Fink *et al.*, 1992, Schiffmann *et al.*, 1991). PD is a basal ganglia associated disorder. The basal ganglia is made up of input nuclei (caudate, putamen and accumbens), output nuclei (globus pallidus interna (GPi), substantia nigra pars reticulate (SNr), and intrinsic nuclei (globus pallidus externa (GPe), subthalamic nucleus (STN) and SNpc, that facilitate planning and initiation of movement (Armentero *et al.*, 2011). There are two pathways that coordinate information from the striatum to the basal ganglia - firstly the direct pathway, facilitating excitation through A<sub>1</sub> ARs and DA type 1 (D<sub>1</sub>) receptors ending in GPi/SNr nuclei, and secondly the indirect pathway, facilitate inhibition through A<sub>2A</sub> AR and DA type 2 (D<sub>2</sub>) receptors projecting to GPe (Armentero *et al.*, 2011, Mori, 2014). A<sub>1</sub> ARs are localized presynaptically of dopamine axon terminals and inhibition of these AR facilitates DA release in the striatum, whereas A<sub>2A</sub> AR antagonism potentiates the postsynaptic response to DA (Shook & Jackson, 2011).

Restoration of motor activity was demonstrated by the effect of A<sub>2A</sub> AR blockade in different animal models of PD, which includes modulated behaviour in unilateral 6-OHDA lesioned rodents, reduction of motor impairment in MPTP-treated non-human primates; as well as reversion of haloperidol-induced catalepsy (Simola *et al.*, 2008, Xu *et al.*, 2005). Mono-administration of the A<sub>2A</sub> AR antagonist KW-6002 as well as co-administration with L-dopa did not present dyskinesia or any side effects, thereby showing synergy between these compounds (Grondin *et al.*, 1999,

Kanda *et al.*, 2000). Recently reports showed that administration of an A<sub>2A</sub> AR antagonist only show a reduction in the OFF time in dyskinesia but not a significant improvement in dyskinesia (Zheng *et al.*, 2018).

Furthermore, several studies indicated that consumption of caffeine and A<sub>2A</sub> AR antagonist administration is able to attenuate dopaminergic neurotoxicity and neurodegeneration (Chen *et al.*, 2001, Ikeda *et al.*, 2002, Xu *et al.*, 2002). In genetically susceptible patients, caffeine intake also reduced the risk of developing PD (Kumar *et al.*, 2015), supporting a previous meta-analysis of 26 observational studies which showed a decreased risk of developing PD with caffeine intake (Costa *et al.*, 2010). Unfortunately, after prolonged treatment with caffeine, rapid tolerance develops, but this tolerance is not shown by selective A<sub>2A</sub> AR antagonists SCH-58261(Halldner *et al.*, 2000, Popoli *et al.*, 2000).

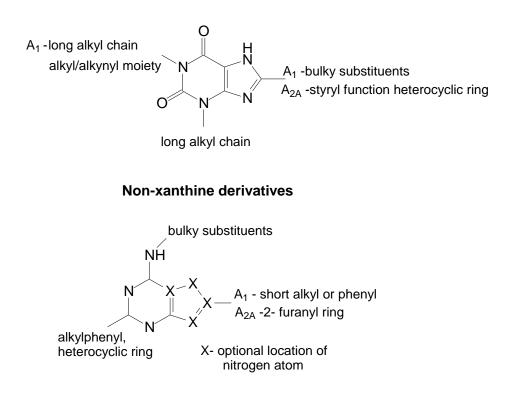
## 3.4 Potential Adenosine Receptor Antagonists

#### 3.4.1 Xanthine A<sub>1</sub> Adenosine Receptors Antagonists

Xanthine derivatives present some physico-chemical disadvantages which include water solubility, CNS penetration and bioavailability (Klotz, 2000, Moro *et al.*, 2006). It was also found that the AR affinity of xanthine based antagonists differ between species, unlike non-xanthine derivatives which display similar AR affinity between different species (Maemoto *et al.*, 1997, Maemoto *et al.*, 2004).

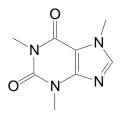
Xanthine based compounds with good A<sub>1</sub> AR affinity, possess bulky C8-substituents as well as long alkyl chains in positions 1 and 3, whereas high affinity of A<sub>2A</sub> AR ligands are C8-styryl substituted with N(1)-alkyl/alkynyl moiety or fused tricyclic xanthines with heteroatoms (Yuzlenko & Kiec-Kononowicz, 2006) (**Figure 3-1**). Promising novel A<sub>1</sub> and A<sub>2A</sub> AR antagonists are generally based on position C8 substitutions with aryl or cycloalkyl groups (Baraldi *et al.*, 2008). Non-xanthine AR antagonists are commonly non-fused monocyclic, fused bi-and tricyclic analogues made of nitrogen, oxygen and sulphur where most often A<sub>1</sub> AR ligands are adenine based with amino group substitution and adjustable nitrogen atoms in the molecule, while A<sub>2A</sub> AR ligands possess good affinity when a furanyl function is present for binding capacity (Yuzlenko & Kiec-Kononowicz, 2006).

#### Xanthine derivatives



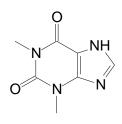
# Figure 3-1: Summary of the general features of xanthine and non-xanthine based A<sub>1</sub> and A<sub>2A</sub> AR antagonists (Yuzlenko & Kiec-Kononowicz, 2006)

Natural xanthines such as caffeine (1,3,7-trimethylxanthine) and theophylline (1,3dimethylxanthine) are known to be non-selective  $A_1/A_{2A}$  antagonists and through different manipulations initiated the discovery of some of the most potent and selective  $A_1$  and  $A_{2A}AR$ antagonists (Yuzlenko & Kiec-Kononowicz, 2006). The xanthine core provided a number of possibilities for xanthine derivatives as they present N1, N3, N7 and C8 positions for substitution to form key pharmaceutically active compounds (Singh *et al.*, 2018). Several studies found that propyl and ethyl substitutions at N1, N3 and N7 positions appear to enhance affinity at  $A_1 AR$ when compared to methyl substitution (Van Der Walt & Terre'Blanche, 2015).



#### Caffeine

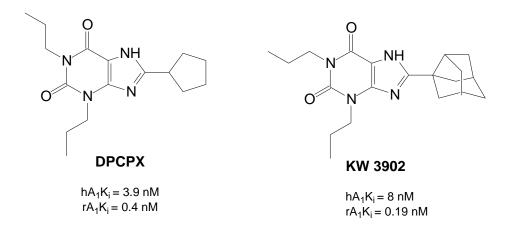
 $hA_1K_i = 44\ 900\ nM\ hA_{2A}K_i = 23\ 000\ nM$  $rA_1K_i = 18\ 800\ nM\ rA_{2A}K_i = 43\ 000\ nM$ 



#### Theophylline

$$\label{eq:Ki} \begin{split} hA_1K_i &= 6\;200\;nM\;hA_{2A}K_i = 4\;200\;nM\\ rA_1K_i &= 8\;500\;nM\;rA_{2A}K_i = 25\;000\;nM \end{split}$$

Manipulation of the xanthine nucleus initiated different potent and selective antagonists, which contain bulky lipophilic substituents leading to the selective A<sub>1</sub> AR antagonist DPCPX. DPCPX has high affinity and selectivity for rat brain A<sub>1</sub> ARs and a 10 fold less affinity for human A<sub>1</sub> ARs (Fredholm *et al.*, 2001). Because of its high affinity and selectivity for A<sub>1</sub> ARs, [<sup>3</sup>H]DPCPX became a ligand of choice in A<sub>1</sub> AR radioligand binding assays (Maemoto *et al.*, 1997). The selective A<sub>1</sub> AR antagonist KW 3902, (rolofylline) showed high affinity for rat and human A<sub>1</sub> ARs and improved presynaptic dysfunction and restored neuronal activity (Dennissen *et al.*, 2016; Müller & Jacobson, 2011).



# 3.4.2 Non-xanthine A<sub>1</sub> Adenosine Receptor Antagonists

Non-xanthine ligands are non-fused monocyclic or fused bi- and tricyclic derivatives with the nitrogen, oxygen and sulphur heteroatoms (Yuzlenko & Kiec-Kononowicz, 2006).

#### 3.4.2.1 Monocyclic

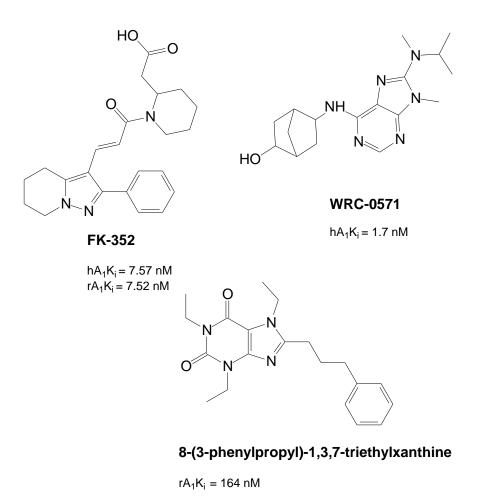
Usually, monocyclic heteroaromatic rings are very rare and most of them have low affinity with only few exceptions of anti-inflammatory action and generally consist of two five-membered heterocycles, thiazoles or thiadiazoles (Chang *et al.*, 2004, Yuzlenko & Kiec-Kononowicz, 2006). Modification of 1,2,4-thiadiazol by substitution of a phenol group resulted in the potent antagonist N-(3-phenyl-1,2,4-thiadiazol-5-yl)-4-hydroxybenzamide (LUF-5437) (Yuzlenko & Kiec-Kononowicz, 2006)



### 3.4.2.2 Bicyclic Fused Heteroaromatic System

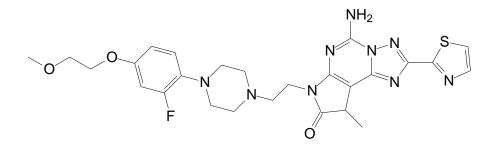
Bicyclic 6:5 fused heteroaromatic compounds form the largest group of published and synthesized non-xanthine A<sub>1</sub> AR antagonists (Yuzlenko & Kiec-Kononowicz, 2006). Pyrazolo[1,5- $\alpha$ ]pyridines have the advantage of water solubility as salts with high selectivity for A<sub>1</sub> ARs and (R)-1-[(E)-3-(2-phenylpyrazolo[1,5-a]pyridin-3-yl)acryloyl]pipendin-2yl acetic acid (FK-352) was identified as a high affinity selective A<sub>1</sub> AR antagonist (Maemoto *et al.*, 1997). Further structure-activity relationships with substitution in the 8-position of adenine afforded the isopropyl-methyl-amine moiety 8-(N-methyl isopropyl)amino-N6-(5'-endohydroxy-endonorbornyl)-9-methyladenine) WRC-0571 with improved A<sub>1</sub> AR affinity (Robeva *et al.*, 1996).

Research by van der Walt and co-workers (2013) showed that by introducing aryl-substituents at position C8, the xanthine structure resulted in improved A<sub>1</sub> AR affinity for (8-(3-phenylpropyl)-1,3,7-triethylxanthine.



## 3.4.2.3 Tricyclic Fused Heteroaromatic System

Tricyclic fused heteroaromatic rings are usually composed of 6:6:5 fused, 6:5:6 fused and 5:6:5 fused rings with different number and arrangement of nitrogen atoms (Yuzlenko & Kiec-Kononowicz, 2006). Most 6:6:5 fused N-heteroaromatic systems have similarities in the number of ring arrangements, and favourable cyclopentyl group presence (Chang *et al.*, 2004).

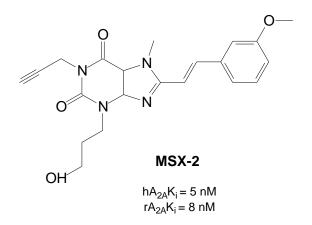


**5-amino-[1,2,4]triazolo-[5,1-f]purin-2-one** hA<sub>1</sub>K<sub>i</sub> = 1.5 nM

#### 3.4.3 Xanthine and Non-xanthine A<sub>2A</sub> AR Antagonists

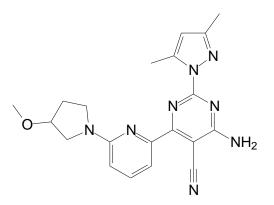
Based on the core xanthine structure some  $A_{2A}$  AR antagonists present good affinity, with some of these  $A_{2A}$  antagonists undergoing clinical trials and only one drug approved for the adjunctive treatment of PD (Yuzlenko & Kiec-Kononowicz, 2006).

The first highly selective A<sub>2A</sub> ligands were 8-styryl derivatives e.g. MSX-2 which contains a propargyl group. 3-(3-Hydroxypropyl)-8-(m-methoxystryl)-7-methyl-1-propargyl xanthine MSX-2 further showed high affinity in radioligand binding assays at the human recombinant A<sub>2A</sub> AR (Cacciari *et al.*,2003; Yuzlenko & Kiec-kononowicz, 2006).



#### 3.4.3.1 Monocyclic Fused Heteroaromatic System

The monocyclic  $A_{2A}$  antagonist (R)-4-amino-2-(3,5-dimethyl-1H-pyrazol-1-yl)-6-(6-(3-methoxypyrrolidin-1-yl)pyridin-2-yl)pyrimidine-5-carbonitrile showed high affinity against  $A_{2A}$  ARs and emerged as potential treatment for PD (Yang *et al.*, 2016).

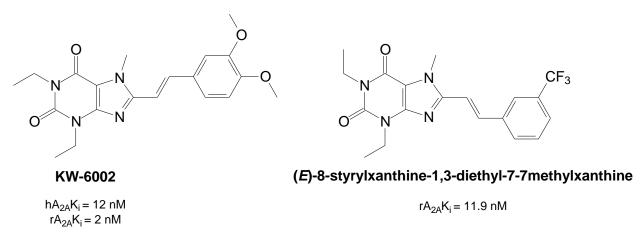




 $hA_{2A}K_{i} = 1.0 \text{ nM}$ 

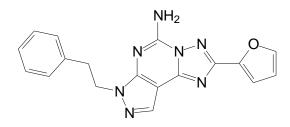
#### 3.4.3.2 Bicyclic Fused Heteroaromatic System

KW-6002, reported as the most advanced A<sub>2A</sub> antagonist, received marketing approval in Japan under the name NOURIAST<sup>™</sup> in 2013 and was also recently approved by the United States FDA. Exploration of the 1,3-diethyl-8-(3,4-dimehtoxystryryl)-7-methylxanthine scaffold which is substituted by two ethyl groups on the xanthine nucleus (Zheng *et al.*, 2014; Hockemeyer *et al.*, 2004; Van der Walt *et al.* 2013), led to the discovery of the structurally related (*E*)-1,3-diethyl-7methyl-8-[(3-trifluoromethyl)styryl]xanthine which also showed high binding affinity against rat striatal A<sub>2A</sub> ARs.



## 3.4.3.3 Tricyclic Fused Heteroaromatic System

[1,2,4]triazolo[5,1-f]purin-2-one derivatives, synthesized by Basu and co-workers (2017), showed significant affinity for  $A_{2A}$  ARs and proved to be good therapeutical drugs for the potential future treatment of PD. Additionally, a series of 5-amino-2-furylopyrazolotriazolo-pyrimidines showed good affinity and selectivity for  $A_{2A}$  ARs, but displayed poor water solubility (Baraldi *et al.* 2002). Interestingly, the report by Lopes and co-workers. (1999) on another tricyclic fused heteroatomic ring system namely SCH-58261 clearly showed how the intraperitoneal injection of either caffeine or the aforementioned selective  $A_{2A}$  AR antagonist (SCH-58261) prevent learning deficit and the expression of A $\beta$  toxicity, which may be helpful in AD.

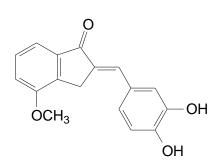


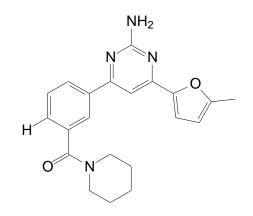
#### SCH 58261

 $hA_{2A}K_i = 1.1 nM$  $rA_{2A}K_i = 1.1 nM$ 

#### 3.4.4 Dual Target A<sub>1</sub>/A<sub>2A</sub> AR Antagonists

Despite its low affinity in both A<sub>1</sub> and A<sub>2A</sub> ARs, the effects of caffeine and coffee consumption are significant in patients with AD and PD, as seen in different studies (Jacobson *et al.*, 1993, Kim *et al.*, 2014, Rodrigues *et al.*, 2015). Recent research on structure-activity relationships of flavonoids and structurally related compounds, lead to the discovery of dual A<sub>1</sub> and A<sub>2A</sub> AR antagonists of the benzylidene indanone family with high affinity in the nanomolar range (Janse van Rensburg *et al.*, 2020, Van Der Walt & Terre'Blanche, 2015). Another dual compound was 4-(5-methylfuran-2-yl)-6-[3-(piperidine-1-carbonyl)phenyl]pyrimidin-2-amine which displayed high affinity with low  $K_i$  values against A<sub>1</sub>/A<sub>2A</sub> ARs in rat brain membranes (Robinson *et al.*, 2015).





2-(3,4-dihydroxybenzylidene)-4-methoxy-2,3-dihydro-1H-inden-1-one

 $rA_1K_i = 42 \text{ nM}$  $rA_{2A}K_i = 78 \text{ nM}$ 

4-(5-methylfuran-2-yl)-6-[3-(piperidine-1carbonyl)phenyl]pyrimidin-2-amine

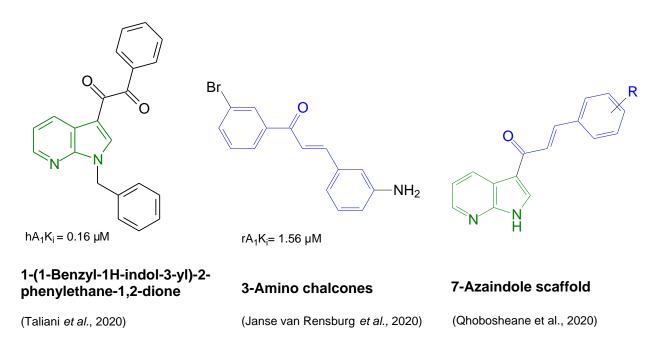
> rA<sub>1</sub>K<sub>i</sub> = 9.54 nM rA<sub>2A</sub>K<sub>i</sub> = 6.34 nM

## 3.4.5 Chalcone Based Adenosine Antagonists

Chalcones are potent organic scaffolds that forms part of the flavonoid group. Chalcones are composed of two aromatic rings that are linked by a three carbon  $\alpha$ , $\beta$ -unsaturated carbonyl system and are considered a privileged scaffold in medicinal chemistry. They have attracted attention not only from synthetic and biosynthetic perspectives but also due to their wide-ranging biological activities, such as antimicrobial, antiviral, antifungal, antimalarial, antileishmanial, anti-inflammatory and anticancer properties (Gaonkar & Vignesh, 2017; Zhuang *et al.*, 2017). Chalcones are a member of the flavonoid family which have been explored as AR antagonists (Van der Walt & Terre'Blanche, 2018). Chalcones have also showed potential in the treatment of neurological conditions by acting as antidepressants and anxiolytics, as well as the inhibition of acetylcholinesterase (AChE), butyrylcholinesterase (BChE) and MAO (Mathew *et al.*, 2019). In addition, several chalcone-coumarin derivatives have shown to possess AR affinity (Vazquez-Rodriguez *et al.*, 2013; Vazquez-Rodriguez *et al.*, 2020) and recently C3 amino-substituted

chalcone derivatives with a bromo substitution on benzylidene ring B were reported to possess selective adenosine  $A_1$  AR affinity in the micromolar range (Janse van Rensburg, *et al.*, 2020).

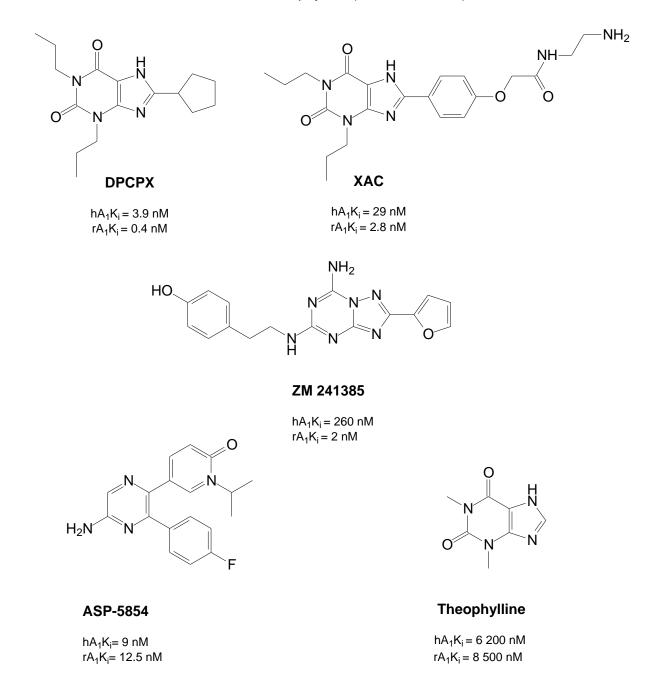
Compounds structurally related to chalcones include benzylidene tetralones, benzylidene indanones, aurones, benzoyl benzofurans, as well as isoxazole and pyrazole derivatives. Due to the open-chain model and scaffold modification of chalcones, a new class of organic compounds such as isoxazole-, pyrazole- and indole-based chalcones was synthesized. In a recent review by Taliani and co-workers (2020) they exploited the indole scaffold to design compounds binding to different pharmacological targets and highlighted. the affinity of 1-benzyl-3-ketoindole derivatives with 1-(1-Benzyl-1*H*-indol-3-yl)-2-phenylethane-1,2-dione exhibiting submicromolar affinity for the A<sub>1</sub> AR of 161 nM. Qhobosheane and co-workers (2020a) synthesized novel C3 substituted chalcone-based derivatives of 7-azaindole and evaluated their protein kinase inhibition activity. Due to the structural similarity of the 7-azaindoles compared to chalcones and indoles it was decided to screen these compounds for novel A<sub>1</sub> AR affinity.



### 3.5 Species Difference in Adenosine Receptor Affinity

Preclinical studies are mostly performed in mice or rats. The A<sub>1</sub> AR of human, rat, and mouse comprises of 326 amino acids and the similarity of the amino acid sequence of the three species is as follows: human vs. rat 95%, human vs. mouse 95%, and rat vs. mouse 98% (Alnouri *et al.*, 2015). In a study by Maemoto and co-workers (1997) xanthine based AR antagonists exhibited higher A<sub>1</sub> affinity for [<sup>3</sup>H]DPCPX binding sites in rat cortical membranes, compared with human brain membranes. In contrast to xanthine antagonists, pyrazolopyridine derivatives displayed similar affinity for [<sup>3</sup>H]DPCPX in both rat and human membranes.

Supporting these findings Fredholm and co-workers (2001) compared literature values of xanthine antagonists showing the following: DPCPX (human 3.9 nM vs rat 0.3 nM), XAC (human 29 nM vs rat 2.8 nM) ZM 241385 (human 260 nM vs rat 2 nM). Non-xanthine AR antagonist, ASP-5854 (5-[5-Amino-3(4-fluorophenyl) pyrazin-2yl]-1-isopropylpyridine-2(1H)-one), further showed similar binding affinity for A<sub>1</sub> ARs in humans, rats, and mice (Mihara *et al.*, 2007). In addition, Klotz and co-workers (1991) also reported xanthine derivatives DPCPX and XAC to be 10-30 times less potent at the human A<sub>1</sub> AR compared to the rat A<sub>1</sub> AR. Interestingly no significant species difference was found for the xanthine, theophylline (Klotz *et al.* 1991).



Regardless of the nature of A<sub>1</sub> AR from different species, above mentioned research provides a caution against extrapolating profiles of potencies for AR antagonist from one species to another.

#### 3.6 Summary

In this chapter, we highlighted the importance and the physiological role of  $A_1$  and  $A_{2A}$  ARs in neurological diseases such as AD and PD, together with the positive effects of the xanthine derivative caffeine on motor and non-motor (cognitive and depression) dysfunctions in PD.  $A_1$  and  $A_{2A}$  AR antagonists were explored as rational drug treatment for AD and PD, and dual  $A_1/A_{2A}$  antagonists may offer a solution in alleviating motor symptoms, depression and cognitive dysfunction and provide neuroprotection simultaneously. The affinity of the AR antagonists is very important and shows the potential of the compound as a drug; however, drug affinity tests done only on rat brain models are not ideal for a number of reasons. As highlighted in chapter 1, there is a strong opposition and debate among the scientific community about the relevance of animal data for humans and the likelihood of successful cross-species translation. The current pilot study will aid in establishing a radioligand binding assay to determine  $A_1$  AR affinity using CHO cells transfected with human  $A_{2A}$ ,  $A_{2B}$  and  $A_3$  ARs in order to test compounds against all the adenosine subtypes. In addition, this study also aims to identify the 7-azaindole scaffold as a promising target for the development of novel compounds for AR affinity.

# CHAPTER 4: RADIOLIGAND BINDING ASSAY USING CHO CELLS EXPRESSING HUMAN ADENOSINE A1 RECEPTORS

## 4.1 Introduction

The  $A_1$  AR radioligand binding assays provide a means for assessing the degree of binding affinity that selected test compounds may possess toward the  $A_1$  ARs. Typically, a radioligand binding study is conducted using proteins that express the desired receptor in the presence of a radioligand known to exhibit a high affinity towards the receptor in question.

RBAs are based upon competitive binding between an analyte and a radiolabelled ligand at a certain target receptor. By using this method, the amount of the ligand bound to a target receptor (ligand-receptor-complex) can be measured in order to determine the radiolabelled ligand's binding affinity for certain receptors, after which the degree of selectivity for the receptor can be calculated by comparing various affinities of receptor subtypes. Any receptor can be used to perform RBAs as long as it has a selective ligand which can be labelled with radioactive isotopes. Currently, determination of adenosine receptor affinity is mainly accomplished by performing RBAs with ligands radiolabelled with either tritium or iodine

Competition binding experiments determines the relative affinity of an unlabelled compound or drug for its target receptor. During these experiments a single concentration of a radioligand is measured in the presence of increasing concentrations of unlabelled compounds with the same target receptor during a state of equilibrium. The concentration of bound radioligand wil decrease as the concentration of the unlabelled ligand increases. The affinity constant ( $K_i$ ) is determined, and indicates the affinity of the unlabelled compound or drug for its target receptor. Further, the radioligands bound to receptors and those free of receptors are separated by the use of filters after incubation, for example a Hoffeler filtration system (Van der Walt, *et al.*, 2013). An acceptable amount of ligand needs to be bound to their receptors in order to prevent losing too much after the unbound ligand are washed away during the process of separation. In order to achieve this, the ligand has to be given an adequate amount of time to bind with their respective receptors. Liquid scintillation counting is used to count the bound radioactivity trapped in the filters after placing the filters in scintillation fluid. For the present study, a Packard Tri-Carb 2100 TR liquid was used, and the scintillation count expressed as counts per minute (CPM).

All cell culture work and radioligand binding assays were done at the North-West University's Laboratory for Analytical and Molecular Biology (LAMB). Furthermore, all materials and reagents were commercially available and purchased from various manufacturers. The Chinese hamster

ovary (CHO) cells expressing the human A<sub>1</sub> AR cells were kindly donated by Prof KN Klotz from the University of Würzburg, Germany.

# 4.2 Ethics

The use of CHO cells for the A<sub>1</sub> AR radioligand binding assay was approved by the Health Sciences Ethics Office for Research, Training and Support, North-West University, application number: NWU-00585-19-A5 (Annexure).

# 4.3 Cloning of Adenosine Receptor and Stable Transfection of Cells

Chen and Okayama (1987) were the first to describe the transfection protocol of mammalian cells by plasmid DNA, while Freund *et al.* (1994) clearly illustrated transfection of the rat  $A_1$  AR into CHO cells. CHO  $A_1$  cells were prepared at the University of Würzburg, Germany; specifically cloning of the human  $A_1$  AR and stable transfection of CHO cells cDNAs were verified according to GenBank entries after sequencing and comparison and found to correspond with other published sequences for the  $A_1$  AR (Klotz *et al.*, 1998).

# 4.4 Culturing of CHO A<sub>1</sub> AR Cells

First and foremost, all cell culture work was done in a sterile environment; all apparatus and equipment were always disinfected with 70% ethanol prior to use.

# 4.4.1 Apparatus and Equipment

- Tube racks
- Spray bottle with 70% ethanol
- Paper towels
- Centrifuge tubes (15 cm<sup>2</sup>/50 cm<sup>2</sup>)
- Cell culture flasks (25 cm<sup>2</sup>/75 cm<sup>2</sup>)
- Serological pipettes (5 mL; 10 mL)
- Waste containers (one for liquid waste, one for plastic waste)
- Pipettor
- Laminar flow hood
- CO<sub>2</sub> incubator
- Water bath
- Light microscope
- Centrifuge

# 4.4.2 Materials and Reagents

- Growth medium: Dulbecco's Modified Eagles Medium with nutrient mixture F12 (DMEM/F12) (PAN Biotech) as well as added 10% Fetal Bovine Serum (FBS) (Gibco), 1% L-Glutamine (Lonza), 1% Non-Essential Amino Acids (NEAA) (Lonza), 1% Penicillin-Streptomycin (Pen-Strep) (Lonza), 0.2mg/mL (0.4%) Geneticin (Gibco)
- Phosphate buffered saline (PBS) (HyClone)
- Trypsin-Versene<sup>™</sup>-Trypsin-EDTA Cell Culture Reagent (Lonza)

# 4.5 Procedure for Culturing

This procedure is valid for the routine culturing of CHO  $A_1$  cells in 25 and 75 cm<sup>2</sup> culture flasks donated from the University of Würzburg, Germany as described by Klotz *et al.* (1998). CHO  $A_1$  cells between passages 4 and 14 were used for the experiments.

# 4.5.1 Reviving Frozen Cell Stocks

The following procedure is effective for the standard revival of CHO  $A_1$  cells after storage at - 150°C for long periods:

The cryovial was removed from the cryogenic refrigerator and rapidly thawed at 37 °C in the water bath, and once thawed 60–80% (presenting a slush), disinfected and placed in the laminar flow hood. (Please note that the cap of the cryovial must never be submerged in the water bath.) To

the thawed cryovial containing the selected cell stock, 1 mL preheated growth medium was slowly added, and subsequently, transferred to a sterile 15 cm<sup>2</sup> tube already containing 4 mL growth medium. The cell-medium mixture was then centrifuged at 140 *g* for 5 minutes; the resulting supernatant was removed by pipetting. The pellet was re-suspended in 5 mL growth medium. The cell-medium mixture was transferred to a sterile 25 cm<sup>2</sup> cell culture flask (taking care to distribute the said mixture evenly over the growth surface) and placed in the CO<sub>2</sub> incubator overnight. The following day, attachment of cells to the growth surface was determined using a light microscope. The spent growth medium was removed from the cell culture flask by pipetting and new growth medium was added to and distributed in the flask, and then, the cell culture flask was returned to the CO<sub>2</sub> incubator.

- For a 25 cm<sup>2</sup> cell culture flask, 5 mL growth medium was added
- For a 75 cm<sup>2</sup> cell culture flask, 15 mL growth medium was added

# 4.5.2 Subculturing Cells

Of note, CHO  $A_1$  cells were subcultured between 80–90% confluences. Again, the spent growth medium was removed from the cell culture flask by pipetting. The flask was then rinsed twice with preheated PBS.

- For a 25 cm<sup>2</sup> cell culture flask, 5 mL PBS was added twice
- For a 75 cm<sup>2</sup> cell culture flask, 10 mL PBS was added twice

Subsequently, preheated Trypsin-Versene<sup>™</sup> was added to and distributed evenly in the flask and the flask was then promptly returned to the CO<sub>2</sub> incubator at 37 °C for 2 minutes. Thereafter, the flask was removed from the incubator and tapped in order to detach the cells from the growth surface.

- For a 25 cm<sup>2</sup> cell culture flask, 1 mL Trypsin-Versene<sup>™</sup> was added
- For a 75 cm<sup>2</sup> cell culture flask, 3 mL Trypsin-Versene<sup>™</sup> was added

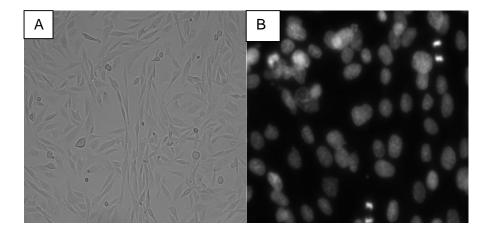
Once the cells were detached, preheated growth medium was added to the flask.

- For a 25 cm<sup>2</sup> cell culture flask, 5 mL growth medium was added
- For a 75 cm<sup>2</sup> cell culture flask, 6 mL growth medium was added

The cell-medium mixture was transferred to a sterile  $15 \text{ cm}^2$  tube and centrifuged at 140 *g* for 5 minutes; the resulting supernatant was removed by pipetting. The pellet was re-suspended in 5 mL preheated growth medium, and upon mixing, the cell suspension was divided between the appropriate number of 25 cm<sup>2</sup> and/or 75 cm<sup>2</sup> cell culture flasks. (Subculture in a ratio between 1:8

and 1:20. A subculture ratio of 1:20 takes about 1 week to reach 80% confluence). Preheated growth medium was then added to each flask to obtain the desired final volume. Finally, the cell culture flasks were put in the  $CO_2$  incubator.

- Final volume of growth medium for a 25 cm<sup>2</sup> cell culture flask is 5 mL
- Final volume of growth medium for a 75 cm<sup>2</sup> cell culture flask is 15 mL



- Figure 4-1: Illustrates the (A) CHO  $A_1$  cells at 50-60 % confluence. (B) Clean CHO  $A_1$  growth without mycoplasma contamination.
- 4.6 Membrane Storage

# 4.6.1 Apparatus and Equipment:

- Tube racks
- Spray bottle with 70% ethanol
- Paper towels
- Centrifuge tubes (15 cm<sup>2</sup>/50 cm<sup>2</sup>)
- Cell culture flasks (25 cm<sup>2</sup>/75 cm<sup>2</sup>)
- Serological pipettes (5 mL; 10 mL)
- Waste containers (one for liquid waste, one for plastic waste)
- Pipettor
- Cell scraper
- Laminar flow hood
- CO<sub>2</sub> incubator
- Water bath
- Light microscope
- Centrifuge

• Freezer

# 4.6.2 Materials and Reagents

- PBS
- 50 mM Tris/HCI (pH 7.7 at 25 °C)

# 4.6.3 Method

For CHO A<sub>1</sub> cells between 80-90% confluences, the spent growth media was removed from the cell culture flask by pipetting. The flask was then rinsed twice with preheated PBS.

- For a 25 cm<sup>2</sup> cell culture flask, 5 mL PBS was added twice
- For a 75 cm<sup>2</sup> cell culture flask, 10 mL PBS was added twice

The cells attached to the growth surface of the cell culture flask were then scraped off in ice-cold hypotonic buffer and transferred to a sterile 50 cm<sup>2</sup> tube (repeated two times). These tubes were then frozen at -20 °C until day of membrane preparation.

- For a 25 cm<sup>2</sup> cell culture flask, 5 mL (50 mM Tris/HCI (pH 7.7 at 25 °C) was added twice
- For a 75 cm<sup>2</sup> cell culture flask, 10 mL (50 mM Tris/HCI (pH 7.7 at 25 °C) was added twice

# 4.7 Membrane Preparation

# 4.7.1 Apparatus and Equipment

- Centrifuge tubes (15 cm<sup>2</sup>/50 cm<sup>2</sup>)
- Serological pipettes (5 mL; 10 mL)
- Pipettor
- Dispersing instrument
- Centrifuge
- Freezer

# 4.7.2 Materials and Reagents

- 50 mM Tris/HCl buffer (pH 7.7 at 25 °C)(Merck)
- GF/B filters (Whatman®)
- Liquid scintillation counter (PerkinElmer)

# 4.7.3 Method

Membrane proteins were prepared by thawing frozen CHO A<sub>1</sub> cells on ice. The resulting cell suspension was homogenized on ice (2 x 15 s at full speed) and the homogenate was then spun for 10 min (4 °C) at 1,000 *g*. The pellet was discarded, and the supernatant was then centrifuged for 30 min at 100,000 *g*. The final membrane pellet was resuspended in 50 mM Tris/HCl buffer (pH 7.7 at 25 °C) at a protein concentration of 1.2 mg/ml and stored at -80 °C (Klotz *et al.*, 1998).

# 4.8 **Protein Concentration**

The protein concentration was determined according to the Bradford assay, using bovine serum albumin as reference standard (Bradford, 1976).

# 4.9 Adenosine A<sub>1</sub> Receptor Radioligand Binding Assays

The degree of binding affinity the test compounds showed toward the human  $A_1$  AR was determined via radioligand binding assays which were adapted from Klotz *et al.* (1998) and Van der Walt and Terre'Blanche (2015). The  $A_1$  AR radioligand binding assay used membrane proteins expressing the human  $A_1$  AR (see sections 4.1–4.6) (Klotz *et al.*, 1998) and 1,3-[<sup>3</sup>H]-dipropyl-8-cyclopentylxanthine ([<sup>3</sup>H]DPCPX) as radioligand (Bruns *et al.*, 1987).

# 4.9.1 Apparatus and Equipment

- Tube racks
- Microcentrifuge tubes (1.5 cm<sup>2</sup>)
- Centrifuge tubes (15 cm<sup>2</sup>/50 cm<sup>2</sup>)
- Polyvials
- GF/B filters
- Serological pipettes (5 mL; 10 mL)
- Pipettor
- Pippette tips (1000 μL, 100 μL, 10 μL, 1 μL)
- Pipettes (1000 µL, 100 µL, 10 µL, 1 µL)
- Hoffeler vacuum system
- Shaking water bath
- Liquid scintillation counter

# 4.9.2 Materials and Reagents

• Dimethyl sulfoxide (DMSO) (Sigma)

- 50 mM Tris/HCl buffer (pH 7.7 at 25 °C) (Merck)
- 1,3-[<sup>3</sup>H]-dipropyl-8-cyclopentylxanthine ([<sup>3</sup>H]DPCPX) (PerkinElmer)
- Adenosine deaminase (ADA)(Sigma)
- CHO A1 membrane proteins

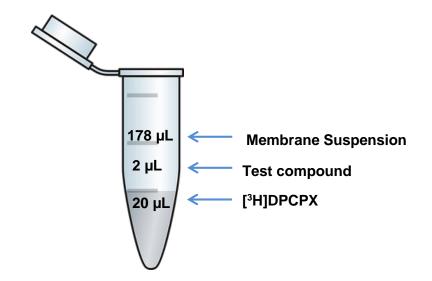


Figure 4-2: Illustration of 2 mL tube with final order of addition: (1) 20  $\mu$ L radioligand solution (containing [<sup>3</sup>H]DPCPX) (2) 2  $\mu$ L test compound (at the desired concentration ranging from 0  $\mu$ M to 100  $\mu$ M) and (3) 178  $\mu$ L membrane suspension (containing CHO A<sub>1</sub> membranes).

# 4.9.3 Method

Each incubation consisted of (in order of additions): (1) 20  $\mu$ L radioligand solution, (2) 2  $\mu$ L test compound (at the desired concentration ranging from 0  $\mu$ M to 100  $\mu$ M) and (3) 178  $\mu$ L membrane suspension. The radioligand solution had a concentration of 1 nM [<sup>3</sup>H]DPCPX (2) and the membrane suspension was equivalent to 20  $\mu$ g/200 mL membrane proteins (expressing the A<sub>1</sub> AR) and 0.2 units/mL adenosine deaminase (ADA). The final volume of all incubations contained 200  $\mu$ L 50 mM Tris/HCl buffer (pH 7.7 at 25 °C) and 1% DMSO (Van der Walt & Terre'Blanche, 2015). After the additions were made, each sample was vortexed and incubated in a shaking water bath at 25 °C for 1 hour. Thirty minutes, post-commencement of incubation, each sample was vortexed again and returned to the shaking water bath. Incubation was terminated by filtering each sample through 25 mm GF/B filter and the pertaining microcentrifuge tube was washed twice with 1 mL Tris/HCl buffer (pH 7.7 at 25 °C). Filtration occurred under reduced pressure using a Hoffeler vacuum system and upon completion each filter was placed into a scintillation vial, along with 4 mL scintillation fluid. The vials were subjected to agitation before being left in a stationary position for two hours. At that time, the residual radioactivity of each filter was measured with a

liquid scintillation counter. Non-specific binding of [ ${}^{3}$ H]DPCPX for the radioligand binding assay was defined as binding in the presence of 10  $\mu$ M DPCPX (Bruns *et al.*, 1987; Van der Walt *et al.*, 2015). Specific binding was defined as the total binding minus the non-specific binding (Van der Walt *et al.*, 2015).

#### 4.10 Statistical Data Analysis

Briefly, all statistical data analyses were done using Microsoft Excel and GraphPad Prism Software based on non-linear curve fitting procedures. Sigmoidal dose response curves, from which  $IC_{50}$  values were calculated, were obtained by plotting the specific binding of [<sup>3</sup>H]DPCPX against the logarithm of the test compounds' concentrations. Subsequently, the  $IC_{50}$  values were used to calculate the inhibition constant (K<sub>i</sub>) values for the competitive inhibition of [<sup>3</sup>H]DPCPX against membrane proteins by means of the Cheng-Prusoff equation, where [C<sup>\*</sup>] is the concentration of the radioligand and K<sub>d</sub> its dissociation constant (**Equation 4–1**). The equilibrium dissociation constant (K<sub>d</sub>) value for the radioligand [<sup>3</sup>H]DPCPX was taken as 3.86 nM (Klotz *et al.*, 1998). All calculated K<sub>i</sub> values were determined in triplicate and given as mean ± standard error of the mean (SEM).

# $K_i = IC_{50} / (1 + [C^*] / K_d)$ Equation 4-1

IC<sub>50</sub> test compound / (1 + [Concentration radioligand [<sup>3</sup>H]DPCPX] / K<sub>d</sub> value radioligand [<sup>3</sup>H]DPCPX)

Component	Function		
DMSO	Used as solvent for test compounds		
50 mM Tris.HCl	Served as buffer solution		
Scintillation fluid	Used to dissolve GF/B filters containing bound radioligand in order to measure radioactivity with a scintillation counter.		
A <sub>1</sub> AR membrane suspension	Contains CHO membranes expressing the $A_1$ AR as well as ADA.		
[ <sup>3</sup> H]DPCPX	Selective A <sub>1</sub> AR antagonist used as radioligand (Bruns <i>et al.</i> , 1987; Van der Walt & Terre'Blanche, 2015)		
ADA	Breakdown endogenous adenosine and prevents binding of endogenous adenosine to ARs (Bruns <i>et al.</i> , 1987)		

Table 4-1:Summary of A1 AR radioligand binding assay using CHO cells.

# **CHAPTER 5: RESULTS AND DISCUSSION**

# 5.1 Introduction

As mentioned earlier, the current pilot study will aid in establishing the radioligand binding assay using CHO cells transfected with the human  $A_1$  AR. This will enable future studies using CHO cells transfected with human  $A_{2A}$ ,  $A_{2B}$  and  $A_3$  ARs in order to test compounds against all the adenosine subtypes. In addition, this study also aimed to identify the 7-azaindole scaffold as a promising target for the development of novel compounds with AR affinity.

# 5.2 Establishing Standard Radioligand Binding Assay for CHO Cells Expressing the A<sub>1</sub> AR

The radioligand binding assay utilizing rat  $A_1$  ARs was adapted, as seen in **Table 5-1**, to establish the assay using CHO  $A_1$  cell membranes.

Parameters	Rat A <sub>1</sub> AR RBA	Human A <sub>1</sub> AR RBA	
Final volume	1000 µL	200 µL	
Membrane suspension	Rat whole brain: 120 μg/890 μL	СНО А₁ cell: 17.8 µg/178 µL	
	ADA: 0.1 units/890 μL	ADA: 0.04 units/178 μL	
Radioligand solution	[3H]DPCPX: 0.1 nM	[3H]DPCPX: 1 nM	
Solvent (DMSO)	1%	1%	
Buffer	1000 µL 50 mM Tris–HCI buffer	200 µL 50 mM Tris–HCI buffer	
	(pH 7.7 at 25 °C)	(pH 7.7 at 25 °C)	
Non-specific binding	10 µM DPCPX	10 µM DPCPX	
Tube for incubation:	5 mL round-bottom polypropylene	2 mL microcentrifuge tube	
	tubes		
Tubes rinsed 2 x with:	4 mL 50 mM Tris–HCl buffer	1 mL 50 mM Tris–HCl buffer	
	(pH 7.7 at 25 °C)	(pH 7.7 at 25 °C)	
Rinsed filter with:	4 mL 50 mM Tris–HCl buffer	4 mL 50 mM Tris–HCI buffer	
	(pH 7.7 at 25 C)	(pH 7.7 at 25 C)	
Scintillation fluid	4 mL	4 mL	
Performed in:	triplicate	triplicate	

Table 5 4.	Dedictionend hinding eccev perspectate utilizing either set or CLO A AD
Table 5-1:	Radioligand binding assay parameters utilizing either rat or CHO A <sub>1</sub> AR

In this study two reference compounds (DPCPX and KW-6002) were used to establish the method and the results are shown in **Table 5-2**. The radioligand binding assays were validated with

DPCPX ( $A_1$  antagonist) and istradefylline ( $A_{2A}$  antagonist) as reference compounds and results were in accordance with literature values.

As reported previously by Maemoto and co-workers (2004) as well as Bulicz and colleagues (2006), the present study also found that the affinity of reference compounds DPCPX and istradefylline have significantly reduced activity against human versus rat  $A_1$  ARs. Affinity at the human  $A_1$  AR binding of DPCPX is almost tenfold lower than rat whole brain membranes expressing the  $A_1$  AR. This phenomenon, is apparently, often observed with xanthine derivatives such as DPCPX and istradefylline (Maemoto *et al.*, 2004).

# Table 5-2:Inhibition constant ( $K_i$ ) values for the binding affinity of referencecompounds against rat (r) and human (h) A1 ARs

	K <sub>i</sub> ±S	K <sub>i</sub> ± SEM (μM) <sup>a</sup>		
Compound	rA <sub>1</sub> <sup>b</sup> vs [ <sup>3</sup> H]DPCPX <sup>c</sup>	hA <sub>1</sub> <sup>d</sup> vs [ <sup>3</sup> H]DPCPX <sup>e</sup>		
	(0.0005) <sup>f</sup>	0.004 ± 0.0002		
DPCPX (A1 antagonist)	(0.0004) <sup>g</sup>	(0.001) <sup>i</sup>		
	(0.0003) <sup>h</sup>	(0.003) <sup>j</sup>		
latradatulling (A., antagonist)	(0.000)k	1.7 ± 0.56		
Istradefylline (A <sub>2A</sub> antagonist)	(0.230) <sup>k</sup>	(0.841) <sup>I</sup>		

<sup>a</sup> All inhibition constant ( $K_i$ ) values were determined in triplicate and expressed as mean  $\pm$  standard error of the mean (SEM) in  $\mu$ M.

<sup>b</sup> Rat receptors were used (rA<sub>1</sub>)

° 0.1 nM [<sup>3</sup>H]DPCPX

<sup>d</sup> Human receptors were used (hA<sub>1</sub>).

<sup>e</sup> 1 nM [<sup>3</sup>H]DPCPX

<sup>f</sup> Literature value obtained from (Van der Walt & Terre'Blanche, 2015)

<sup>g</sup> Literature value obtained from (Janse van Rensburg et al., 2017)

<sup>h</sup> Literature value obtained from (Lohse et al., 1987)

<sup>i</sup> Literature value obtained from (Maemoto et al., 2004)

<sup>j</sup> Literature value obtained from (Bulicz et al., 2006)

<sup>k</sup> Literature value obtained from (Müller & Jacobson, 2011)

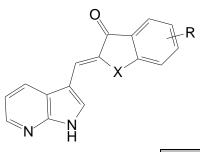
<sup>1</sup> Literature value obtained from (Müller & Jacobson, 2011)

Therefore, the current study demonstrated the reliable *in vitro* assessment of radioligand binding affinity in human A<sub>1</sub> AR transfected CHO cells. The already designed method of radioligand binding assays using transfected CHO cells, expressing A<sub>1</sub> AR has already been validated (Klotz *et al.*, 1998), and is now established at the NWU's LAMB as a tool to measure the binding affinity of novel compounds.

# 5.3 Novel 7-Azaindole-Chalcone Core Binding against A1 AR in Rat and Human

The degree of binding affinity that the test compounds showed toward human (h)  $A_1$  and rat (r)  $A_1$  ARs were determined via radioligand binding assays in triplicate (inhibition constant  $K_i$ ,  $\mu$ M) and expressed as mean ± standard error of the mean (SEM) (**Table 5-3**). The 7-azaindole derivatives under investigation were previously synthesized and evaluated as protein kinase inhibitors (Qhobosheane *et al.*, 2020a; Qhobosheane *et al.*, 2020b). Thirty-five 7-azaindole compounds (Qhobosheane *et al.*, 2020a; Qhobosheane *et al.*, 2020b) were screened (at concentrations of 100  $\mu$ M, 10  $\mu$ M and 1  $\mu$ M) and five compounds warranted full RBA against rat  $A_1$  ARs (**Table 5-3**). Thereafter, the affinity of these compounds were determined with the CHO (h)  $A_1$  AR assay and the inhibition constant ( $K_i$ ) value in  $\mu$ M was calculated by methods reported previously (Klotz *et al.*, 1998; Van der Walt & Terre'Blanche, 2015) (**Table 5-3**). These radioligand binding assays were carried out under the parameters shown in **Table 5-1**.

# Table 5-3:Inhibition constant ( $K_i$ ) values for the binding affinity of 7-azaindolederivatives against rat (r) and human (h) A1 ARs



			K <sub>i</sub> ± SEM (μM) <sup>a</sup>	
Compound	(X)	R	rA₁ <sup>ь</sup> vs [³H]DPCPX°	hAı <sup>d</sup> vs [³H]DPCPX <sup>e</sup>
1a	-Cinnamoyl		$0.90 \pm 0.07$	0.92 ± 0.13
1b	(CH <sub>2</sub> ) <sub>2</sub>	-	1.7 ± 0.12	1.9 ± 0.67
1c	(CH <sub>2</sub> ) <sub>2</sub>	6-OH	3.5 ± 0.28	2.7 ± 0.28
1d	(CH <sub>2</sub> ) <sub>2</sub>	6-OCH₃	1.9 ± 0.11	2.5 ± 0.26
1e	CH <sub>2</sub>	5,6-diOCH₃	1.1 ± 0.07	1.7 ± 0.43

<sup>a</sup> Inhibition constant ( $K_i$ ) values were determined in triplicate and expressed as mean ± standard error of the mean (SEM) in  $\mu$ M.

<sup>b</sup> Rat receptors were used (rA<sub>1</sub>)

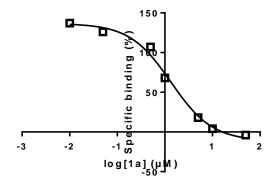
° 0.1 nM [<sup>3</sup>H]DPCPX

<sup>d</sup> Human receptors were used (hA<sub>1</sub>)

<sup>e</sup> 1 nM [<sup>3</sup>H]DPCPX

The competition binding curve (**Figure 5-1**), demonstrates the successful binding of compound **1a** to CHO (h) A<sub>1</sub> AR cells, showing the affinity of 7-azaindole-chalcone core by calculating the

concentration of 50% inhibition of radioligand binding ( $IC_{50}$ ) and using equation 4-1 to calculate affinity ( $K_i$ ).



# Figure 5-1: Typical dose-response curve for our *in vitro* model validation. The average binding-response curve for compound 1a on our human A<sub>1</sub> determined via a radioligand binding assay using CHO cell membranes expressing the human A<sub>1</sub> AR with [<sup>3</sup>H]DPCPX as radioligand.

The evaluation of 7-azaindole conjugated (chalcone/tetralone/indanone) scaffolds showed promising affinity for A<sub>1</sub> ARs in both rat and human species in the low micromolar range. Compound **1a** recorded the best A<sub>1</sub> AR affinity among the test compounds with rat A<sub>1</sub>  $K_i$  value of 0.90 µM and a human A<sub>1</sub> $K_i$  value of 0.92 µM, showing similar affinities in rat compared to human A<sub>1</sub> ARs (**Table 5-3**). This is also consistent with the nature of non-xanthine derivatives and previous reports by Maemoto and co-workers (2004), that non-xanthine ligands exhibit similar affinity for [<sup>3</sup>H]DPCPX binding sites in the brain membranes of different species – including rat and human. The second best affinity was the 7-azaindole-indanone derivative **1e** with dimethoxy substitution on the fused 6- and 5-membered rings, showing A<sub>1</sub> AR affinity of 1.1 µM in rat and a  $K_i$  value of 1.7µM in human A<sub>1</sub> ARs (CHO cells) (**Table 5-3**). The same trend was observed with similar affinities in rat compared to human A<sub>1</sub> ARs for compounds **1b**, **1c** and **1d**. The 1-indanone scaffold has also been reported by Janse van Rensburg and colleagues (2019a, 2019b), showing moderate affinity towards A<sub>1</sub> ARs.

Three compounds (**1b**, **1c**, **1d**) comprising of a conjugated 7-azaindole- $\alpha$ -tertralone structure were compared to **1a** with a conjugated 7-azaindole-chalcone structure and **1e** with a conjugated 7-azaindole-indanone structure. Compounds **1b** (unsubstituted tetralone), **1c** (hydroxyl substituted tetralone) and **1e** (methoxy substituted tetralone) showed a two- to three-fold decrease in affinity for both the rat and human A<sub>1</sub> ARs (**Table 5-3**) compared to **1a** and **1e**. The  $\alpha$ -tetralone scaffold has also been shown to possess affinity for the A<sub>1</sub> ARs (**Janse van Rensburg** *et al.*, 2017; Legoabe *et al.*, 2018). However, seeing that the difference between K<sub>i</sub> values are

small and only a limited number of derivatives were evaluated, these structure-activity relationships should be seen as preliminary. In general, the conjugated 7-azaindole-chalcone derivative **1a** showed better affinity than the indanone (**1e**) and tetralone (**1b**, **1c**, **1d**) derivatives. These limited results show that the 7-azaindole moiety may hold promise in the design of  $A_1$  AR ligands; where the 7-azaindole moiety is conjugated with a chalcone, tetralone or indanone moiety (**Figure 5-2**).

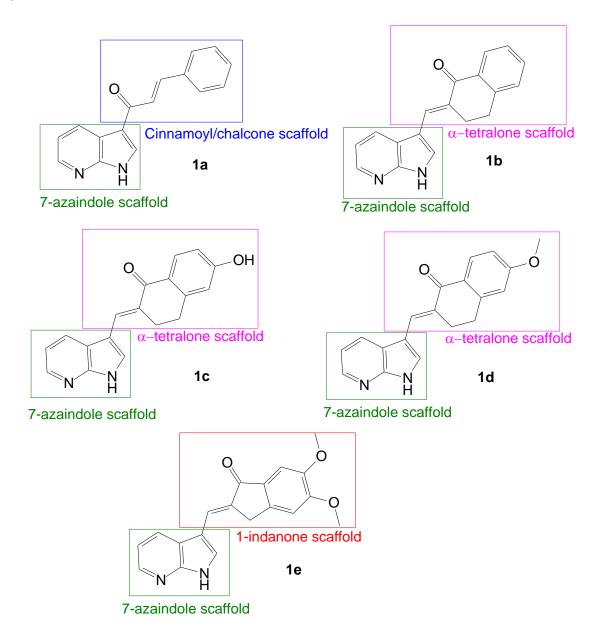


Figure 5-2: Structural relationships of 7-azaindole derivatives containing either a chalcone, tetralone or indanone moiety.

# **CHAPTER 6: CONCLUSION**

Animal research played a vital role in scientific and medical research the past century and continues to aid our understanding of various diseases and the development of new drugs. Understanding how humans respond to drugs is an essential question in drug research and often this cannot be addressed because it is not possible to perform some experiments in human volunteers. An alternative is usually to generate responses in animal models and translating it to humans.

RBAs are widely used for the screening of new potential receptor ligands and are ideally suitable for structure-activity relationship (SAR) analysis and molecular modelling studies. The adenosine A<sub>1</sub> RBAs are routinely performed with rat whole brain membranes that express the adenosine A<sub>1</sub> receptor subtype (Van der Walt & Terre'Blanche, 2015). However, using rat membranes are costly and the use of animals in scientific and therapeutic research has been a subject of hot discussion for many years. There are strong opposing opinions among enthusiasts and sceptics about the relevance of animal data for humans and the likelihood of successful cross-species translation.

Furthermore, cell-culture based tests have considerably reduced the use of rodents in the initial screening of potential new drugs, while speeding up the process so that more compounds can be screened in the same period of time. Prompted by the above discussions, the current pilot study undertook to establish the A<sub>1</sub> AR RBA using CHO cells expressing human A<sub>1</sub> ARs. Reference compounds DPCPX and istradefylline (KW-6002) were included for comparison with literature values. Further, five known 7-azaindole hybrids, previously evaluated as potential protein kinase inhibitors (Qhobosheane, *et al.*, 2020b), were screened as novel adenosine A<sub>1</sub> AR antagonists in the rat (unpublished results) and then selected to be evaluated as A<sub>1</sub> AR antagonists in CHO cells. The A<sub>1</sub> AR radioligand binding assay, using transfected CHO cells, was successfully established and affinity values of reference compounds correlated with literature values studies, thus showing that the above radioligand binding assay is reliable and reduced the challenges in ethics by minimizing the harvesting of rat brains and high costs.

Since the discovery of AR and their connection to various neurodegenerative diseases, the development of xanthine analogues has shown shortcomings of poor oral bioavailability, Bloodbrain barrier (BBB) penetration and species differences. On the other hand, non-xanthine antagonists display similar affinity for  $A_1$  ARs in both rat and human  $A_1$  AR membranes. The five non-xanthine 7-azaindole derivatives displayed  $A_1$  AR affinity in the low micromolar range and showed similar affinity for both rat and human  $A_1$  AR membranes.

The 7-azaindole-chalcone conjugated compound **1a** showed the highest binding affinity for A<sub>1</sub> ARs in rat and human (hA<sub>1</sub>K<sub>i</sub> = 0.92  $\mu$ M; rA<sub>1</sub>K<sub>i</sub> = 0.90  $\mu$ M). The 7-azaindole-indanone conjugated compound **1e** showed the second highest affinity (hA<sub>1</sub>K<sub>i</sub> = 1.7  $\mu$ M; rA<sub>1</sub>K<sub>i</sub> = 1.1  $\mu$ M). The 7-azaindole- $\alpha$ -tetralone fused compounds (**1b**, **1c**, **1d**) showed a two- to three-fold lower binding affinity for A<sub>1</sub> ARs in rat and human.

In conclusion, the  $A_1$  AR radioligand binding assay, using transfected CHO cells, was successfully established and affinity values of reference compounds correlated well with the results in literature. It also reduced challenges in ethics by minimizing the harvesting of rat brains and high costs. Although the compounds were limited for extensive SARs, the 7-azaindole-chalcone core appears to be the most promising lead for the development of compounds with improved  $A_1$  AR affinity with regards to improved cognitive deficits associated with neurodegenerative diseases.

# BIBLIOGRAPHY

Agostinho, P., Cunha, R.A. & Oliveira, C. 2010. Neuroinflammation, oxidative stress and the pathogenesis of Alzheimer's disease. *Current pharmaceutical design*, 16(25):2766-2778.

Ahlskog, J.E. & Muenter, M.D. 2001. Frequency of levodopa-related dyskinesias and motor fluctuations as estimated from the cumulative literature. *Journal of movement disorders*, 16(3):448-458.

Akiyama, H., Barger, S., Barnum, S., Bradt, B., Bauer, J., Cole, G.M., Cooper, N.R., Eikelenboom, P., Emmerling, M. & Fiebich, B.L. 2000. Inflammation and Alzheimer's disease. *Neurobiology of aging*, 21(3):383-421.2016.

Alam, Q., Alam, M.Z., Mushtaq, G., Damanhouri, G.A., Rasool, M., Kamal, M.A. & Haque A. 2016. Inflammatory Process in Alzheimer's and Parkinson's Diseases: Central Role of Cytokines. *Current pharmaceutical design*, 22(5):541-548.

Allen, M., Zou, F., Chai, H.S., Younkin, C.S., Miles, R., Nair, A.A., Crook, J.E., Pankratz, V.S., Carrasquillo, M.M. & Rowley, C.N. 2012. Glutathione S-transferase omega genes in Alzheimer and Parkinson disease risk, age-at-diagnosis and brain gene expression: an association study with mechanistic implications. *Molecular Neurodegeneration*, 7(1):13-24.

Almeida, L., Vaz-da-Silva, M., Silveira, P., Falcão, A., Maia, J., Loureiro, A., Torrão, L., Machado, R., Wright, L. & Soares-da-Silva, P. 2004. Pharmacokinetic–Pharmacodynamic Interaction Between BIA 3-202, a Novel COMT Inhibitor, and Levodopa/Carbidopa. *Clinical Neuropharmacology*, 27(1):17-24.

Amor, S., Peferoen, L.A., Vogel, D.Y., Breur, M., van der Valk, P., Baker, D. & van Noort, J.M. 2014. Inflammation in neurodegenerative diseases–an update. *Immunology*, 142(2):151-166.

Alnouri, M.W., Jepards, S., Casari, A., Schiedel, A.C., Hinz, S. & Muller, C.E. 2015. Selectivity is species-dependent: Characterization of standard agonists and antagonists at human, rat, and mouse adenosine receptors. *Purinergic Signal*, 11(3):389-407.

Ambrósio, A.F., Malva, J.O., Carvalho, A.P. & Carvalho, C.M. 1997. Inhibition of N-, P/Q- and other types of Ca2+ channels in rat hippocampal nerve terminals by the adenosine A1 receptor. *European journal of pharmacology*, 340:301–310.

Angulo, E., Casadó, V., Mallol, J., Canela, E.I., Viñals, F., Ferrer, I., Lluis, C. & Franco, R. 2003. A1 Adenosine Receptors Accumulate in Neurodegenerative Structures in Alzheimer's

Disease and Mediate Both Amyloid Precursor Protein Processing and Tau Phosphorylation and Translocation. *Brain Patholology*, 13(4):440-451.

Arendash G.W., Mori,T., Cao C., Mamcarz, M., Runfeldt, M., Dickson, A., Rezai-Zadeh, K., Tane, J., Citron, B.A., Lin, X., Echeverria, V. &, Potter, H. 2009. Caffeine reverses cognitive impairment and decreases brain amyloid-beta levels in aged Alzheimer's disease mice. *Journal of Alzheimer's Disease*, 17(3):661–680.

Armentero, M.T., Pinna, A., Ferré, S., Lanciego, J.L., Müller, C.E. & Franco, R. 2011. Past, present and future of A2A adenosine receptor antagonists in the therapy of Parkinson's disease. *Pharmacology & therapeutics*, 132:280-299.

Ascherio, A., Zhang, S.M., Hernán, M.A., Kawachi, I., Colditz, G.A., Speizer, F.E. & Willett, W.C. 2001. Prospective study of caffeine consumption and risk of Parkinson's disease in men and women. *Annals of neurology*, 50(1):56-63.

Azdad, K., Gall, D., Woods, A.S., Ledent, C., Ferré, S. & Schiffmann, S.N. 2009. Dopamine D 2 and adenosine A 2A receptors regulate NMDA-mediated excitation in accumbens neurons through A 2A–D 2 receptor heteromerization. *Neuropsychopharmacology*, 34(4):972.

Ball, P. & Knuppen, R. 1980. Catecholoestrogens (2-and 4-Hydroxyoestrogens). *European Journal of endocrinology*, 92(4):1-127.

Basma, A.N., Morris, E.J., Nicklas, W.J. & Geller, H.M. 1995. L-dopa cytotoxicity to PC12 cells in culture is via its autoxidation. *Journal of neurochemistry*, 64(2):825-832.

Baraldi, P., Tabrizi, M.A., Gessi, S. & Borea, P.A. 2008. Adenosine receptor antagonists: Translating medicinal chemistry and pharmacology into clinical utility. *Chemical reviews*, 108(1):238–263.

Baraldi, P.G., Cacciari, B., Romagnoli, R., Spalluto, G., Monopoli, A., Ongini, E., Varani, K., Borea, P.A. 2002. 7-Substituted 5-amino-2-(2-furyl)pyrazolo[4,3-e]-1,2,4-triazolo[1,5c]pyrimidines as A2A adenosine receptor antagonists: A study on the importance of modifications at the side chain on the activity and solubility. *Journal of medicinal chemistry*, 45(1):115–126.

Bassil, N. & Grossberg, G.T. 2009. Novel regimens and delivery systems in the pharmacological treatment of Alzheimer's disease. *CNS drugs*, 23(4):293-307.

Belaidi, A.A. & Bush, A.I. 2016. Iron neurochemistry in Alzheimer's disease and Parkinson's disease: targets for therapeutics. *Journal of neurochemistry*, 139 Suppl 1:179-197.

Borovac, J.A. 2016. Side effects of a dopamine agonist therapy for Parkinson's disease: a mini-review of clinical pharmacology. *The Yale journal of biology and medicine*, 89(1):37-47.

Braak, H., Alafuzoff, I., Arzberger, T., Kretzschmar, H. & Del Tredici, K. 2006. Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathology*, 112(4):389-404.

Bradford, M.M. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1-2):248-254.

Brambilla, R., Cottini, L., Fumagalli, M., Ceruti, S., Abbracchio, M. P. 2003. Blockade of A2A adenosine receptors prevents basic fibroblast growth factor-induced reactive astrogliosis in rat striatal primary astrocytes. *Glia*, 43:190–194.

Brothers, H.M., Marchalant, Y. & Wenk, G.L. 2010. Caffeine attenuates lipopolysaccharideinduced neuroinflammation. *Neuroscience Letters* 480:97–100.

Briyal, S., Nguyen, C., Leonard, M. & Gulati, A. 2015. Stimulation of endothelin B receptors by IRL-1620 decreases the progression of Alzheimer's disease. *Neuroscience*, 301:1-11.

Brocks, D.R. 1999. Anticholinergic drugs used in Parkinson's disease : an overlooked class of drugs from a pharmacokinetic perspective. *Journal of pharmaceutical sciences*, 2:39-46.

Bruns, R.F., Fergus, J.H., Badger, E.W., Bristol, J.A., Santay, L.A., Hartman, J.D., Hays, S.J. & Huang, C.C. 1987. Binding of the A 1-selective adenosine antagonist 8-cyclopentyl-1, 3dipropylxanthine to rat brain membranes. *Naunyn-Schmiedeberg's archives of pharmacology*, 335(1):59-63.

Bruns, R.F., Lu, G.H. & Pugsley, T.A. 1986. Characterization of the A2 adenosine receptor labeled by [3H]NECA in rat striatal membranes. *Molecular Pharmacology*, 29(4):331-346.

Bulicz, J., Bertarelli, D.C., Baumert, D., Fülle, F., Müller, C.E. & Heber, D. 2006. Synthesis and pharmacology of pyrido [2, 3-d] pyrimidinediones bearing polar substituents as adenosine receptor antagonists. *Bioorganic & medicinal chemistry*, 14(8):2837-2849.

Burnstock, G. 2006. Pathophysiology and therapeutic potential of purinergic signaling. *J Pharmacological reviews*, 58(1):58-86.

Burnstock, G. 2017. Purinergic signalling and neurological diseases: an update. *CNS* & *Neurological Disorders - Drug Targets*, 16(3):257-265.

Cacciar, i B., Pastorin, G. & Spalluto, G, 2003. Medicinal chemistry of A2A adenosine receptor antagonists. *Current topics in medicinal chemistry*, 3(4):403 411.

Calne, D.B. & Peppard, R.F. 1987. Aging of the nigrostriatal pathway in humans. *The Canadian journal of neurogical sciences*, 14(3):424-427.

Campagna, J., Spilman, P., Jagodzinska, B., Bai, D., Hatami, A., Zhu, C., Bilousova, T., Jun, M., Elias, C.J., Pham, J., Cole, G., LaDu, M.J., Jung, M.E., Bredesen, D.E. & John, V. 2018. A small molecule ApoE4-targeted therapeutic candidate that normalizes sirtuin 1 levels and improves cognition in an Alzheimer's disease mouse model. *Scientific Reports*, 8(1):17574-17588.

Cao C, Cirrito JR, Lin X, Wang L, Verges DK, Dickson A, Mamcarz M, Zhang C, Mori T, Arendash GW, Holtzman DM & H., P. 2009. Caffeine suppresses amyloid-beta levels in plasma and brain of Alzheimer's disease transgenic mice. *Journal of Alzheimer's Disease*, 17:681-697.

Chakrabarti, S., Khemka, V.K., Banerjee, A., Chatterjee, G., Ganguly, A. & Biswas, A. 2015. Metabolic Risk Factors of Sporadic Alzheimer's Disease: Implications in the Pathology, Pathogenesis and Treatment. *Aging and disease*, 6(4):282-299.

Chang, L., Brussee, J. & Ijzerman, A.P. 2004. Non-xanthine antagonists for the adenosine A1 receptor. *Chemistry & Biodiversity*, 1:1591–1626.

Chapuis, J., Moisan, F., Mellick, G., Elbaz, A., Silburn, P., Pasquier, F., Hannequin, D., Lendon, C., Campion, D. & Amouyel, P. 2008. Association study of the NEDD9 gene with the risk of developing Alzheimer's and Parkinson's disease. *Hum Mol Genet*, 17(18):2863-2867.

Chen, C. & Okayama, H. 1987. High-efficiency transformation of mammalian cells by plasmid DNA. *Molecular and cellular biology*, 7(8):2745-2752.

Chen, J-F., Eltzschig, H.K. & Fredholm, B.B. 2013. Adenosine receptors as drug targets—what are the challenges? *Nature reviews drug discovery*, 12(4):265-286.

Chen, J.F. & Chern, Y. 2011. Impacts of methylxanthines and adenosine receptors on neurodegeneration: human and experimental studies. *Handbook of experimental pharmacology*, 200:267–310.

Chen, J., Lee, C. & Chern, Y. 2014. Adenosine receptor neurobiology: overview. *International review of neurobiology*, 119:1–50.

Chen, J.-F., Huang, Z., Ma, J., Zhu, J., Moratalla, R., Standaert, D., Moskowitz, M.A., Fink, J.S. & Schwarzschild, M.A. 1999. A2A adenosine receptor deficiency attenuates brain injury induced by transient focal ischemia in mice. *Journal of neuroscience*, 19(21):9192-9200.

Chen, X., Ghribi, O. & Geiger, J.D. 2010. Caffeine protects against disruptions of the bloodbrain barrier in animal models of Alzheimer's and Parkinson's diseases. *Journal of Alzheimer's Disease*, 20(s1):S127-S141.

Chen, J.F., Xu, K., Petzer, J.P., Staal, R., Xu, Y.H., Beilstein, M., Sonsalla, P.K., Castagnoli, K., Castagnoli, N., Jr & Schwarzschild, M.A. 2001. Neuroprotection by caffeine and A(2A) adenosine receptor inactivation in a model of Parkinson's disease. *The journal of neuroscience*, 21(10):RC143.

Cheignon, C., Tomas, M., Bonnefont-Rousselot, D., Faller, P., Hureau, C. & Collin, F. 2018. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox biology*, 14:450-464.

Chin-Chan, M., Navarro-Yepes, J. & Quintanilla-Vega, B. 2015. Environmental pollutants as risk factors for neurodegenerative disorders: Alzheimer and Parkinson diseases. *Frontiers in cellular neurosciences*, 9:124.

Chinta, S.J., Woods, G., Rane, A., Demaria, M., Campisi, J. & Andersen, J.K. 2015. Cellular senescence and the aging brain. *Experimental Gerontology*, 68:3-7.

Ciruela, F., Casado, V., Rodrigues, R.J., Luján, R., Burgueno, J., Canals, M., Borycz, J.,
Rebola, N., Goldberg, S.R., Mallol, J., Cortés, A, Canela, E.I., López-Giménez, J.F., Milligan,
G., Lluis, C., Cunha, R.A., Ferré, S. & Franco, R. 2006. Presynaptic control of striatal
glutamatergic neurotransmission by adenosine A1-A2A receptor heteromers. *Journal of neurosciences*, 26:2080–2087.

Costa, J., Lunet, N., Santos, C., Santos, J. & Vaz-Carneiro, A. 2010. Caffeine exposure and the risk of Parkinson's disease: A systematic review and meta-analysis of observational studies. *Journal of Alzheimer's disease*, 20(1):S221–S238.

Cunha, R.A. 2005. Neuroprotection by adenosine in the brain: From A(1) receptor activation to A (2A) receptor blockade. *Purinergic Signal*, 1:111–134.

Cunha, R.A. & Agostinho, P.M. 2010. Chronic caffeine consumption prevents memory disturbance in different animal models of memory decline. *Journal of Alzheimer's Disease*, 20(s1):S95-S116.

Cupino, T. & Zabel, MK,. 2013. Alzheimer's silent partner: cerebral amyloid angiopathy. *Translational stroke Research*, 5:330-337.

Cummings, J., Lee, G., Ritter, A., Sabbagh, M. & Zhong, K. 2019. Alzheimer's disease drug development pipeline: 2019. *Alzheimer's & Dementia: Translational Research & Clinical Interventions*, 5:272-293.

Grünblatt, E., Ruder, J., Monoranu, C.M., Riederer, P., Youdim, M.B. & Mandel, S.A. 2018. Differential alterations in metabolism and proteolysis-related proteins in human Parkinson's disease substantia nigra. *Neurotoxicity research*, 33(3):560-568.

Dai, S.S., Li, W., An, J. H., Wang, H., Yang, N., Chen, X. Y., Zhao, Y., Li, P., Liu, P., Chen, J.F., Zhou, Y. G. 2010. Adenosine A2A receptors in both bone marrow cells and nonbone marrow cells contribute to traumatic brain injury. *Journal of neurochemistry*, 113:1536–1544.

Dall'Igna OP, F.P., Gomes MW, Souza DO, Cunha RA,Lara DR 2007. Caffeine and adenosine A(2a) receptor antagonists prevent beta-amyloid (25-35)-induced cognitive deficits in mice. *Experimental Neurology*, 203:241-245.

Dall'Igna, O.P., Porciúncula, L.O., Souza, D.O., Cunha, R.A. & Lara, D.R. 2003. Neuroprotection by caffeine and adenosine A2A receptor blockade of β-amyloid neurotoxicity. *British journal of pharmacology*, 138(7):1207-1209.

Dallos, V., Heathfield, K., Stone, P. & Allen, F.A.D. 1970. Use of Amantadine in Parkinson's Disease. Results of a Double-blind Trial. *British medical journal*, 4(5726):24-26.

Dauer, W. & Przedborski, S. 2003. Parkinson's disease: mechanisms and models. *Neuron*, 39(6):889-909.

Deep-Brain Stimulation for Parkinson's Disease Study Group, Obeso, J.A., Olanow, C.W., Rodriguez-Oroz, M.C., Krack, P., Kumar. R. & Lang, A.E. 2001. Deep-brain stimulation of the subthalamic nucleus or the pars interna of the globus pallidus in Parkinson's disease. *The New England journal of medicine*, 345(13):956-963. De Erausquin, G., Costa, E. & Hanbauer, I. 1994. Calcium homeostasis, free radical formation, and trophic factor dependence mechanisms in Parkinson's disease. *Pharmacological Reviews*, 46(4):467-482.

De Mendonça A., Almeida, T., Bashir, Z.I. & Ribeiro, J.A. 1997. Endogenous adenosine attenuates long-term depression and depotentiation in the CA1 region of the rat hippocampus. *Neuropharmacology*, 36:161-167.

De Mendonça, A., Sebastião, A.M. & Ribeiro, J.A. 2000. Adenosine: does it have a neuroprotective role after all? *Brain Research Reviews*, 33(2-3):258-274.

De Smet, Y., Ruberg, M., Serdaru, M., Dubois, B., Lhermitte, F. & Agid, Y. 1982. Confusion, dementia and anticholinergics in Parkinson's disease. *Journal of neurology, Neurosurgery & Psychiatry*, 45(12):1161-1164.

Dennissen, F.J., Anglada-Huguet, M., Sydow, A., Mandelkow, E. & Mandelkow, E.M. 2016. Adenosine A1 receptor antagonist rolofylline alleviates axonopathy caused by human Tau ΔK280. *Proceedings of the National Academy of Sciences*, 113(41):11597-11602.

Di Stefano, A., Iannitelli, A., Laserra, S. & Sozio, P. 2011. Drug delivery strategies for Alzheimer's disease treatment. *Expert opinion on drug delivery*, 8(5):581-603.

Dickson, D.W. 2018. Neuropathology of Parkinson disease. *Parkinsonism & related disorders*, 46:S30-S33.

Doody, R.S., Gavrilova, S.I., Sano, M., Thomas, R.G., Aisen, P.S., Bachurin, S.O., Seely, L. & Hung, D. 2008. Effect of dimebon on cognition, activities of daily living, behaviour, and global function in patients with mild-to-moderate Alzheimer's disease: a randomised, double-blind, placebo-controlled study. *Lancet*, 372(9634):207-215.

Drury, A.N. & Szent-Györgyi, A. 1929. The physiological activity of adenine compounds with especial reference to their action upon the mammalian heart. *Journal of physiology*, 68(3):213-237.

Duyckaerts, C. & Dickson, D. 2011 In Neurodegeneration: the molecular pathology of dementia and movement disorders, 2<sup>ND</sup> Edition (ed Dennis Dickson, Roy O. Weller):462-496

Ehrt, U., Broich, K., Larsen, J.P., Ballard, C. & Aarsland, D. 2010. Use of drugs with anticholinergic effect and impact on cognition in Parkinson's disease: a cohort study. *Journal of neurology, neurosurgery & psychiatry*, 81(2):160-165.

El Yacoubi, M., Bouali, S., Popa, D., Naudon, L., Leroux-Nicollet, I., Hamon, M., Costentin, J., Adrien, J. & Vaugeois, J.-M. 2003. Behavioral, neurochemical, and electrophysiological characterization of a genetic mouse model of depression. *Proceedings of the National Academy of Sciences*, 100(10):6227-6232.

Eskelinen, M.H., Ngandu, T., Tuomilehto, J., Soininen, H. & Kivipelto, M. 2009. Midlife coffee and tea drinking and the risk of late-life dementia: a population-based CAIDE study. *Journal of Alzheimer's Disease*, 16(1):85-91.

Fabbrini, G., Brotchie, J.M., Grandas, F., Nomoto, M. & Goetz, C.G. 2007. Levodopa-induced dyskinesias. *Journal of movement disorders*, 22(10):1379-1389

Ferrer, I., Rovira, M.B., Guerra, M.L.S., Rey, M.J. & Costa-Jussá, F. 2004. Neuropathology and pathogenesis of encephalitis following amyloid  $\beta$  immunization in Alzheimer's disease. *Brain Pathology*, 14(1):11-20.

Fink, J.S., Weaver, D.R., Rivkees, S.A., Peterfreund, R.A., Pollack, A.E., Adler, E.M. & Reppert, S.M. 1992,. Molecular cloning of the rat A2 adenosine receptor: selective co-expression with D2 dopamine receptors in rat striatum. *Brain research. Molecular brain research*, 14:186-195.

Fiorito, V., Chiabrando, D. & Tolosano, E. 2018. Mitochondrial targeting in neurodegeneration: a heme perspective. *Pharmaceuticals*, 11(3):87.

Frampton, J.E. 2014. Pramipexole extended-release: a review of its use in patients with Parkinson's disease. *Drugs*, 74(18):2175-2190.

Franceschi, C., Bonafè, M., Valensin, S., Olivieri, F., De Luca, M., Ottaviani, E. & De Benedictis,G. 2000. Inflamm-aging: an evolutionary perspective on immunosenescence. *Annals of the New York Academy of Sciences*, 908(1):244-254.

Fredholm, B. & Dunwiddie, T. 1988. How does adenosine inhibit transmitter release? *Trends in pharmacological sciences*, 9(4):130-134.

Fredholm, B.B., Bättig, K., Holmén, J., Nehlig, A. & Zvartau, E.E. 1999. Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacological reviews*, 51(1):83-133.

Fredholm, B.B., IJzerman, A.P., Jacobson, K.A., Klotz, K.-N. & Linden, J. 2001. International Union of Pharmacology. XXV. Nomenclature and classification of adenosine receptors. *Pharmacological reviews*, 53(4):527-552.

Freund, S., Ungerer, M. & Lohse, M.J. 1994. A 1 adenosine receptors expressed in CHO-cells couple to adenylyl cyclase and to phospholipase C. *Naunyn-Schmiedeberg's archives of pharmacology*, 350(1):49-56.

Gaonkar, S.L. & Vignesh, U. 2017. Synthesis and pharmacological properties of chalcones: a review. *Research on chemical intermediates*, 43(11):6043-6077.

García-Vázquez, R., Rebitski, E.P., Viejo, L., de los Ríos, C., Darder, M. & García-Frutos, E.M. 2020. Clay-based hybrids for controlled release of 7-azaindole derivatives as neuroprotective drugs in the treatment of Alzheimer's disease. *Applied Clay Science*, 189:105541.

Gelber, R.P., Petrovitch, H., Masaki, K.H., Ross, G.W. & White, L.R. 2011. Coffee intake in midlife and risk of dementia and its neuropathologic correlates. *Journal of Alzheimer's Disease*, 23(4):607-615.

Gimenez-Llort, L., Fernandez-Teruel, A., Escorihuela, R.M., Fredholm, B.B., Tobena, A., Pekny, M. & Johansson, B. 2002. Mice lacking the adenosine A1 receptor are anxious and aggressive, but are normal learners with reduced muscle strength and survival rate. *The European journal of neuroscience*, 16:547-550.

Gimenez-Llort, L., Schiffmann, S.N., T. Shmidt, L. Canela, L. Camon, M. Wassholm, M. Canals,
A. Terasmaa, A. Fernandez-Teruel, A. Tobena, E. Popova, S. Ferre, L.Agnati, F. Ciruela, E.
Martinez, J. Scheel-Kruger, C. Lluis, R. Franco, K. Fuxe & M.Bader. 2007. Working memory
deficits in transgenic rats overexpressing humanadenosine A2A receptors in the brain. *Neurobiology of learning and memory*, 87:42–56.

Gomes, C.V., Kaster, M.P., Tomé, A.R., Agostinho, P.M. & Cunha, R.A. 2011. Adenosine receptors and brain diseases: Neuroprotection and neurodegeneration. *Biochimica et Biophysica Acta*, 1808((5):1380–1399.

Gottwald, M.D. & Aminoff, M.J. 2008. New frontiers in the pharmacological management of Parkinson's disease. *Drugs of today*, 44(7):531-545.

Graham, D.G., Tiffany, S.M., Bell, W.R. & Gutknecht, W.F. 1978. Autoxidation versus covalent binding of quinones as the mechanism of toxicity of dopamine, 6-hydroxydopamine, and related compounds toward C1300 neuroblastoma cells in vitro. *Molecular pharmacology*, 14(4):644-653.

Grondin, R., Bedard, P. J., Hadj Tahar, A., Gregoire, L., Mori, A., Kase, H. 1999. Antiparkinsonian effect of a new selective adenosine A2A receptor antagonist in MPTP-treated monkeys. *Neurology*, 52:1673–1677.

Grünblatt, E., Ruder, J., Monoranu, C.M., Riederer, P., Youdim, M.B. & Mandel, S.A. 2018. Differential alterations in metabolism and proteolysis-related proteins in human Parkinson's disease substantia nigra. *Neurotoxicity research*, 33(3):560-568.

Gulati, A., Hornick, M.G., Briyal, S. & Lavhale, M.S. 2018. A novel neuroregenerative approach using ET(B) receptor agonist, IRL-1620, to treat CNS disorders. *Physiological research*, 67(Supplementum 1):S95-S113.

Haas, C. 2012. Strategies, development, and pitfalls of therapeutic options for Alzheimer's disease. *Journal of Alzheimer's Disease*, 28(2):241-281.

Halldner, L., Lozza, G., Lindström, K., & Fredholm, B. B. 2000. Lack of tolerance to motor stimulant effects of a selective adenosine A(2A) receptor antagonist. European journal of pharmacology, 406:345–354.

Kurz, A., Basun, H., Burger, K., Mortberg, A., Frolich, L., Schroder, J., Schonknecht, P. & Moller, H.J. 2009. Lithium trial in Alzheimer's disease : a randomized, single-blind, placebocontrolled, multicenter 10-week study. *The journal of clinical psychiatry*, 70(6): 922-931.

Haselkorn, M.L., Shellington, D.K., Jackson, E.K., Vagni, V.A., Janesko-Feldman, K., Dubey, R.K., Gillespie, D.G., Cheng, D., Bell, M.J., Jenkins, L.W., Homanics, G.E., Schnermann, J. & Kochanek, P.M. 2010. Adenosine A1 receptor activation as a brake on the microglial response after experimental traumatic brain injury in mice. *Journal of neurotrauma*, 27:901–910.

Hauw, J., Seilhean, D., Piette, F., Uchihara, T. & Duyckaerts, C. 1996. Alzheimer's disease lesions: from morphology to cell biology. *J Bulletin de l'Academie nationale de medecine*, 180(7):1687-1700.

Herrmann, N., Chau, S.A., Kircanski, I. & Lanctot, K.L. 2011. Current and emerging drug treatment options for Alzheimer's disease. *Drugs*, 71(15):2031-2065.

Hertel, F., Züchner, M., Weimar, I., Gemmar, P., Noll, B., Bettag, M. & Decker, C. 2006. Implantation of electrodes fordeep brain stimulation of the subthalamic nucleus in advanced Parkinson's disease with the aid of intraoperative microrecording under general anesthesia. *Neurosurgery*, 59(5):E1138-E1138. Hirsch, E.C., Vyas, S. & Hunot, S. 2012. Neuroinflammation in Parkinson's disease. *Parkinsonism & related disorders*, 18:S210-S212.

Holmes, C., Wilkinson, D., Dean, C., Vethanayagam, S., Olivieri, S., Langley, A., Pandita-Gunawardena, N.D., Hogg, F., Clare, C. & Damms, J. 2004. The efficacy of donepezil in the treatment of neuropsychiatric symptoms in Alzheimer disease. *Neurology*, 63(2):214-219.

Horgusluoglu, E., Nudelman, K., Nho, K. & Saykin, A.J. 2017. Adult neurogenesis and neurodegenerative diseases: a systems biology perspective. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*, 174(1):93-112.

Hroudov, J., Singh, N. & Fišar, Z. 2014. Mitochondrial Dysfunctions in Neurodegenerative Diseases: Relevance to Alzheimer's Disease. *BioMed Research International*, 2014:1-9.

Ittner, L.M. & Götz, J. 2011. Amyloid-β and tau—a toxic pas de deux in Alzheimer's disease. *J Nature Reviews Neuroscience*, 12(2):65-72.

Hockemeyer, J., Burbiel, J.C. & Müller, C.E. 2004. Multigram-scale syntheses, stability, and photoreactions of A2A adenosine receptor antagonists with 8-styrylxanthine structure: potential drugs for Parkinson's disease. *The Journal of organic chemistry*, 69(10):3308-3318.

Huang QJ, J.H., Hao XL, Minor TR.. 2004. Brain IL-1 beta was involved in reserpineinduced behavioral depression in rats. *Acta Pharmacologica sinica*, 25(3):293-296.

Ikeda, K., Kurokawa, M., Aoyama, S., Kuwana, Y. 2002. Neuroprotection by adenosine A2A receptor blockade in experimental models of Parkinson's disease. *Journal of neurochemistry*, 80: 262–267.

Jacobson, K.A., Gallo-Rodriguez, C., Melman, N., Fischer, B., Maillard, M., van Bergen, A., van Galen, P.J. & Karton, Y. 1993. Structure-activity relationships of 8-styrylxanthines as A2-selective adenosine antagonists. *Journal of medicinal chemistry*, 36(10):1333–1342.

Jacobson, K.A. & Gao, Z.-G. 2006. Adenosine receptors as therapeutic targets. *Nature reviews. Drug discovery*, 5(3):247.

Jankovic, J. 2008. Parkinson's disease: clinical features and diagnosis. *Journal of Neurology, Neurosurgery & Psychiatry*, 79(4):368-376.

Jarvis, M.J. 1993. Does caffeine intake enhance absolute levels of cognitive performance? *Psychopharmacology*, 110(1-2):45-52.

Jarvik, L. & Greenson, H. 1987. About a peculiar disease of the cerebral cortex. *Alzheimer's disease and associated disorders*, 1(1):3-8.

Jenner, P., Dexter, D., Sian, J., Schapira, A. & Marsden, C. 1992. Oxidative stress as a cause of nigral cell death in Parkinson's disease and incidental Lewy body disease. *Annals of neurology*, 32(S1):S82-S87.

Johansson, B., Halldner, L., Dunwiddie, T.V., Masino, S.A., Poelchen, W., Giménez-Llort, L., Escorihuela, R.M., Fernández-Teruel, A., Wiesenfeld-Hallin, Z., Xu, X.J., Hrdemark, A., Betsholtz, C., Herlenius, E, Fredholm, B.B. 2001. Hyperalgesia, anxiety, and decreased hypoxic neuroprotection in mice lacking the adenosine A1 receptor,. *Proceedings of the National Academy of sciences of the United States of America*, 98:9407-9412.

Jones, P.A., Smith, R. A., Stone, T. W. 1998. Protection against hippocampal kainate excitotoxicity by intracerebral administration of an adenosine A2A receptor antagonist. *Brain research*, 800(2):328–335.

Kaakkola, S. 2000. Clinical pharmacology, therapeutic use and potential of COMT inhibitors in Parkinson's disease. *Drugs*, 59(6):1233-1250.

Kanda, T., Jackson, M. J., Smith, L. A., Pearce, R. K., Nakamura, J., Kase, H., *et al.* 2000. Combined use of the adenosine A(2A) antagonist KW-6002 with L-DOPA or with selective D1 or D2 dopamine agonists increases antiparkinsonian activity but not dyskinesia in MPTP-treated monkeys. *Experimental neurology*, 162: 321–327.

Kaster, M.P., Rosa, A.O., Rosso, M.M., Goulart, E.C., Santos, A.R. & Rodrigues, A.L.S. 2004. Adenosine administration produces an antidepressant-like effect in mice: evidence for the involvement of A1 and A2A receptors. *Neuroscience letters*, 355(1-2):21-24.

Koutsilieri, E., Lutz, M. & Scheller, C. 2013. Autoimmunity, dendritic cells and relevance for Parkinson's disease. *Journal of Neural Transmission*, 120(1):75-81.

Kim, W.S., Kågedal, K. & Halliday, G.M. 2014. Alpha-synuclein biology in Lewy body diseases. *Alzheimer's research & therapy*, 6(5-8):73.

Klotz, K.-N., Vogt, H. & Tawfik-Schlieper, H. 1991. Comparison of A 1 adenosine receptors in brain from different species by radioligand binding and photoaffinity labelling. *Naunyn-Schmiedeberg's archives of pharmacology*, 343(2):196-201.

Klotz K-N, Hessling, J., Hegler J, Owman C, Kull B, Fredholm BB, and Lohse MJ. 1998. Comparative pharmacology of human adenosine receptor subtypes - characterization of stably transfected receptors in CHO cells. *Naunyn Schmiedebergs Archives of pharmacology*, 357:1– 9.

Klotz, K.-N. 2000. Adenosine receptors and their ligands. *Naunyn-Schmiedeberg's archives of pharmacology*, 362(4-5):382-391.

Klotz, K., Hessling, J., Hegler, J., Owman, C., Kull, B., Fredholm, B. & Lohse, M. 1997. Comparative pharmacology of human adenosine receptor subtypes–characterization of stably transfected receptors in CHO cells. *J Naunyn-Schmiedeberg's archives of pharmacology*, 357, 1-9.

Kopf, S.R., Melani, A., Pedata, F. & Pepeu, G. 1999. Adenosine and memory storage. *Psychopharmacology*, 146: 214-219.

Kumar, P.M., Paing, S.S.T., Li, H., Pavanni, R., Yuen, Y., Zhao, Y. & Tan, E.K. 2015. Differential effect of caffeine intake in subjects with genetic susceptibility to Parkinson's Disease. *Scientific reports*, 5(1):1-3.

Kumar, P. & Clark, M. 2005. Degenerative neuronal diseases. *Kumar and Clark Clinical Medicine. Saunders Press: London*:1254-1255.

Kvernmo, T., Härtter, S. & Burger, E. 2006. A review of the receptor-binding and pharmacokinetic properties of dopamine agonists. *Clinical therapeutics*, 28(8):1065-1078.

Langston, J.W., Ballard, P., Tetrud, J.W. & Irwin, I. 1983. Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis. *Science*, 219(4587):979-980.

LeWitt, P.A. 2000. New drugs for the treatment of Parkinson's disease. *Pharmacotherapy*, 20(1 Pt 2):26S-32S.

Lees, A.J. 2002. Drugs for Parkinson's disease. *Journal of Neurology, Neurosurgery & Psychiatry*, 73(6):607-610.

Legoabe, L.J., Van der Walt, M.M. & Terre'Blanche, G. 2018. Evaluation of 2-benzylidene-1tetralone derivatives as antagonists of A1 and A2A adenosine receptors. *Chemical biology & drug design*, 91(1):234-244. Liang, Z., Zhao, Y., Ruan, L., Zhu, L., Jin, K., Zhuge, Q., Su, D.-M. & Zhao, Y. 2017. Impact of aging immune system on neurodegeneration and potential immunotherapies. *Progress in neurobiology*, 157:2-28.

Lipton, S.A. 2006. NMDA Receptors, Glial Cells, and Clinical Medicine. Neuron, 50(1):9-11.

Lindsay, J., Laurin, D., Verreault, R., Hébert, R., Helliwell, B., Hill, G.B. & McDowell, I. 2002. Risk factors for Alzheimer's disease: a prospective analysis from the Canadian Study of Health and Aging. *American journal of epidemiology*, 156(5):445-453.

Lobato, K.R., Binfaré, R.W., Budni, J., Rosa, A.O., Santos, A.R.S. & Rodrigues, A.L.S. 2008. Involvement of the adenosine A1 and A2A receptors in the antidepressant-like effect of zinc in the forced swimming test. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 32(4):994-999.

Lohse, M.J., Klotz, K.-N., Lindenborn-Fotinos, J., Reddington, M., Schwabe, U. & Olsson, R.A. 1987. 8-Cyclopentyl-1, 3-dipropylxanthine (DPCPX)—a selective high affinity antagonist radioligand for A 1 adenosine receptors. *Naunyn-Schmiedeberg's archives of pharmacology*, 336(2):204-210.

Londos, C., Cooper, D. & Wolff, J. 1980. Subclasses of external adenosine receptors. *Proceedings of the National Academy of Sciences*, 77(5):2551-2554.

Lopes, L.V., Cunha, R.A. & Ribeiro, J.A. 1999. Cross-talk between A(1)/A(2A) adenosine receptors in the rat hippocampus and cortex. *British journal of pharmacology*, 127:U20-U20.

Lopes, V.L., Sebastiao, M.A. & Ribeiro, A.J. 2011. Adenosine and Related Drugs in Brain Diseases: Present and Future in Clinical Trials. *Current Topics in medicinal chemistry*, 11(8):1087-1101.

Lu, G., Zhou, Q.-X., Kang, S., Li, Q.-L., Zhao, L.-C., Chen, J.-D., Sun, J.-F., Cao, J., Wang, Y.-J. & Chen, J. 2010. Chronic morphine treatment impaired hippocampal long-term potentiation and spatial memory via accumulation of extracellular adenosine acting on adenosine A1 receptors. *Journal of neuroscience*, 30(14):5058-5070.

Lucas, M., Mirzaei, F., Pan, A., Okereke, O.I., Willett, W.C., O'Reilly, E.J., Koenen, K., Ascherio, A. . 2011. Coffee, caffeine and risk of depression among women. *Archives of Internal Medicine*, 171:1571-1578.

Maia, L. & De Mendonça, A. 2002. Does caffeine intake protect from Alzheimer's disease? . *European journal of neurology*, 9: 377-382.

Maemoto, T., Tada, M., Mihara, T., Ueyama, N., Matsuoka, H., Harada, K., Yamaji, T., Shirakawa, K., Kuroda, S., Akahane, A., Iwashita, A., Matsuoka, N. & Mutoh, S. 2004. Pharmacological Characterization of FR194921, a New Potent, Selective, and Orally Active Antagonist for Central Adenosine Receptors. *Journal of pharmacological sciences*, *96(1)*:42-52.

Maemoto, T., Finlayson, K., Olverman, H.J., Akahane, A., Horton, R.W. & Butcher, S.P. 1997. Species differences in brain adenosine A1 receptor pharmacology revealed by use of xanthine and pyrazolopyridine based antagonists. *British journal of pharmacology*, 122(6):1202-1208.

Mangialasche, F., Solomon, A., Winblad, B., Mecocci, P. & Kivipelto, M. 2010. Alzheimer's disease: clinical trials and drug development. *Lancet Neurology*, 9(7):702-716.

Mannisto, P. 1999. Catechol-O-methyltransferase (COMT) : biochemistry, molecular biology, pharmacology, and clinical efficacy of the new selective COMT inhibitors. *Pharmacological reviews*, 51(4):593-628.

Martyn, C. & Gale, C. 2003. Tobacco, coffee, and Parkinson's disease: Caffeine and nicotine may improve the health of dopaminergic systems. *British Medical Journal*, 326(7389):561-562.

Massoud, F., Desmarais, J.E. & Gauthier, S. 2011. Switching cholinesterase inhibitors in older adults with dementia. *International psychogeriatry*, 23(3):372-378.

McKhann, G.M., Knopman, D.S., Chertkow, H., Hyman, B.T., Jack Jr, C.R., Kawas, C.H., Klunk, W.E., Koroshetz, W.J., Manly, J.J. & Mayeux, R. 2011. The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's dementia*, 7(3):263-269.

Mihara, T., Mihara, K., Yarimizu, J., Mitani, Y., Matsuda, R., Yamamoto, H., Aoki, S., Akahane, A., Iwashita, A. & Matsuoka, N. 2007. Pharmacological characterization of a novel, potent adenosine A1 and A2A receptor dual antagonist, 5-[5-amino-3-(4-fluorophenyl)pyrazin-2-yl]-1-isopropylpyridine-2(1H)-one (ASP5854), in models of Parkinson's disease and cognition. *Journal of pharmacoloty and experimental therapeutics*, 323((2):708–719.

Mishina, M., Ishiwata, K., Naganawa, M., Kimura, Y., Kitamura, S., Suzuki, M., Hashimoto, M., Ishibashi, K., Oda, K., Sakata, M., Hamamoto, M., Kobayashi, S., Katayama, Y. & Ishii, K.

2011. Adenosine A2A Receptors Measured with [11C]TMSX PET in the Striata of Parkinson's Disease Patients. *PLoS ONE*, 6(2):1-8.

Montastruc, J.L., Rascol, O. & Senard, J.M. 1999. Treatment of Parkinson's disease should begin with a dopamine agonist. *Movement disorders*, 14(5):725-730.

Mori, A. 2014 Mode of action of adenosine A2A receptor antagonists as symptomatic treatment for Parkinson's disease.. London.: Academic Press. 119:88-116.

Moro, S., Gao, Z.G., Jacobson, K.A. & Spalluto, G. 2006. Progress in the pursuit of therapeutic adenosine receptor antagonists. *Medicinal Research Reviews*, 26(2):131-159.

Muller, C.E. 2002. P2-pyrimidinergic receptors and their ligands. *Current pharmaceutical design*, 8(26):2353-2369.

Müller, C.E. & Jacobson, K.A. 2011. Recent developments in adenosine receptor ligands and their potential as novel drugs. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1808(5):1290-1308.

Münchau, A. & Bhatia, K.P. 2000. Pharmacological treatment of Parkinson's disease. *Postgraduate medical journal,* 76(900):602-10.

Nagatsu, T. & Sawada, M. 2009. L-dopa therapy for Parkinson's disease: Past, present, and future. *Parkinsonism & Related disorders*, 15:S3-S8.

Nussbaum, R.L. & Ellis, C.E. 2003. Alzheimer's disease and Parkinson's disease. *J New England journal of medicine*, 348(14):1356-1364.

Obeso, J.A., Olanow, C.W. & Nutt, J.G. 2000. Levodopa motor complications in Parkinson's disease. *Trends Neurosci*, 23(10):S2-7.

Pahwa, R., Factor, S.A., Lyons, K.E., Ondo, W.G., Gronseth, G., Bronte-Stewart, H., Hallett, M., Miyasaki, J., Stevens, J. & Weiner, W.J. 2006. Practice Parameter: Treatment of Parkinson disease with motor fluctuations and dyskinesia (an evidence-based review): [RETIRED]. *Neurology*, 66(7):983-95.

Pagano, G., Rengo, G., Pasqualetti, G., Femminella, G.D., Monzani, F., Ferrara, N. & Tagliati,
M. 2015. Cholinesterase inhibitors for Parkinson's disease: a systematic review and metaanalysis. *Journal of Neurology. Neurosurgery and Psychiatry*, 86(7):767-773. Palacios, N., Gao, X., McCullough, M.L., Schwarzschild, M.A., Shah, R. Gapstur, S., Ascherio,
A. 2012; Caffeine and risk of Parkinson's disease in a large cohort of men and women. *Movement disorders*, 27(10):1276–1282.

Palmer, T. & Stiles, G. 1995. Adenosine receptors. Neuropharmacology, 34(7):683-694.

Ongini, E., Dionisotti, S., Gessi, S., Irenius, E. & Fredholm, B.B. 1999. Comparison of CGS 15943, ZM 241385 and SCH 58261 as antagonists at human adenosine receptors. *Naunyn-Schmiedeberg's archives of pharmacology*, 359:7-10.

Panza, F., Solfrizzi, V., Barulli, M., Bonfiglio, C., Guerra, V., Osella, A., Seripa, D., Sabbà, C., Pilotto, A. & Logroscino, G. 2015. Coffee, tea, and caffeine consumption and prevention of late-life cognitive decline and dementia: a systematic review. *The journal of nutrition, health & aging*, 19(3):313-328.

Parkinson, J. 2002. An essay on the shaking palsy. *The journal of Neuropsychiatry and clinical neurosciences*, 14(2):223-236.

Perl, D.P. 2010. Neuropathology of Alzheimer's disease. *Mount Sinai journal of medicine*, 77(1):32-42.

Pechlivanova MD, T.J., Alova LH, Petkov VV, Nikolov RP, Yakimova KS. 2012. Effect of longterm caffeine administration on depressive-like behavior in rats exposed to chronic unpredictable stress. *Behavioral Pharmacology*, 23:339-347.

Perry, E.K., Tomlinson, B.E., Blessed, G., Bergmann, K., Gibson, P.H. & Perry, R.H. 1978. Correlation of cholinergic abnormalities with senile plaques and mental test scores in senile dementia. *British medical journal*, 2(6150):1457-1459.

Pinna, A., Tronci, E., Schintu, N., Simola, N., Volpini, R., Pontis, S., Cristalli, G. & Morelli, M. 2010. A new ethyladenine antagonist of adenosine A2A receptors: Behavioral and biochemical characterization as an antiparkinsonian drug. *Neuropharmacology*, 58(3):613-623.

Pham, N., Nanri A, Kurotani K, Kuwahara K, Kume A, Sato M, Hayabuchi H & T., M. 2013. Green tea and coffee consumption is inversely associated with depressive symptoms in a Japanese working population *Public Health Nutrition*, 17(3):625-633.

Polinsky, R.J. 1998. Clinical pharmacology of rivastigmine: a new-generation acetylcholinesterase inhibitor for the treatment of Alzheimer's disease. *Clinical Therapy*, 20(4):634-647.

Popoli, P., Regio, R. & Pezzola, A. 2000. Effects of SCH 58261, an adenosine A (2A) receptor antagonist, on quinpirole-induced turning in 6-hydroxydopamine-lesioned rats. Lack of tolerance after chronic caffeine intake. Neuropsychopharmacology 22:522-529.

Pracharova, J., Saltarella, T., Radosova Muchova, T., Scintilla, S., Novohradsky, V., Novakova, O., Intini, F.P., Pacifico, C., Natile, G., Ilik, P., Brabec, V. & Kasparkova, J. 2015. Novel antitumor cisplatin and transplatin derivatives containing 1-methyl-7-azaindole: synthesis, characterization, and cellular responses. *Journal of Medicinal Chemistry*, 58(2):847-859.

Porsolt, R.D., Bertin, A. & Jalfre, M. 1977. Behavioral despair in mice: a primary screening test for antidepressants. *Archives Internationales de Pharmacodynamie et de Therapie*, 229 327–336.

Prediger, R., Da Cunha, C. & Takahashi, R. 2005. Antagonistic interaction between adenosine A2A and dopamine D2 receptors modulates the social recognition memory in reserpine-treated rats. *Behavioural pharmacology*, 16(4):209-218.

Preti, D., Baraldi, P.G., Moorman, A.R., Borea, P.A. & Varani, K. 2015. History and perspectives of A2A adenosine receptor antagonists as potential therapeutic agents. *Medicinal research reviews*, 35(4):790-848.

Prince, M.J., Wu, F., Guo, Y., Robledo, L.M.G., O'Donnell, M., Sullivan, R. & Yusuf, S. 2015. The burden of disease in older people and implications for health policy and practice. *Lancet*, 385(9967):549-562.

Qhobosheane, M.A., Legoabe, L.J., Josselin, B., Bach, S., Ruchaud, S. & Beteck, R.M. 2020a. Synthesis and evaluation of C3 substituted chalcone-based derivatives of 7-azaindole as protein kinase inhibitors. *Chemical biology & drug design, 96(6):* 1395-1407.

Qhobosheane, M.A., Legoabe, L.J., Josselin, B., Bach, S., Ruchaud, S., Petzer, J.P. & Beteck, R.M. 2020b. Synthesis and evaluation of 7-azaindole derivatives bearing benzocycloalkanone motifs as protein kinase inhibitors. *Bioorganic & Medicinal Chemistry*, 28(11):115468.

Ralevic, V. & Burnstock, G. 1998. Receptors for purines and pyrimidines. *Pharmacological reviews*, 50(3):413-492.

Ramlackhansingh, A.F., Bose, S.K., Ahmed, I., Turkheimer, F.E., Pavese, N. & Brooks, D.J.2011. Adenosine 2A receptor availability in dyskinetic and nondyskinetic patients withParkinson disease. *Neurology*, 76(21):1811-1816.

Rebola, N., Simões, A.P., Canas, P.M., Tomé, A.R., Andrade, G.M., Barry, C.E., Agostinho, P.M., Lynch, M.A. & Cunha, R.A. 2011. Adenosine A2A receptors control neuroinflammation and consequent hippocampal neuronal dysfunction. *Journal of neurochemistry*, 117(1):100-111.

Ribeiro, J.A. & Sebastiao, A.M. 2010. Caffeine and adenosine. *Journal of Alzheimer's disease*, 20:S3–S15.

Ribeiro, J., Sebastiao, A. & De Mendonça, A. 2002. Adenosine receptors in the nervous system: pathophysiological implications. *Progress in neurobiology*, 68(6):377-392.

Riederer, P. & Laux, G. 2011. MAO-inhibitors in Parkinson's Disease. *Experimental Neurobiology*, 20(1):1-17.

Rivera-Oliver, M. & Díaz-Ríos, M. 2014 Using caffeine and other adenosine receptor antagonists and agonists as therapeutic tools against neurodegenerative diseases: A review. *Life Sciences*, 101 1–9.

Robeva, A.S., Woodard, R.L., Jin, X., Gao, Z., Bhattacharya, S., Taylor, H.E., Rosin, D.L. & Linden, J. 1996. Molecular characterization of recombinant human adenosine receptors. *Drug development research*, 39(3-4):243-252.

Robinson, D.M. & Plosker, G.L. 2006. Galantamine extended release. *CNS drugs*, 20(8):673-681.

Robinson, S.J., Petzer, J.P., Terre'Blanche, G., Petzer, A., Van der Walt, M.M., Bergh, J.J. & Lourens, A.C. 2015. 2-Aminopyrimidines as dual adenosine A1/A2A antagonists. *European journal of medicinal chemistry*, 104:177-188.

Rocca, W.A. 2018. The burden of Parkinson's disease: a worldwide perspective. *The Lancet Neurology*, 17(11):928-929.

Rockenstein, E., Mante, M., Adame, A., Crews, L., Moessler, H. & Masliah, E. 2007. Effects of Cerebrolysin on neurogenesis in an APP transgenic model of Alzheimer's disease. *Acta neuropathoogyl*, 113(3):265-275.

Rodrigues, F.B., Caldeira, D., Ferreira, J.J. & Costa, J. 2015. Caffeine and neuroprotection in Parkinson's disease. The Adenosinergic System. In: Morelli, M., Simola, M. & Wardas, J eds. *The Adenosinergic System - A Non-Dopaminergic Target in Parkinson's Disease.* Springer, *pp.* 233-272.

Ross, G.W. 2000. Association of coffee and caffeine intake with the risk of Parkinson disease. *Journal of the American medical association*, 283:2674-2679.

Ruusunen, A., Lehto, S.M., Tolmunen, T., Mursu, J., Kaplan, G.A. & Voutilainen, S. 2010. Coffee, tea and caffeine intake and the risk of severe depression in middle-aged Finnish men: the Kuopio Ischaemic Heart Disease Risk Factor Study. *Public health nutrition*, 13(8):1215-1220.

Sallaberry, C., Nunes, F., Costa, M.S., Fioreze, G.T., Ardais, A.P., Botton, P.H.S., Klaudat, B., Forte, T., Souza, D.O. & Elisabetsky, E. 2013. Chronic caffeine prevents changes in inhibitory avoidance memory and hippocampal BDNF immunocontent in middle-aged rats. *Neuropharmacology*, 64:153-159.

Salvemini, D., Kim, S.F. & Mollace, V. 2013;. Reciprocal regulation of the nitric oxide and cyclooxygenase pathway in pathophysiology: relevance and clinical implications. *American Journal of Physiology - Regulatory, Integrative and Comparative Physiology*, 304((7):R473–487.

Sarasija, S. & Norman, K. 2018. Role of Presenilin in Mitochondrial Oxidative Stress and Neurodegeneration in Caenorhabditis elegans. *Antioxidants*, 7(9):111. doi: 10.3390/antiox7090111

Scheltens, P., Blennow, K., Breteler, M.M.B., de Strooper, B., Frisoni, G.B., Salloway, S. & Van der Flier, W.M. 2016. Alzheimer's disease. *Lancet*, 388(10043):505-517.

Schiffmann, S., Jacobs O, & Vanderhaeghen. JJ. 1991. Striatal restricted adenosine A2 receptor (RDC8) is expressed by enkephalin but not by substance P neurons: An in situ hybridization histochemistry study. *Journal of Neurochemistry*, 57:1062–1067.

Schwarzschild, M.A., Chen, J.-F. & Ascherio, A. 2002. Caffeinated clues and the promise of adenosine A2A antagonists in PD. *Journal of Neurology*, 58(8):1154-1160.

Shook, B.C., Rassnick, S., Chakravarty, D., Wallace, N., Ault, M., Crooke, J., Barbay, J.K., Wang, A., Leonard, K. & Powell, M.T. 2010. Optimization of arylindenopyrimidines as potent adenosine A2A/A1 antagonists. *Bioorganic & medicinal chemistry letters*, 20(9):2868-2871.

Simola, N., Morelli, M., Pinna, A. 2008. Adenosine A2A receptor antagonists and Parkinson's disease: State of the art and future directions. *Current pharmaceutical design*, 14:1475–89002.

Simola, N., Pinna, A., Frau, L. & Morelli, M. 2014. Protective Agents in Parkinson's Disease: Caffeine and Adenosine A2A Receptor Antagonists. *Handbook of Neurotoxicity*:2281-2298.

Singh, N., Shreshtha, A.K., Thakur, M.S. & Patra, S. 2018. Xanthine scaffold: scope and potential in drug development. Heliyon, 4(10):e00829.

Šimić, G., Leko, M.B., Wray, S., Harrington, C.R., Delalle, I., Jovanov-Milošević, N., Bažadona, D., Buée, L., de Silva, R. & Di Giovanni, G. 2017. Monoaminergic neuropathology in Alzheimer's disease. *Progress in neurobiology*, 151:101-138.

Sowell, R.A., Owen, J.B. & Butterfield, D.A. 2009. Proteomics in animal models of Alzheimer's and Parkinson's diseases. *Ageing research reviews*, 8(1):1-17.

Štarha, P., Hošek, J., Vančo, J., Dvořák, Z., Suchý Jr, P., Popa, I., Pražanová, G. & Trávníček, Z. 2014. Pharmacological and molecular effects of platinum (II) complexes involving 7azaindole derivatives. *PLoS One*, 9(3):e90341.

Szekely, C.A., Thorne, J.E., Zandi, P.P., Ek, M., Messias, E., Breitner, J.C.S. & Goodman, S.N. 2004. Nonsteroidal Anti-Inflammatory Drugs for the Prevention of Alzheimer's Disease: A Systematic Review. *Neuroepidemiology*, 23(4):159-169.

Tabakman, R., Lecht, S. & Lazarovici, P. 2004. Neuroprotection by monoamine oxidase B inhibitors: a therapeutic strategy for Parkinson's disease? *Bioessays*, 26(1):80-90.

Taliani, S., Da Settimo, F., Martini, C., Laneri, S., Novellino, E. & Greco, G. 2020. Exploiting the Indole Scaffold to Design Compounds Binding to Different Pharmacological Targets. *Molecules*, 25(10):2331.

Tan, S.H., Karri, V., Tay, N.W.R., Chang, K.H., Ah, H.Y., Ng, P.Q., Ho, H,S., Keh, H.W. & Candasamy, M. 2019. Emerging pathways to neurodegeneration: Dissecting the critical molecular mechanisms in Alzheimer's disease, Parkinson's disease. *Biomedicine Pharmacotherapy*, 111:765-777.

Tariot, P.N. & Aisen, P.S. 2009. Can lithium or valproate untie tangles in Alzheimer's disease? *The journal of clinical psychiatry*, 70(6):919-921.

Tholfsen, L.K., Larsen, J.P., Schulz, J., Tysnes, O.-B. & Gjerstad, M.D. 2015. Development of excessive daytime sleepiness in early Parkinson disease. *Neurology*, 85(2):162-168.

Trifilieff, P., Rives, M.-L., Urizar, E., Piskorowski, R.A., Vishwasrao, H.D., Castrillon, J., Schmauss, C., Slättman, M., Gullberg, M. & Javitch, J.A. 2011. Detection of antigen interactions ex vivo by proximity ligation assay: endogenous dopamine D2-adenosine A2A receptor complexes in the striatum. *Biotechniques*, 51(2):111-118. Truban, D., Hou, X., Caulfield, T.R., Fiesel, F.C. & Springer, W. 2017. PINK1, Parkin, and mitochondrial quality control: what can we learn about Parkinson's disease pathobiology? *Journal of Parkinson's disease*, 7(1):13-29.

Tsutsui S, Schnermann J, Noorbakhsh F, Henry S, Yong VW, Winston BW & al., e. 2004. A1 adenosine receptor upregulation and activation attenuates neuroinflammation and demyelination in a model of multiple sclerosis. *Journal of neurosciences*, 24(6):1521–1529.

van Boxtel, M.P.J., Schmitt, J.A.J., Bosma, H. & Jolles, J. 2003. The effects of habitual caffeine use on cognitive change: a longitudinal perspective. *Pharmacology Biochemisty and Behavior*, 75(4):921-927.

van Calker D, M.M., Hamprecht B 1979. Adenosine regulates via two different types of receptors, the accumulation of cyclic AMP in cultured brain cells. *Journal of neurochemistry*, 33:999–1005.

Van der Walt, M.M., Terre'Blanche, G., Petzer, A., Lourens, A.C.U., Petzer, J.P. 2013. The adenosine A2A antagonistic properties of selected C8-substituted xanthines. *Bioorganic Chemistry*, 49:49-58.

Van Der Walt, M.M. & Terre'Blanche, G. 2015. 1,3,7-triethyl-substituted xanthines - possess nanomolar affinity for the adenosine A1 receptor. *Bioorganic & medicinal chemistry*, 23:6641-6649.

Van der Walt, M.M. & Terre'Blanche, G. 2018. Benzopyrone represents a privilege scaffold to identify novel adenosine A1/A2A receptor antagonists. *Bioorganic chemistry*, 77:136-143.

Van der Walt, M.M. & Terre'Blanche, G. 2015. 1, 3, 7-Triethyl-substituted xanthines—possess nanomolar affinity for the adenosine A1 receptor. *Bioorganic & medicinal chemistry*, 23(20):6641-6649.

Van Rensburg, H.D.J., Legoabe, L.J. & Terre'Blanche, G. 2020. C3 amino-substituted chalcone derivative with selective adenosine r A 1 receptor affinity in the micromolar range. *Chemical Papers*, 75:1581–1605

Van Rensburg, H.D., Terre'Blanche, G., Van der Walt, M. & Legoabe, L. 2017. 5-Substituted 2benzylidene-1-tetralone analogues as A1 and/or A2A antagonists for the potential treatment of neurological conditions. *Bioorganic Chemistry*, 74:251-259. Van Rensburg, H.D., Legoabe, L., Terre'Blanche, G. & Van der Walt, M. 2019b. 2– Benzylidene–1–Indanone Analogues as Dual Adenosine A1/A2a Receptor Antagonists for the Potential Treatment of Neurological Conditions. *Drug research*, 69(07):382-391.

Van Rensburg, H.D.J., Legoabe, L.J., Terre'Blanche, G. & Van der Walt, M.M. 2019a. Methoxy substituted 2-benzylidene-1-indanone derivatives as A 1 and/or A 2A AR antagonists for the potential treatment of neurological conditions. *MedChemComm*, 10(2):300-309

Vazquez-Rodriguez, S., Figueroa-Guíñez, R., Matos, M.J., Santana, L., Uriarte, E., Lapier, M., Maya, J.D. & Olea-Azar, C. 2013. Synthesis of coumarin–chalcone hybrids and evaluation of their antioxidant and trypanocidal properties. *MedChemComm*, 4(6):993-1000.

Vazquez-Rodriguez, S., Vilar, S., Kachler, S., Klotz, K.-N., Uriarte, E., Borges, F. & Matos, M.J. 2020. Adenosine receptor ligands: Coumarin–Chalcone hybrids as modulating agents on the activity of hARs. *Molecules*, 25(18):4306.

Van Bulck, M., Sierra-Magro, A., Alarcon-Gil, J., Perez-Castillo, A. & Morales-Garcia, J.A. 2019. Novel Approaches for the Treatment of Alzheimer's and Parkinson's Disease. *International journal of molecular science*, 20(3):719. doi: 10.3390/ijms20030719.

Veinbergs, I., Mante, M., Mallory, M. & Masliah, E. 2000. Neurotrophic effects of Cerebrolysin in animal models of excitotoxicity. *Journal of neural transmission. Supplement,* 59:273-280.

Weiner, M.W., Veitch, D.P., Aisen, P.S., Beckett, L.A., Cairns, N.J., Cedarbaum, J., Green, R.C., Harvey, D., Jack, C.R. & Jagust, W. 2015. 2014 Update of the Alzheimer's Disease Neuroimaging Initiative: a review of papers published since its inception. *Alsheimer's & dementia*, 11(6):e1-e120.

Wenk, G. 2006. Neuropathologic changes in Alzheimer's disease: Potential targets for treatment. *Journal of clinical psychiatry*, 67 Suppl 3:3-7.

Wilkinson, D. 2012. A review of the effects of memantine on clinical progression in Alzheimer's disease. *Interational journal of geriatric Psychiatry*, 27(8):769-776.

Wolf, E., Seppi, K., Katzenschlager, R., Hochschorner, G., Ransmayr, G., Schwingenschuh, P., Ott, E., Kloiber, I., Haubenberger, D. & Auff, E. 2010. Long-term antidyskinetic efficacy of amantadine in Parkinson's disease. *Movement disorders*, 25(10):1357-1363.

Wong, Y.C. & Krainc, D. 2017. α-synuclein toxicity in neurodegeneration: mechanism and therapeutic strategies. *Nature medicine*, 23:1-13.

Xie, A., Gao, J., Xu, L. & Meng, D. 2014. Shared mechanisms of neurodegeneration in Alzheimer's disease and Parkinson's disease. *Biomedical research international*, 2014:648740. doi: 10.1155/2014/648740.

Xu, K., Xu, Y.H., Chen, J.F. & Schwarzschild, M.A. 2002. Caffeine's neuroprotection against 1methyl-4-phenyl-1,2,3,6-tetrahydropyridine toxicity shows no tolerance to chronic caffeine administration in mice. *Neuroscience Letters*, 322:13–16.

Xu, K., Bastia, E., Schwarzschild, M. 2005. Therapeutic potential of adenosine A(2A) receptor antagonists in Parkinson's disease. *Pharmacology & therapeutics*, 105:267–310.

Xuesong, C., Othman, G. & Jonathan, D. 2010. Caffeine protects against disruptions of the BBB in animal models of Alzheimer's and Parkinson's disease. *Journal of Alzheimer's disease*, 20:127–146.

Xiong, H., Baskys, A. & Wojtowicz, J.M. 1996. Brain-derived peptides inhibit synaptic transmission via presynaptic GABAB receptors in CA1 area of rat hippocampal slices. *Brain research*, 737(1-2):188-194.

Xiong, H., Wojtowicz, J.M. & Baskys, A. 1995. Brain tissue hydrolysate acts on presynaptic adenosine receptors in the rat hippocampus. *Canadian journal of physiology and pharmacology*, 73(8):1194-1197.

Yadav, S., Gupta, S.P., Srivastava, G., Srivastava, P.K. & Singh, M.P. 2012. Role of secondary mediators in caffeine-mediated neuroprotection in maneb-and paraquat-induced Parkinson's disease phenotype in the mouse. *Neurochemical research*, 37(4):875-884.

Yamada, K., Kobayashi, M., Mori, A., Jenner, A. Kanda, T. 2013. Antidepressant-like activity of the adenosine A2A receptor antagonist, istradefylline (KW-6002), in the forced swim test and the tail suspension test in rodents. *Pharmacology, biochemistry & behaviour*, 114–115:23–30.

Yang, Z., Li, L., Zheng, J., Ma, H., Tian, S., Li, J., Zhang, H., Zhen, X. & Zhang, X. 2016. Identification of a new series of potent adenosine A2A receptor antagonists based on 4-amino-5-carbonitrile pyrimidine template for the treatment of Parkinson's disease. *ACS chemical neuroscience*, 7(11):1575-1584.

Yasuda, S., Sugiura, H., Tanaka, H., Takigami, S. & Yamagata, K. 2011. p38 Map kinase inhibitors as potential therapeutic drugs for neural diseases. *Central nervous system agents in medicinal chemistry*, 11:45–59.

Yu, L., Shen, H., Coelho, J., Araújo, I., Huang, Q., Day, Y., Rebola, N., Canas, P., Rapp, E., Ferrara, J., Taylor, D., Müller, C., Linden, J., Cunha, R., Chen, J. 2008. Adenosine A2A receptor antagonists exert motor and neuroprotective effects by distinct cellular mechanisms. *Annals of neurology*, 63:338–346.

Zheng, J., Zhang, X. & Zhen, X. 2018. Development of Adenosine A2A Receptor Antagonists for the Treatment of Parkinson's Disease: A Recent Update and Challenge. *ACS chemical neuroscience*, 10(2):783-791.

Zhou, S.J., Zhu, M.E., Shu, D., Du, X.P., Song, X.H., Wang, X.T., Zheng, R.Y., Cai, X.H., Chen, J.F. & He, J.C. 2009. Preferential enhancement of working memory in mice lacking adenosine A2A receptors. *Brain research*, 1303:74-83.

Zhuang, C., Zhang, W., Sheng, C., Zhang, W., Xing, C. & Miao, Z. 2017. Chalcone: a privileged structure in medicinal chemistry. *Chemical reviews*, 117(12):7762-7810.

## ANNEXURE



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North-West University Animal Care, Health and Safety Research Ethics Committee (NWU-AnimCareREC)

Tel: 018 299-1208 Email: <u>Ethics-AnimCare@nwu.ac.za</u> (for animal studies)

18 September 2019

## ETHICS APPROVAL LETTER OF STUDY

Based on approval by the North-West University Animal Care, Health and Safety Research Ethics Committee (NWU-AnimCareREC) on 18/09/2019, the NWU-AnimCareREC hereby approves your study as indicated below. This implies that the NWU-AnimCareREC grants its permission that, provided the general conditions specified below are met and pending any other authorisation that may be necessary, the study may be initiated, using the ethics number below.

Study title: Radioligand binding assays of selected A1/A2A adenosine antagonists using rat membrane cells and Chinese Hamster Ovary- cells expressing human adenosine receptors Principal Investigator/Study Supervisor/Researcher: Prof G Terre'Blanche Student: T Tutubala - 23368322																
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Approval of the study is provided for a year, after which continuation of the study is dependent on receipt and review of an annual monitoring report and the concomitant issuing of a letter of continuation. A monitoring report is due at the end of September annually until completion.																
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While this ethics approval is subject to signed in the application form, the fol-	llowin	g ge	nera	al teri	ms ar	d co	ndi	tions	s w	ill a <sub>l</sub>	oply					d
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- without any delay in case of any adverse event or incident (or any matter that interrupts sound ethical principles) during the course of the study.
- The approval applies strictly to the proposal as stipulated in the application form. Should any
  amendments to the proposal be deemed necessary during the course of the study, the principal
  investigator/study supervisor/researcher must apply for approval of these amendments at the NWUAnimCareREC, prior to implementation. Should there be any deviations from the study proposal

without the necessary approval of such amendments, the ethics approval is immediately and automatically forfeited.

- Annually a number of studies may be randomly selected for active monitoring.
- The date of approval indicates the first date that the study may be started.
- In the interest of ethical responsibility, the NWU-AnimCareREC reserves the right to:
- request access to any information or data at any time during the course or after completion of the study;
  - to ask further questions, seek additional information, require further modification or monitor the conduct of your research or the informed consent process;
  - withdraw or postpone approval if:
  - · any unethical principles or practices of the study are revealed or suspected;
  - it becomes apparent that any relevant information was withheld from the NWU-AnimCareREC or that information has been false or misrepresented;
  - submission of the annual monitoring report, the required amendments, or reporting of adverse events or incidents was not done in a timely manner and accurately; and/or
  - new institutional rules, national legislation or international conventions deem it necessary.
- NWU-AnimCareREC can be contacted for further information via <u>Ethics-AnimCare@nwu.ac.za</u> or 018 299 1208

NWU-AnimCareREC would like to remain at your service and wishes you well with your study. Please do not hesitate to contact the NWU-AnimCareREC for any further enquiries or requests for assistance.

Yours sincerely,

Digitally signed by Christiaan B Brink Date: 2019.09.18 16:34:28 +02'00'

Prof Tiaan Brink Chairperson NWU-AnimCareREC

Digitally signed by Prof Minrie Greeff Date: 2019.09.18 20:13:34 +02'00'

Prof Minrie Greeff Head of the Faculty of Health Sciences Ethics Office

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